# MESTECH

### Comparison of fluorogenic substrates for the detection of faecal indicator bacteria in water samples using a continuous fluorometric assay.

Ciprian Briciu-Burghina, Brendan Heery, Fiona Regan

Marine and Environmental Sensing Technologies Hub (MESTECH), National Centre for Sensor Research (NCSR), School of Chemical Sciences, Dublin City University, Dublin.

Ciprian.Briciu@dcu.ie

#### Background:

At present standard methods employed for the microbiological monitoring of bathing waters require at least 18 hours to perform and are based on culturing techniques. This is a huge drawback when immediate action is required. Real-time and on-line monitoring are key factors for consideration in current method development for continuous indicator organism detection in order to meet early warning requirements and water safety plans.

Methods utilising β-D-Glucuronidase (GUD) activity as an indicator of Escherichia Coli presence use labelled glucuronides to produce optical signals. Fluorometric assays for the measurement of Escherichia Coli GUD activity are traditionally performed using the fluorogenic substrate 4-methyl-umbelliferone-β-D-glucuronide (4-MUG) which upon hydrolysis releases the fluorophore 4-methyl-umbelliferone (4-MU). The major drawback of 4-MU is its high pKa (7.8), which causes only partial dissociation at pHs around the optimum pH for GUD activity (6.5-7.0). To overcome this issue researchers have employed discontinuous enzyme assays which require the addition of alkali.

In this context we explore the spectrophotometric properties of three fluorogenic substrates and their respective aglycons (Fig.1 ) for the continuous measurement of GUD activity and we apply the developed method for the rapid detection of Escherichia Coli in environmental water samples.

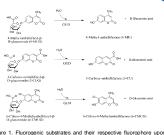
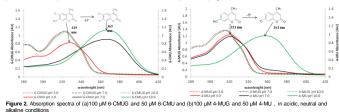


Figure 1. Fluoroge ca

#### **Results:**

► UV-VIS characterisation. UV-VIS spectroscopy was used to determine the absorption for the fluorophores and substrates at different pН values and the protonation/deprotonation behaviour of the fluorophores (Fig 2.)



#### Fluorescence spectroscopy characterisation.

When the excitation wavelength is selected to maximise the emission, the fluorescence intensity of 6-CMU in the 6.8-7.0 pH range is 6 times higher than that of 4-MU and 2.5 times higher than the fluorescence of 3-CU (Fig. 3).

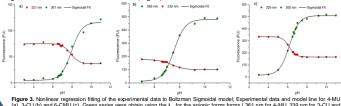


Figure 3. Nonlinear regression fitting of the e (a), 3-CU (b) and 6-CMU (c). Green series we ion fitting of the experimental data to Boltzman Sigmoidal model; Experimental data and model line for 4-MU ). Green series were obtain using the  $\lambda_{ex}$  for the anionic forms forms (361 nm for 4-MU, 339 nm for 3-CU and 5 nm for 6-CMU). Red series were obtained using the λ<sub>ex</sub> for the neutral forms (321 nm for 4-MU, 339 for 3-CU and 329 for 6-CMU).

10mL GUD extraction buffer (pH 6.8)

>Results

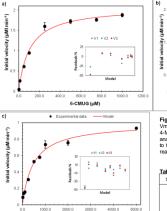
0532x+1.6537

1 mL 6-CMUG

1mM(pH 6.8)

Proof of concept: Rapid method for E.Coli detection

>GUD-Substrate kinetics. One way to investigate the interaction between GUD and the three substrates is through the use of Michaelis Menten parameters: K<sub>m</sub> and  $V_{\text{max}}$  A comparison between these parameters for the three substrates can give insights into the GUD's preferred molecule, catalysis rates and optimal substrate concentration. By conducting studies in the same conditions (pH, temperature, GUD concentration) the optimal substrate for GUD assay can be selected. Initial reaction velocities were plotted against substrate concentration (Fig 4 a,b,c).



3-CUG (µM

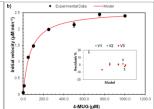


Figure 4. Michaelis-Menten models used to estimate Km and ax for the GUD catalysed hydrolysis of (a) 6-CMUG, (b) IUG and (c) 3-CUG. The inset shows the mean residual 4-MUG and (c) 3-CUG. The inset shows the mean residual analysis; V1, V2, V3 are the reaction velocities corresponding to the 3 runs. Reaction rates were followed for 10 min, with readings takes at each 5 s.

Substrate	K <sub>m</sub> (μM)	V <sub>max</sub> (µM min <sup>-1</sup> )	V <sub>max</sub> / K <sub>n</sub>
4-MUG	70.82	2.56	0.031
3-CUG	479.28	0.99	0.002
6-CMUG	106.88	2.07	0.019

#### Conclusions:

A continuous fluorometric method for the measurement of E.Coli GUD activity has been developed using 6-CMUG and offers a more straightforward approach for the evaluation of kinetic data. Benefits of this method as compared to a continuous one, include less sample manipulation, less reagent consumption, less experimental errors and better LOD.

The method was applied for the detection of E. Coli from environmental water samples and was successful in predicting E. Coli concentrations below the EU threshold for "excellent quality" in 1h.

## Acknowledgements:

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GUD activity

>Procedure

10-300 mL

ulture based meth

Figure 5. Linear regression between *E. Coli* concentrations determined using Colilert 18 and GUD activity from environmental water samples; (a) sea water samples, (b) fresh water samples; 31 individual water samples were used from which different dilutions were prepared and assayed for m which different dilutions.

GUD A









