

Effect of acute ingestion of exogenous ketone supplements on exercise metabolism, physical and cognitive performance in athletes

Mark Evans

Doctor of Philosophy



Dublin City University

Supervisor: Dr. Brendan Egan

Submitted to Dublin City University, April 2019

Declaration:

I hereby certify that this material, which I now submit for assessment on the programme of study leading to the award of PhD is entirely my own work, and that I have exercised reasonable care to ensure that the work is original, and does not to the best of my knowledge breach any law of copyright, and has not been taken from the work of others save and to the extent that such work has been cited and acknowledged within the text of my work.

Signed: _____ ID No: _____ Date: _____

Acknowledgements

Thank you to my family, especially my overly supportive parents, Therese and Philip Evans. Over the last four years you have got me through some difficult times and helped me celebrate the good ones. I may have gone overboard when you told me to stay in college for as long as possible. I won't ever be able to repay you for everything but I'll try my best. Thank you.

To Deirdre Duke, for all her love and support I am forever grateful. You've helped me to keep some perspective over the last four years and remember what's important in the grand scheme. I could not have done it without you. Thank you.

To some of my close friends; Neil Higgins, Stephen James, Steven Chapman, Gavin Mullan, Conor Quinn, Alan White, David McCrea, Greg Hawe, Peter Tierney, Mattias Fitzgerald, Matthew Coleman, Robert Fitzgerald, Robert Burke and Daniel Deasy. You'll never have to ask how long until I finish ever again and thank the lord for that.

Thank you to everyone in Dublin City University, University College Dublin and Waterford Institute of Technology who contributed to this work in one way or another. A PhD is never a solo effort and I'm grateful for all your help and hard work.

I would like to thank the Irish Research Council and Deepak Sharma of VITA Actives Ltd. for their support over the last four years.

Finally, to Dr. Brendan Egan, my BSc, MSc and PhD thesis supervisor and mentor. Thank you for guiding me through my academic career to date and going above and beyond your role every step of the way. I will always be grateful for the input, help and opportunity you have provided me over the last seven years.

Mark Evans

**Effect of acute ingestion of exogenous ketone supplements on exercise metabolism,
physical and cognitive performance in athletes**

Abstract

Ketone bodies, namely beta-hydroxybutyrate (β HB), acetoacetate (AcAc) and acetone, are produced in the liver during physiological states and manipulations that result in reduced carbohydrate availability. Exogenous ketone supplements, namely ketone esters and ketone salts, have been developed with the aim of achieving acute nutritional ketosis i.e. β HB concentrations > 0.5 mM. We investigated whether exogenous ketone supplements had an effect on the metabolic response to exercise, physical and cognitive performance in athletes.

A literature review was undertaken to examine the relationship between elevated ketone bodies, achieved via intravenous fusion of ketone bodies or fasting, and metabolism both at rest and during exercise¹. The review focused on seminal work performed in the 1970s and 1980s and identified a number of metabolic effects that may have relevance to improve performance and recovery in athletic populations.

Study 1 investigated the effect of a commercially-available ketone salt product formulation on the metabolic response to a graded exercise session in trained endurance athletes². We observed an elevation in β HB concentrations (0.4-0.5 mM), a reduction in plasma glucose concentrations, but no effect on plasma lactate concentrations or exercise efficiency.

Study 2 investigated the effect of co-ingestion with carbohydrate of a ketone ester supplement in the form of a (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone mono ester on

physical and cognitive performance in team sport athletes in response to a simulated soccer task³. Ingestion of the ketone ester had no effect on 15 m sprint times during the simulated task, or on reaction time or sustained attention performed afterwards. Compared to carbohydrate alone, we observed a preservation in executive function, measured by a decision making task, but a possible impairment in performance in a short high intensity intermittent performance test in the ketone ester condition.

Study 3 investigated the effect in trained runners of co-ingestion with carbohydrate of a (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone mono ester on the metabolic response to submaximal exercise, and performance in 10 km time trial and cognitive tasks. We observed no effect on endurance performance or cognitive performance with the ketone ester compared to the carbohydrate alone condition.

Future research should focus on exploring the optimal dosage and timing of exogenous ketone supplements around exercise to confer performance benefits, if any. It remains to be confirmed which exercise modalities may benefit from exogenous ketone supplementation.

References

1. **Evans M**, Cogan KE, Egan B (2017) Metabolism of ketone bodies during exercise and training: physiological basis for exogenous supplementation. *Journal of Physiology* 595(9):2857–71.
2. **Evans M**, Patchett E, Nally R, Kearns R, Larney M, Egan B (2018) Effect of acute ingestion of A-hydroxybutyrate salts on the response to graded exercise in trained cyclists. *European Journal of Sport Science* 18(3):376–86.
3. **Evans M**, Egan B (2018). Intermittent running and cognitive performance after ketone ester ingestion. *Medicine & Science in Sports & Exercise* 50(11):2330-2338.

Peer-reviewed Publications Arising from this Thesis

Literature review

Published in the Journal of Physiology in May 2017 (appendix A).

Evans M, Cogan KE, Egan B (2017) Metabolism of ketone bodies during exercise and training: physiological basis for exogenous supplementation. *Journal of Physiology* 595(9):2857–71.

Original research article

Published in the European Journal of Sport Science in April 2018 (appendix B).

Evans M, Patchett E, Nally R, Kearns R, Larney M, Egan B (2018) Effect of acute ingestion of β -hydroxybutyrate salts on the response to graded exercise in trained cyclists. *European Journal of Sport Science* 18(3):376–86.

Original research article

Published in Medicine and Science in Sports and Exercise in November 2018 (appendix C).

Evans M, Egan B (2018). Intermittent running and cognitive performance after ketone ester ingestion. *Medicine & Science in Sports & Exercise* 50(11):2330-2338.

Original research article

Published in Medicine and Science in Sports and Exercise in November 2019 (Appendix D)

Evans M, McSwiney FT, Brady AJ (2019). No benefit of ingestion of a ketone mono ester supplement on 10-km running performance. *Medicine & Science in Sports & Exercise* doi: 10.1249/MSS.0000000000002065

Conference Proceedings Arising from this Thesis

Oral presentation

European College of Sport Science annual meeting, Vienna 2016.

Ingestion of the ketone body beta-hydroxybutyrate alters the metabolic response to exercise in trained cyclists.

Poster presentation

Cell Symposia, Gothenburg 2017.

Ingestion of the ketone body beta-hydroxybutyrate alters the metabolic response to exercise in trained cyclists.

Oral presentation

European College of Sport Science annual meeting, Dublin 2018.

Intermittent running and cognitive performance after ketone ester ingestion.

Invited speaker

International Sport and Exercise Nutrition Conference, Newcastle 2018.

Exogenous ketone supplements: effects on exercise metabolism and performance.

Additional Peer-reviewed Publications Independent of this Thesis

Original research article

Published in the International Journal of Sport Nutrition and Exercise Metabolism in May 2018.

Evans M, Tierney P, Gray N, Hawe G, Macken M, Egan B (2018) Acute ingestion of caffeinated chewing gum improves repeated sprint performance of team sport athletes with low habitual caffeine consumption. *International Journal of Sport Nutrition and Exercise Metabolism* 28(3):221-227.

Original research article

Published in the European Journal of Applied Physiology in February 2018.

Cogan KE, **Evans M**, Iuliano E, Melvin A, Susta D, Neff K, De Vito G, Egan B (2018) Co-ingestion of protein or a protein hydrolysate with carbohydrate enhances anabolic signaling, but not glycogen resynthesis, following recovery from prolonged aerobic exercise in trained cyclists. *European Journal of Applied Physiology* 118(2):349-359.

Table of Contents

Declaration.....	i
Acknowledgements	ii
Abstract	iv
Peer-reviewed Publications Arising from this Thesis	vi
Conference Proceedings Arising from this Thesis	vii
Additional Peer-reviewed Publications Independent of this Thesis	viii
Table of Contents	ix
List of Figures	xii
List of Tables.....	xiii
List of Abbreviations.....	xiv
Chapter 1	1
Introduction	2
History and development of exogenous ketone supplements	3
Endogenous versus exogenous ketosis for athletic performance.....	7
Ketogenic supplements	9
Aims of the Thesis	12
Chapter 2	14
Introduction	15
Exercise metabolism and biochemistry.....	17
Overview.....	17
Substrate utilisation during exercise.....	18
Ketone bodies in circulation	21
Ketogenesis.....	22
Ketolysis in extra-hepatic tissues.....	23
Measurement of β HB	24
Effect of aerobic exercise training on enzymes of ketogenesis and ketolysis	26
Ketone body metabolism during exercise	28
Ketone body metabolism during exercise under conditions of <i>exogenous</i> ketosis	31
Metabolic effects of ingesting ketone salts and ketone esters at rest and during exercise	32
Ketone salts at rest.....	32
Ketone salts during exercise	33
Ketone esters at rest	34
Ketone esters during exercise	36
Ketone body metabolism after exercise: post-exercise ketosis	37
Modulation of post-exercise ketosis by aerobic exercise training and nutrition intervention	38
Metabolic consequences of post-exercise ketosis during recovery: a role for exogenous ketones as a recovery aid?.....	39
Effects beyond fuelling: β HB as a HDAC inhibitor	45
Effects on physical performance	47
Ketone esters	47
Ketone salts	48
Effects on cognitive performance.....	49
State-of-the-art in exogenous ketone supplementation and exercise performance	53
Chapter 3	54
Abstract	55

Introduction	56
Methods	57
Participants	57
Experimental design	58
Incremental exercise tests	59
Pre-trial preparation	60
Experimental trials	60
Blood analysis.....	62
Data analysis	62
Statistical analysis.....	63
Results.....	64
Discussion.....	69
Chapter 4	75
Abstract	76
Introduction	77
Methods	79
Participants	79
Experimental design	80
Incremental exercise test and familiarisation.....	81
Cognitive test battery	81
Pre-trial preparation	82
Experimental trials.....	83
Blood analysis.....	85
Statistical analysis.....	85
Results.....	86
Pre-exercise hydration status.....	86
Plasma β HB, glucose and lactate concentrations	86
15 m sprint times and shuttle run time to exhaustion	89
Cognitive performance.....	90
Gastrointestinal symptoms	91
Identification of KE and best performance trials	91
Discussion.....	93
Chapter 5	100
Abstract	101
Introduction	102
Methods	104
Participants	104
Experimental design	104
Assessment of $\dot{V}O_{2peak}$ and submaximal running speeds.....	105
Cognitive test battery	106
Pre-trial preparation	107
Main experimental trials.....	107
Blood analysis.....	111
Statistical analysis.....	111
Results.....	112
Plasma β HB, glucose and lactate concentrations	112
Submaximal exercise	114
10-km TT performance	117
Cognitive performance.....	117
Gastrointestinal symptoms	120

Identification of CHO+KME and best performance trials	120
Discussion.....	120
Chapter 6	127
Main research findings	128
Study 1	128
Study 2	128
Study 3	129
Metabolic response to ketone salt ingestion during exercise.....	129
Metabolic response to ketone ester ingestion during exercise.....	130
Effect of acute ingestion of a β HB ketone monoester on physical performance.....	131
Cognitive performance after exercise after ingestion of a β HB ketone monoester	134
Emerging issues and future directions for research in exogenous ketone supplementation in athletic performance	139
<i>βHB measurement</i>	140
<i>Recovery</i>	141
<i>Exogenous ketone dose</i>	142
<i>Substrate oxidation</i>	142
<i>Product racemity</i>	143
<i>Storage of samples</i>	143
Concluding remarks	144
References	145
Appendix A: Published Journal of Physiology article	163
Appendix B: Published European Journal of Sport Science article	178
Appendix C: Published Medicine and Science in Sport and Exercise article	189
Appendix D: Accepted and published ahead of print Medicine and Science in Sport and Exercise article.....	198

List of Figures

Figure 1. 1 Chemical structures of ketogenic supplements	11
Figure 2.1 Overview of ketogenesis, ketolysis and the metabolic and ergogenic potential of exogenous ketones.....	15
Figure 2. 2 The effect of exercise intensity and exercise duration on substrate metabolism during acute exercise	20
Figure 2. 3 Energy provision in skeletal muscle during exercise	20
Figure 2.4 Changes in β HB concentrations under various physiological states.	22
Figure 2.5 Metabolic pathways of ketone body metabolism in liver and skeletal muscle.....	25
Figure 2.6 β HB as a metabolic regulator and signalling metabolite.....	45
Figure 3.1 Experimental design schematic	62
Figure 3.2 Plasma β HB (a), glucose (b) and lactate (c), respiratory exchange ratio (d), and HR (e) responses after β HB salt ingestion and water ingestion.	66
Figure 4.1 Schematic of the study protocol. CHO, carbohydrate-electrolyte solution; PLA, placebo; RTE, shuttle run to exhaustion	85
Figure 4.2 Plasma β HB (A), glucose (B), and lactate (C) responses to ketone ester and placebo ingestion.	88
Figure 4.3 HR (A) and RPE (B) responses to ketone ester and placebo ingestion.....	89
Figure 4.4 Fifteen-meter sprint times (A) and shuttle run time to exhaustion (B) during ketone ester and placebo trials.....	90
Figure 5.1 Schematic of the study protocol.	110
Figure 5.2 Plasma β HB (A), glucose (B), and lactate (C) concentrations during ketone monoester and placebo trials.....	114
Figure 5.3 10-km time-trial performance (A), individual differences between CHO+KME compared to CHO+PLA (B), and running speeds for each 2 km split during the 10-km time-trials (C) in response to ketone monoester and placebo ingestion	118

List of Tables

Table 3.1 Participant anthropometrics and fitness profile	58
Table 3.2 Cardiorespiratory responses during graded exercise in CON or KET.....	67
Table 4.1 Cognitive performance measures assessed before and after each trial	92
Table 5.1 Physiological responses to 1 h of treadmill running at ~65% $\dot{V}O_{2\max}$ with carbohydrate co-ingested with either placebo or a ketone monoester	115
Table 5.2 Cognitive performance in response to ketone monoester and placebo ingestion	119
Table 6. 1 Summary of the effects of ketogenic supplements on physical and cognitive performance.....	136

List of Abbreviations

3-hydroxybutyrase dehydrogenase	BDH
Acetoacetate	AcAc
Acetoacetyl coenzyme-A	AcAc-CoA
Acetyl coenzyme-A acetyltransferase	ACAT
Area under the curve	AUC
ATP-phosphocreatine system	PCr
Adenosine tri-phosphate	ATP
Acetyl coenzyme-A	Ac-CoA
Acetyl-CoA carboxylase	ACC
Beats per minute	bpm
Body mass index	BMI
beta-hydroxybutyrate	β HB
β HB salts condition	KET
Calorie	kcal
Calorie per kilogram	kcal.kg ⁻¹
Calorie per minute	kcal.min ⁻¹
Carbohydrate	CHO
Carbon dioxide	CO ₂
Carbon dioxide production	$\dot{V}CO_2$
Tricarboxylic acid cycle	TCA
Delta efficiency	DE
Dry weight	DW
Dual-energy X-ray absorptiometry	DXA
D- β -hydroxybutyrate	D β HB
Effect size	ES
Free fatty acids	FFA
Gastrointestinal	GI
Gram	g
Gram per hour	g.h ⁻¹
Gram per kilogram body mass	g.kg ⁻¹ body mass
Gram per kilogram body mass per hour	g.kg ⁻¹ body mass.h ⁻¹
Gram per minute	g.min ⁻¹
Gross efficiency	GE
Heart rate	HR
High-intensity interval training	HIIT
Histone acetyltransferases	HAT
Histone deacetylase	HDAC
Hour	h
Hydroxymethylglutaryl-CoA synthase	HMGCS
Intramuscular triglyceride	IMTG
Joule	J
Ketone bodies	KB
Ketone ester condition	KE
Ketone monoester plus carbohydrate condition	CHO+KME
Ketone salts	KS

Kilogram	kg
Kilojoule	kJ
Kilojoule per minute	kJ.min ⁻¹
Kilometre	km
Kilometre per hour	km.h ⁻¹
L-βeta-hydroxybutyrate	LβHB
Loughborough intermittent shuttle test	LIST
Mammalian target of rapamycin	mTOR-1
Maximal oxygen consumption	$\dot{V}O_{2peak}$
Maximum heart rate	HR _{max}
Medium chain triglycerides	MCT
Meter	m
Milligram	mg
Milligram per kilogram body mass	mg.kg ⁻¹ body mass
Millilitre	mL
Millilitre per minute	mL.min ⁻¹
Millilitres per kilogram body mass per minute	mL.kg ⁻¹ body mass.min ⁻¹
Millimoles per litre	mM
Millisecond	ms
Minute	min
Monocarboxylate transporters	MCT
Multi tasking test	MTT
Oxygen consumption	$\dot{V}O_2$
Oxygen pulse	O ₂ pulse
Peak power output	W _{max}
Peroxisome proliferator-activated receptor gamma coactivator 1-alpha	PGC-1 α
Post-exercise ketosis	PEK
Placebo condition	PLA
Placebo plus carbohydrate condition	CHO+PLA
Pyruvate dehydrogenase	PDH
Rapid visual information processing task	RVP
Rating of perceived exertion	RPE
Reaction time task	RTI
Respiratory exchange ratio	RER
Respiratory minute volume	VE
Respiratory quotient	RQ
Revolutions per minute	RPM
<i>R,S</i> -1,3-butanediol acetoacetate diester	KDE
(<i>R</i>)-3-hydroxybutyl (<i>R</i>)-3-hydroxybutyrate ketone monoester	KME
Second	s
Solute ligand carrier	SLC
Solute ligand carrier protein 16A	SLC-16A
Standard deviation	SD
Succinyl-CoA:3-oxoacid CoA transferase	OXCT
Time trial	TT
Watt	W
Watts per second	W.s ⁻¹
Yoyo-intermittent recovery test level 1	YoYo-IR1

Chapter 1

Introduction

Ketone bodies (KB), namely β -hydroxybutyrate (β HB), acetoacetate (AcAc) and acetone, are produced in the liver during physiological states and nutritional manipulations that result in reduced carbohydrate availability, including fasting, starvation and ketogenic diets (Robinson & Williamson, 1980; Laffel, 1999). The practical relevance for athletic seeking performance gains generated from these states are negligible (Burke, 2015). This led to the exploration of exogenous ketone supplements as means to achieve acute nutritional ketosis to raise ketone body concentrations without reducing carbohydrate availability. The use of exogenous ketone supplements in professional sport was rumoured as long ago as 2012, while their use was more recently confirmed in professional cycling (Abraham, 2015; Pinckaers et al., 2016). Since then several original research articles have been published investigating the effect of various exogenous ketone supplements on the metabolic response to exercise and physical and cognitive performance (Cox et al. 2016; Leckey et al. 2017; O'Malley et al. 2017; Rodger et al. 2017; Waldman et al. 2017; Holdsworth et al. 2017; Vandoorne et al. 2017). While interest has peaked only in the last few years, the work on ketones provided exogenously, either through injections or infusions goes back to the 1940s where their potential to alter whole body and local tissue metabolism was first realised. These data remained largely isolated until recently. The purpose of this introductory chapter is to explore the historical development of exogenous ketone supplements, and the current interest in these supplements for athletic performance. Chapter 2 will provide an extensive review of the physiology and metabolism of ketone bodies at rest, during exercise and in response to training.

History and development of exogenous ketone supplements

During the 1940s, a series of experiments identified β HB and AcAc as two of sixteen metabolites with the ability to increase the metabolic efficiency of animal sperm (Lardy & Phillips, 1945; Lardy et al., 1945). Simultaneously, β HB and AcAc injections were administered to study their effects on glucose metabolism in animal models (Nath & Brahmachari, 1944; Nath & Brahmachari 1946; Nath & Brahmachari, 1948; Tidwell & Axelrod, 1948; Tidwell & Nagler, 1952; Chari & Wertheimer, 1953). The blood glucose response to KB administration was divergent in these works. In a set of experiments administering β HB and AcAc injections to rabbits and guinea pigs, progressive hyperglycaemia was observed and it was hypothesised that KB cause a hypersecretion of insulin and damage of the pancreatic islet cells (Nath & Brahmachari, 1944; Nath & Brahmachari, 1946; Nath & Brahmachari, 1948). Conversely, hypoglycaemia was observed in rats following AcAc administration and glucose tolerance was unaltered following simultaneous AcAc/glucose administration. A “sugar sparing action of ketone bodies” was suggested, associated with decreased glycogenolysis and lower blood glucose concentrations (Tidwell & Axelrod, 1948; Tidwell & Nagler, 1952). Furthermore, KB inhibit glycolysis and increase the conversion of glucose to glycogen as demonstrated in rat heart skeletal muscle in vitro (Maizels et al., 1977), perfused heart model in dogs (Laughlin et al., 1994), and reduce hepatic glucose output when combined with somatostatin in type 2 diabetic patients (Henry et al., 1990).

The describing of KB for exogenous delivery in the novel form of ketone esters occurred some 40 years ago, and outlined a speculated primary role in parenteral nutrition (Brunengraber, 1997). The use of KB, whether as esters of AcAc or β HB, as a source of parenteral nutrition was devised as an alternative to carbohydrate to deliver more energy at a lower osmolality, avoid the pathophysiological side effects that accompany intravenous

glucose infusion, and avoid the catabolism of protein that accompanies instances of critical illness and trauma in humans (Birkhahn, 1983). Initially, glycerol monobutyrate was used due to its structural similarity to β HB with no toxic effects being observed in rats during infusion (Birkhahn et al., 1977). Monoacetoacetin, a monoester of glycerol and AcAc was also a viable intravenous substrate for rats. Body weight gains were observed and nitrogen balance in growing mice was maintained with 71% of energy needs met with monoacetoacetin infused over 7 d compared to isocaloric carbohydrate infusion, with total KB concentrations elevated to 0.4-1.2 mM in various animal models (Birkhahn et al., 1979; Birkhahn, 1983). To further increase total KB concentrations and the amount of energy per osmol provided parenterally, triesters of glycerol and acetoacetate were developed but were not completely water-soluble and therefore unsuitable for intravenous infusion (Brunengraber, 1997). The solution to this was to use β HB, improving the water-solubility of the compounds. Glycerol β HB mono- and triesters maintained nitrogen balance and body weight similar to that of isocaloric carbohydrate infusion when providing 50% of energy needs in rats over 7 d (Birkhahn et al., 1997). As well as glycerol, 1,3-butanediol (BD) can be bound via an esterification reaction to β HB or AcAc. BD is converted to D- β HB in the liver via of alcohol and aldehyde dehydrogenase (Kies et al., 1973), elevating circulating β HB concentrations and having metabolic effects such as protein sparing in burned rats (Maiz et al., 1985).

These data on the metabolic action of ketone esters remained largely isolated until the last 5 years, but KB infusion studies in humans from the late 1960s to 1980s contribute significantly to knowledge on the role of KB during exercise (Hagenfeldt & Wahren, 1968; Hagenfeldt & Wahren, 1971; Fery et al., 1974; Sherwin et al., 1975; Balasse et al., 1978; Fery & Balasse, 1983; Fery & Balasse, 1986; Fery & Balasse, 1988). KB disposal into skeletal muscle is elevated as much as fivefold during exercise; reflected by a decrease in circulating KB concentrations at the onset of exercise. β HB is the primary KB extracted from

circulation and a net production of AcAc is observed. This drop in β HB is accompanied by an increase in the metabolic clearance rate (MCR) and oxidation of KB in skeletal muscle (Fery & Balasse, 1986). MCR is a measure of the ability of tissues to remove ketones from the blood, analogous to arteriovenous difference per unit time, but when measured during exercise is taken to represent an index of the ability of exercise to stimulate the capacity of working muscle to extract and utilise ketones (Fery & Balasse, 1983; Balasse & Fery, 1989). Taken together, the earlier work on KB and their potential role in parenteral nutrition informed the development and safety of ketone esters, whereas the infusion work in humans undertaking exercise informed the role that KB may play as an alternative fuel source for skeletal muscle (Chapter 2).

Methods of exogenous ketone supplementation producing acute nutritional ketosis

Investigating effects of ketosis on skeletal muscle metabolism has been typically achieved by endogenous ketosis using fasting of various durations (Balasse & Fery, 1989), or by exogenous ketosis produced by either ketone salt ingestion (Johnson & Walton, 1972), or infusion of AcAc or β HB (Fery & Balasse, 1988; Mikkelsen et al., 2015). Endogenous ketosis may also be achieved by carbohydrate (CHO) restriction, particularly by ketogenic diets (Paoli et al., 2013). The practical relevance for athletes seeking performance gains of metabolic responses generated from prolonged fasting is negligible, whereas benefits of ketogenic dieting for performance with a high intensity component are equivocal (Burke, 2015). This has led to the exploration of exogenous ketone ingestion as a means to achieve acute nutritional ketosis. Importantly, because endogenous ketosis results in concomitant elevations in free fatty acids (FFA) and alterations in glucose, insulin and counter-regulatory hormones, isolating the metabolic effects specific to KB has proved challenging. Therefore,

exogenous ketone supplementation is a means to address these questions and explore potential for performance and therapeutic benefits.

Oral administration of KB in their free acid form is expensive and ineffective at producing ketosis, so buffering the free acid form with sodium/potassium/calcium salts has been explored and are commercially available. These too are relatively ineffective at increasing β HB concentrations, but may be improved by co-ingestion with medium chain triglycerides (C:8, C:10), at least in rats (Kesi et al., 2016). However, ingestion of large quantities of ketone salts (KS) is impractical due to resulting gastrointestinal distress, and potentially undesirable consequences of cation overload or acidosis (Veech, 2004).

The development of ketone esters provides an alternative method to increase β HB concentrations, and are well-tolerated in rodents and humans (Clarke et al., 2012; Cox et al., 2016; Kesi et al., 2016). Two prominent ketone esters in the published literature are the R,S-1,3-butanediol acetoacetate diester (KDE) (Kesi et al., 2016; Leckey et al., 2017), originally developed by Dr. Henri Brunengraber (Case Western Reserve University) as a metabolic therapy for parenteral and enteral nutrition (Desrochers et al., 1995), and the (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester (KME) (Clarke et al., 2012; Cox et al., 2016), originally developed by Dr. Richard Veech (National Institutes for Health) and Dr. Kieran Clarke (University of Oxford) to improve the physical and cognitive performance in warfighters (Ford and Glymour, 2014).

Acute ingestion of either ester can result in short-term (~0.5 to 6 h) nutritional ketosis indicated by β HB concentrations >0.5 mM (Clarke et al., 2012; Leckey et al., 2017). For KME, ingestion at a dose 573 mg.kg^{-1} body mass (BM) resulted in β HB concentrations of ~ 3.0 mM after 10 minutes and rising to ~ 6.0 mM 30 min after ingestion (Cox et al., 2016). Ingestion of two 250 mg.kg^{-1} body mass KDE at 50 and 30 min prior to a 31.2 km cycling time trial, respectively, elevates serum β HB concentrations to ~ 0.4 mM. Whole blood β HB

concentrations reached ~0.6 mM during the time trial and peaked at ~1.4 mM during a 1 h recovery period (Leckey et al., 2017). Acute nutritional ketosis is therefore achieved without the impracticality of prolonged fasting or ketogenic dieting.

Endogenous versus exogenous ketosis for athletic performance

The link between increasing muscle glycogen stores and enhanced exercise capacity is well-established, accomplished by increasing intake of dietary carbohydrates in the days prior to exercise or through exogenous carbohydrate feeding during exercise (Bergström et al., 1967; Coyle et al., 1986; Jeukendrup, 2004). Accordingly, sports nutrition guidelines recommend high carbohydrate intakes in preparation for and during competition to maximise muscle glycogen stores and improve performance (Burke et al., 2004). Despite this, low carbohydrate, high fat ketogenic diets are recommended in some quarters as a means to take advantage of the body's large fat reserves, namely adipose tissue and intramuscular triglycerides (IMTG), as a source of fuel during exercise (Volek et al., 2015). A period of “keto-adaptation” is often cited as necessary gain full benefit of a high fat diet (LCHF) for athletic performance. LCHF (<50g carbohydrate.day⁻¹) is associated with a preservation of submaximal exercise capacity (~60% $\dot{V}O_{2peak}$), a reduction in RER reflecting an increasing in fat utilisation during exercise and an elevation of circulating β HB to ~0.5-2.5 mM, depending on the length of the LCHF (Phinney et al., 1980; Phinney et al., 1983; Volek et al., 2016; McSwiney et al., 2018). Furthermore, muscle glycogen stores are maintained during 20 months of LCHF (~70% fat) despite elevated β HB concentrations, possibly explaining the preservation of exercise capacity and the ability to synthesise muscle glycogen from gluconeogenic substrates (Volek et al., 2016). These dietary approaches can be considered as “endogenous ketosis”, but are proposed to impair high intensity exercise performance through an attenuation of pyruvate dehydrogenase activity (PDH), a key glycolytic enzyme

(Burke, 2015). This performance impairment was demonstrated as a reduction in 1 km sprint power output following 6 d adaptation to a LCHF (~68% fat), while 100 km time trial performance was maintained (Stellingwerff et al., 2006; Havemann et al., 2006). Of note is the brief (<1 wk) adaptation period to the LCHF, which proponents of the diet say is too short to fully adapt to endogenous ketosis as an alternative fuelling strategy (Volek et al., 2015). However, while 3 wk adaptation to a LCHF improves maximal aerobic capacity ($\dot{V}O_{2\text{peak}}$) in a group of Australian elite race walkers, it conversely impairs gains in performance in response to a 3 wk intense training camp in elite race walkers (Burke et al., 2017).

“Exogenous ketosis”, also known as the aforementioned acute nutritional ketosis, can be produced through ingestion of a ketone ester or ketone salt (Clarke et al., 2012; Cox et al., 2016; Stubbs et al., 2017) but does not require a LCHF to elevate β HB concentrations, and results in a similar increase in β HB concentrations to endogenous ketosis. Elevations in β HB concentrations are achieved with concomitant exogenous ketone supplement and carbohydrate intake, regardless of prior feeding status (Stubbs et al., 2017; Myette-Côté et al., 2018). Exogenous ketone ingestion aims to circumvent the impairment of high intensity exercise performance in athletes by elevating β HB concentrations without the impractical but necessary low carbohydrate intake implemented during endogenous ketosis. However, similarly to endogenous ketosis, rats supplemented for 5 d with exogenous ketones exhibit a reduction in cardiac PDH activity versus rats fed a high carbohydrate diet (Murray et al., 2016), but this inhibition remains to be directly confirmed either in human skeletal muscle, or in response to a single bolus ingestion. Nevertheless, given the effects of acute ingestion of KME to reduce glycolytic flux during exercise (Cox et al., 2016), it is likely that some of these effects are being mediated via inhibition of PDH.

Ketogenic supplements

Ketones esters and KS are exogenous ketone supplements orally ingested with the aim of elevating circulating β HB concentrations. β HB has two enantiomers, designated D- and L-, or R- and S-, respectively. Currently-available commercial β HB assays and handheld point-of-care monitors determine the concentration of the D- enantiomer. D- β HB is the circulating and primary form of β HB (Tsai et al., 2006), and L- β HB is not a substrate for mitochondrial 3-hydroxybutyrate dehydrogenase, a key enzyme in the ketolytic pathway, and thus is not metabolised to AcAc (Schofield et al., 1982). The constituent parts of ketone esters and KS are different and this alters the metabolic response to the respective ketogenic supplements after acute ingestion. Ketone esters, namely KME and KDE, are D- β HB and AcAc molecules respectively, attached via an ester bond either to another ketone body or a ketone body precursor such as BD. KS are a racemic mixture of D/L- β HB or non-racemic D- β HB molecules attached to a sodium/calcium/potassium mineral salt (Figure 1.1). All investigations in to the efficacy of elevating β HB concentrations using KS have used commercially-available supplements containing a racemic mixture of β HB, typically containing 50% of the D- and L-enantiomers (Stubbs et al., 2017; O'Malley et al., 2017; Rodger et al., 2017; Fischer et al., 2018; Waldman et al., 2018). KME has a chiralic purity of 99%, meaning it exclusively contains the D- β HB isomer, whereas commercially-available KS have a purity of 50%, making them less effective at elevating the metabolically-active D- β HB enantiomer (Stubbs et al., 2017). One serving of KME (DeltaG, HVMN Ketone, HVMN, United States) provides 25g D- β HB, whereas one serving of KS (KetoCana, KetoSports, United States) provides 11.7 g β HB comprising of 50% each of the D- and L- β HB enantiomers.

In addition to the ingestion of KB-containing formulations, medium chain

triglycerides are ketogenic agents and may be coingested with KS to augment the β HB response to KS ingestion (Kesi et al., 2016). Briefly, medium chain triglycerides are fatty acids 6 to 12 carbons in length and are transported to the liver directly via the portal vein and oxidised rapidly (Bach & Babayan, 1982). Acute ingestion of 25 g medium chain triglyceride prior to exercise elevates β HB to ~ 0.5 mM, lowering blood glucose concentrations but has no effect on glycogen utilisation during 60 min cycling at 60% $\dot{V}O_{2\text{peak}}$ (Decombaz et al., 1983). Simultaneous carbohydrate and medium chain triglyceride ingestion increases the rate of medium chain triglyceride oxidation compared to medium chain triglyceride ingestion alone (Jeukendrup et al., 1995), with the latter impairing performance in a cycling time trial due to a high incidence of gastrointestinal distress (Jeukendrup et al., 1998). However, this impairment in performance was not observed with concomitant carbohydrate and medium chain triglyceride ingestion (Jeukendrup et al., 1998).

The original article on the effect of a (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester (KME) on cycling time trial performance remains the *only* report of a performance benefit of an exogenous ketone supplement (Cox et al., 2016). Ingestion of 573 mg.kg⁻¹ body mass of KME improved 30 min maximal distance time trial performance by $\sim 2\%$, whereas 31.2 km cycling time trial performance was impaired by $\sim 2\%$ with ingestion of the R,S-1,3-butanediol acetoacetate diester (Leckey et al., 2017). Racemic ketone salts have no performance benefit for short duration, high intensity efforts (O'Malley et al., 2017; Rodger et al., 2017; Waldman et al., 2018). At the time of commencing our experimental work, the *Cell Metabolism* report (Cox et al., 2016) was the only published work in the area and informed the dosing and timing strategies employed in studies 1, 2 and 3 of this thesis.

Given the increasing interest but relative lack of scientific research on exogenous ketone supplements, there is a need to investigate whether ketogenic supplements alter the

metabolic response to various exercise challenges and improve athletic performance. With these gaps in knowledge in mind, three human intervention studies were undertaken to investigate whether administration of β HB in the form of KS or ketone esters altered the metabolic response to exercise and/or improved physical and cognitive performance in response to various exercise challenges in trained athletes.

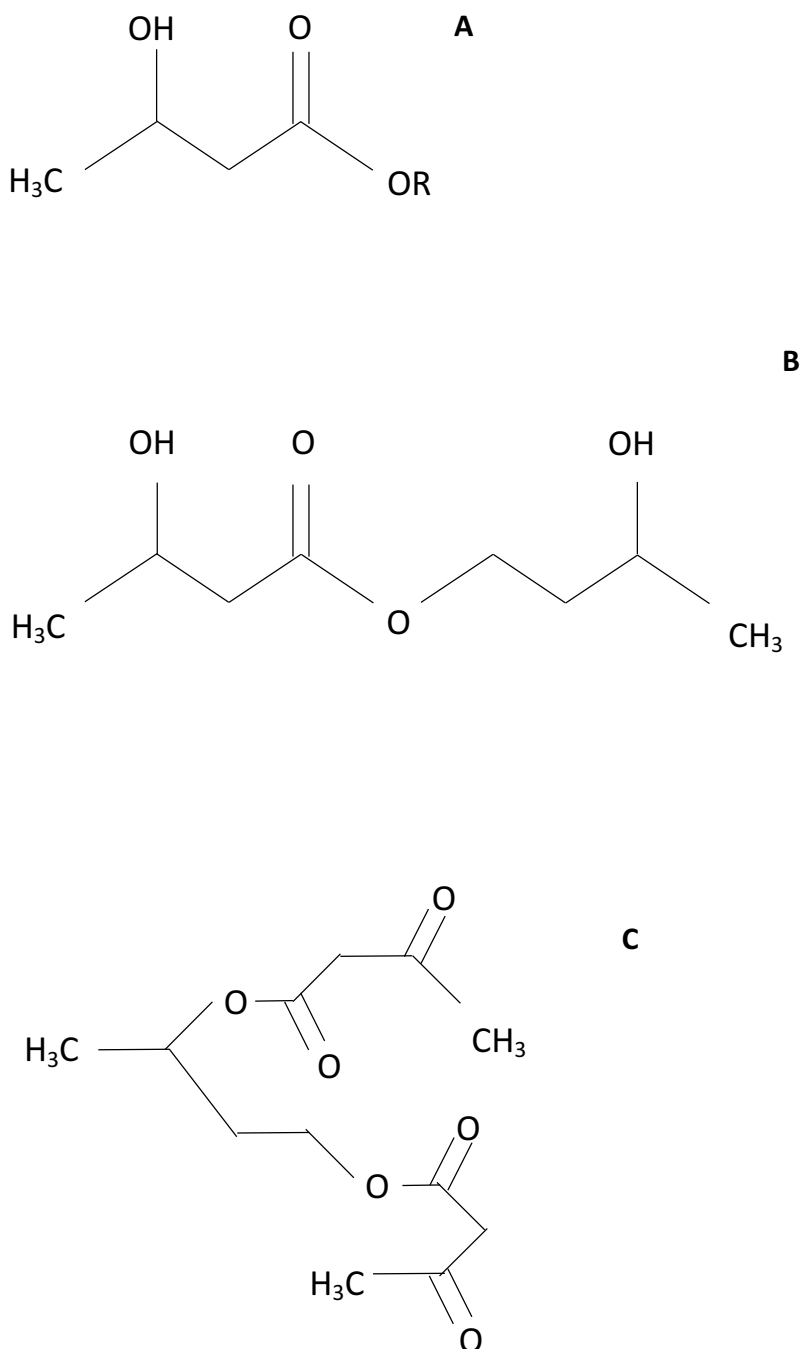


Figure 1. 1 Chemical structures of ketogenic supplements

A, racemic ketone salts. B, the (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester. C, R,S-1,3-butanediol acetoacetate diester.

Aims of the Thesis

- During pilot testing, discover the optimal dose and timing of racemic ketone ingestion to maximise peak β HB concentrations after acute ingestion of racemic ketone salts.
- Use this dose and timing information to investigate the effect of racemic ketone salts on the metabolic response to a graded exercise session in trained cyclists. Specifically, are the β HB concentrations achieved similar to those of the (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester and will they cause the characteristic reduction in plasma glucose and attenuate the exercise induced rise in plasma lactate concentrations.
- Investigate whether carbohydrate plus the (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester has an ergogenic effect on short duration, high intensity exercise performance during a 75 min soccer simulation protocol and a subsequent intermittent run to volitional exhaustion in team sport athletes compared to carbohydrate ingestion alone. It is of interest to whether the metabolic effects of (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester ingestion, namely a reduction in plasma glucose and an attenuation in the exercise-induced rise in plasma lactate, are observed in this intermittent sporting context.
- Investigate whether ingestion of carbohydrate plus the (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester has an ergogenic effect on 10 km self-paced, treadmill based time trial performance following a 1 h pre load at 65% $\dot{V}O_{2\text{peak}}$ compared to carbohydrate alone.
- Investigate whether ingestion of carbohydrate plus the (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester improves cognitive performance in intermittent

and endurance contexts compared to carbohydrate ingestion alone. Reaction time, decision-making and sustained attention will be measured.

Chapter 2

Sections from this literature review are included in the published Journal of Physiology article (Appendix A): **Evans M**, Cogan KE, Egan B (2017) Metabolism of ketone bodies during exercise and training: physiological basis for exogenous supplementation. *Journal of Physiology* 595(9):2857–71.

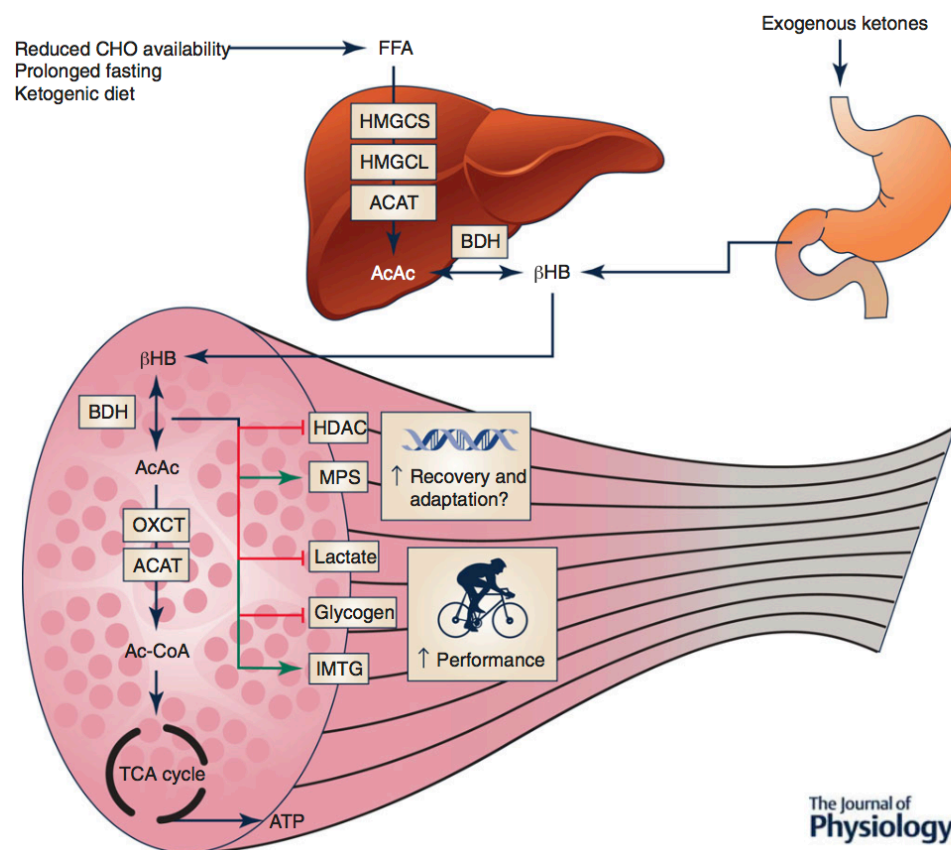


Figure 2.1 Overview of ketogenesis, ketolysis and the metabolic and ergogenic potential of exogenous ketones.

Endogenous ketogenesis and exogenous ketone ingestion result in an increase in circulating β HB. β HB enters the muscle where it alters the metabolic response to exercise and may result in improved recovery and adaptation from exercise training or improved physical performance.

Introduction

Over the past century, exercise physiologists have appreciated the role of carbohydrate and fat in energy provision to exercising skeletal muscle. Much of the work examining the metabolic response during exercise and the impact of exercise on metabolic regulation and adaptive responses to training has focussed on the relative contribution of

these fuels (Egan & Zierath, 2013). Optimising training and nutrition strategies by manipulating the relative intakes of these macronutrients is central to supporting elite sports performance (Cermak & van Loon, 2013; Bartlett et al., 2015; Burke, 2015). An alternative fuel source to CHO and fat are ketone bodies (KB), namely acetoacetate (AcAc), acetone, and β -hydroxybutyrate (β HB), which are produced in the liver during physiological states and nutritional manipulations that result in reduced CHO availability, most commonly during prolonged fasting, starvation, and ketogenic [very low CHO (~5%), low protein (~15%), high fat (~80%)] diets (Robinson & Williamson, 1980; Laffel, 1999). This relative glucose deprivation and concomitant elevation in circulating free-fatty acids results in the production of KB to replace glucose as the primary fuel for peripheral tissues such as the brain, heart and skeletal muscle in these states.

Aside from a role as an alternative fuel source, KB exert a range of metabolic effects including attenuating glucose utilisation in peripheral tissues, anti-lipolytic effects on adipose tissue, and potential attenuation of proteolysis in skeletal muscle (Figure 2.1) (Robinson & Williamson, 1980). KB are utilised by working muscle during exercise (Fery & Balasse, 1986; Fery & Balasse, 1988), and the capacity to uptake and oxidise KB during exercise is higher in exercise-trained skeletal muscle (Winder et al., 1975). Despite these observations, in addition to a glucose sparing action (Maizels et al., 1977) and potential to lower the exercise-induced rise in plasma lactate concentrations (Fery & Balasse, 1988), the potential performance benefits of KB when provided as an exogenous fuel source had until recently received little attention, but had been postulated (Cox & Clarke, 2014; Pinckaers et al., 2016). Since then several articles have been published investigating the effect of various exogenous ketone supplements on the metabolic response to exercise and physical and cognitive performance (Cox et al. 2016; Leckey et al. 2017; O'Malley et al. 2017; Rodger et al. 2017; Holdsworth et al. 2017; Vandoorne et al. 2017; Waldman et al., 2018). Apart from a role as

an alternative fuel source, KB may act as signalling molecules to regulate gene expression and adaptive responses (Shimazu et al., 2013; Zou et al., 2016). Moreover, therapeutic roles for KB have long been proposed in a variety of disease states including aberrant glucose metabolism, genetic myopathies, hypoxic states and neurodegenerative pathologies (Veech, 2004). For therapeutic effects, exogenous ketones are ingested in the form of β HB salts or ketone esters to produce acute (~0.5 to 6 h) nutritional ketosis (Clarke et al., 2012; Kesl et al., 2016), but a surge in interest in KB as a performance aid for athletes arose when ketone ester supplementation was confirmed in professional cycling (Abraham, 2015; Pinckaers et al., 2016).

Exercise metabolism and biochemistry

Overview

Skeletal muscle has the capacity to alter the rate of energy production in response to movement, powered by actin-myosin crossbridge cycling contraction (Podolsky and Schoenberg, 1983). Exercise causes skeletal muscle to rapidly increase its rate of ATP hydrolysis, providing an immediate source of energy for muscular work by increasing ATP turnover more than 100-fold compared to resting values (Gaitanos et al., 1993). Metabolic pathways that drive ATP production are activated during exercise that help to maintain ATP stores, facilitating the continuation of muscular work. Skeletal muscle is the primary site of carbohydrate and lipid metabolism for energy production, illustrated by a greater than 30-fold demand increase in intramuscular oxygen consumption during strenuous exercise and an approximate 70- to 100- fold increase in tricarboxylic acid cycle flux under similar conditions (Anderson and Saltin, 1985; Gibala et al., 1998). The relative contribution of CHO and lipid to total energy provision during exercise is dependent on the modality, the frequency, intensity, and duration of individual exercise sessions (Egan and Zierath, 2013).

Substrate utilisation during exercise

The relative contribution of different metabolic pathways to energy provision is dependent on the relative intensity and power output of the exercise bout. Power output determines the ATP demand while intensity determines the relative contributions of CHO, lipid and other fuel sources. Isotope tracer methodology has provided us with a detailed view of the impact of these factors on substrate utilisation during exercise (Romijn et al., 1993; van Loon et al., 2001). At low to moderate intensities of exercise ($<65\% \dot{V}O_{2\text{peak}}$), the primary source of fuel for skeletal muscle is glucose, derived from hepatic glycogenolysis, and free fatty acids, liberated from adipose tissue. These responses are dependent on various hormonal responses, including adrenaline, noradrenaline, insulin, and glucagon (Kjaer, 2006). As exercise intensity increases ($>65\% \dot{V}O_{2\text{peak}}$), the contribution of CHO increases while the use of FFA declines. This trend continues up to maximal intensities, where the vast majority of energy is being provided by muscle glycogen (Van Loon et al., 2001). Conversely to exercise intensity, as exercise duration increases ($> 60 \text{ min}$) lipid oxidation increases (Romijn et al., 1993) (Figure 2.2). Muscle glycogen, the predominant fuel during moderate to intense exercise, is converted to glucose in a process known as glycogenolysis. This process is under the control of the enzyme glycogen phosphorylase, regulated by allosteric modulation by AMP and IMP (Hargreaves, 2006). The end product of glycogenolysis is glucose molecules that are converted to glucose-6-phosphate and subsequently pyruvate in the cytoplasm before crossing the mitochondrial membrane, converted to acetyl Co-A and incorporated into the tricarboxylic acid cycle to produce ATP. As the duration of exercise increases, or the intensity of exercise decreases, the contribution of lipids to fuel provision increases. FFA, liberated from adipose tissue and intramuscular triglycerides are the two sources of lipids that muscle can oxidise during exercise. Lipids, in the form of triglycerides, are stored in adipose tissue and sequentially broken down to glycerol and three fatty acids by

the enzymes adipose triglyceride lipase, hormone sensitive lipase and monoacylglycerol lipase under the control of the hormone insulin and the catecholamines. Each fatty acid passes out of the adipocyte in to the blood and are transported across the plasma membrane of the muscle cell via fatty acid transports, including fatty acid binding protein, fatty acid transport protein and fatty acid translocase. Fatty acids are transported across the carnitine shuttle and undergo β -oxidation in the mitochondria in the mitochondria to produce acetyl CoA, which enters the tricarboxylic acid cycle to produce ATP and fuel muscular work (Figure 2.3).

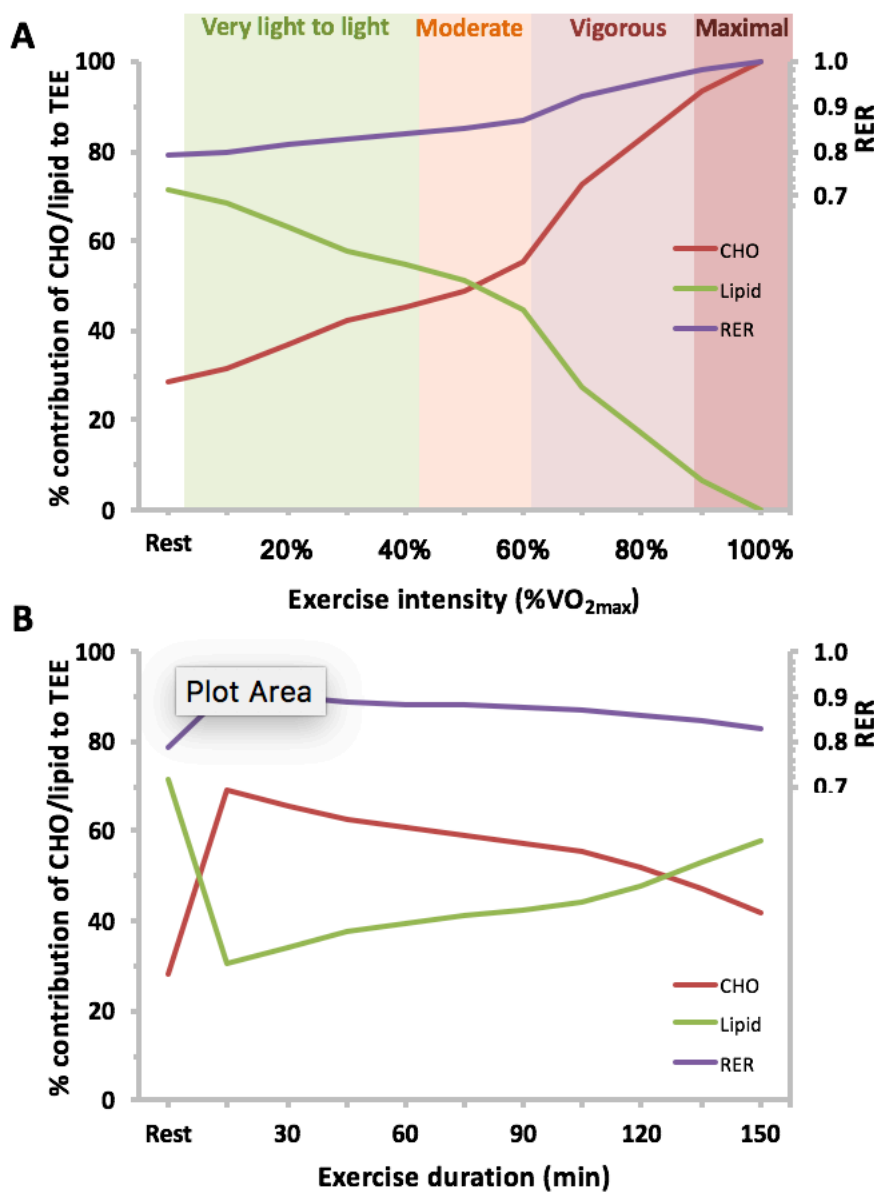


Figure 2. 2 The effect of exercise intensity and exercise duration on substrate metabolism during acute exercise

(A) Substrate contribution to exercise of increasing intensity. Up to 30%VO₂max, oxidation of lipid sources (mostly plasma FFAs) accounts for the majority of energy provision. As exercise intensity increases, absolute CHO oxidation rate and relative contribution to energy provision increases. The lipid oxidation rate increases up to 60%–70% VO₂max, after which it declines as intensity increases. In relative terms, the contribution of lipid oxidation to energy provision decreases proportionally with increasing exercise intensity, as reflected by a steady rise in RER. (B) Substrate contribution to exercise at a fixed intensity (e.g., 65%VO₂max) for an extended duration. An initial rise in RER occurs at the onset of exercise, reflecting the increase in relative contribution of CHO to energy provision compared to resting metabolism. Thereafter, a small but steady decline in RER is observed with extended duration of exercise, reflecting the declining relative contribution of CHO to energy provision as a function of the increasing relative contribution of lipid.

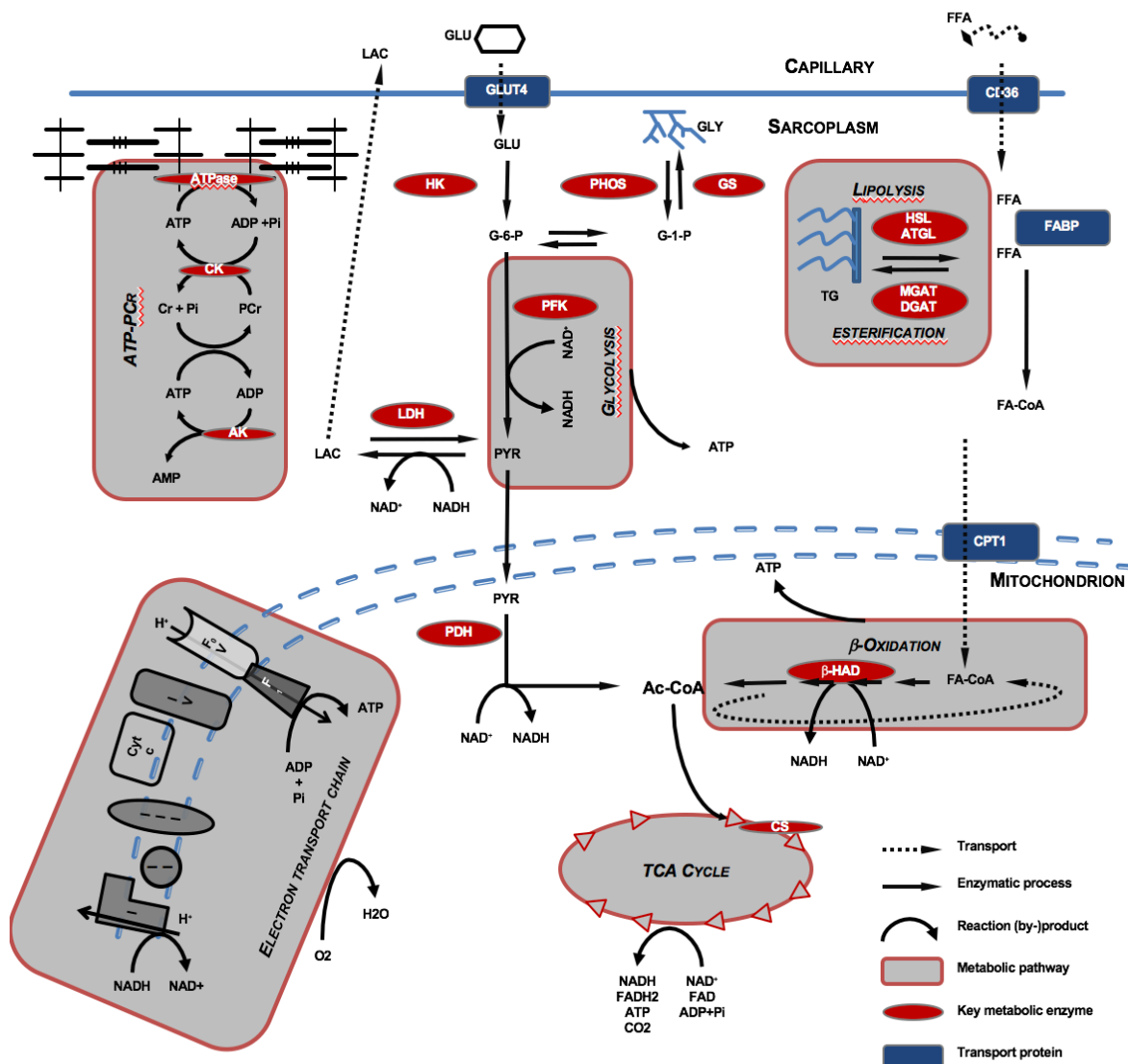


Figure 2. 3 Energy provision in skeletal muscle during exercise

ATP hydrolysis, catalyzed by myosin ATPase, powers skeletal muscle contraction. Metabolic pathways of ATP generation in skeletal muscle include (1) the ATP-phosphagen system wherein the degradation of PCr by creatine kinase (CK) produces free Cr and Pi, which is transferred to ADP to re-form ATP; the adenylate kinase (AK) (myokinase) reaction catalyzes the formation of ATP and AMP from two ADP molecules; (2) anaerobic glycolysis, where glucose-6-phosphate derived from muscle glycogen (GLY) (catalyzed by glycogen phosphorylase, PHOS) or circulating blood glucose (GLU) (catalyzed by hexokinase, HK), is catabolized

pyruvate (PYR), which is reduced to lactate (LAC) by lactate dehydrogenase (LDH), and produces ATP by substrate level phosphorylation; (3) processes of carbohydrate (glycolysis) and lipid (β-oxidation) metabolism producing acetyl-CoA (Ac-CoA), which enters the tricarboxylic acid (TCA) cycle in the mitochondria, coupled to oxidative phosphorylation in the electron transport chain (ETC). The two main metabolic pathways, i.e., glycolysis and oxidative phosphorylation, are linked by the enzyme complex pyruvate dehydrogenase (PDH). GLUT4 facilitates glucose uptake to the sarcoplasm, which may undergo glycolysis or during rest/inactivity, be stored as glycogen via glycogen synthase (GS). Fatty acyl translocase (FAT/CD36) facilitates long-chain fatty acid transport at the sarcolemma, and, in concert with fatty acid binding protein (FABPpm) and carnitine palmitoyltransferase 1 (CPT1), across the mitochondrial membrane. FFAs entering the cell may be oxidized via β-oxidation or be diverted for storage as IMTG via esterification by monoacylglycerol acyltransferase (MGAT) and diacylglycerol acyltransferase (DGAT). Liberation of FFAs from IMTG stores via lipolysis in skeletal muscle during exercise occurs via the activities of HSL and ATGL. All pathways of ATP generation are active during exercise, but the relative contribution of each is determined by the intensity and duration of contraction, as a function of the relative power (rate of ATP production) and capacity (potential amount of ATP produced). CS, citrate synthase; Cyt c, cytochrome c; PFK, phosphofructokinase

Overview of ketone body metabolism

Ketone bodies in circulation

Plasma KB concentrations reflect the balance between hepatic production (“ketogenesis”) and peripheral breakdown and utilisation (“ketolysis”) in extra-hepatic tissues, both of which are under various levels of control as detailed in previous reviews (Robinson & Williamson, 1980; Laffel, 1999). Ketogenesis is an evolutionarily-conserved adaptive response playing a critical role in survival during an energy crisis by providing a substrate for the brain, which cannot utilise FFA as a fuel source. AcAc, acetone, and βHB comprise the KB, although βHB is not technically a ketone because the ketone moiety has been reduced to a hydroxyl group. AcAc and βHB are short-chain, four carbon organic acids that act as FFA-derived circulating substrates to provide energy to extra-hepatic tissues, whereas the contribution of acetone, readily generated by the spontaneous decarboxylation of AcAc, to energy provision is negligible. Plasma KB concentrations are <0.1 mM in the postprandial state, whereas hyperketonemia is accepted as KB concentrations exceeding 0.2 mM (Robinson & Williamson, 1980). Various states of CHO restriction, depletion and dysregulation produce hyperketonemia to different degrees (Figure 2.4).

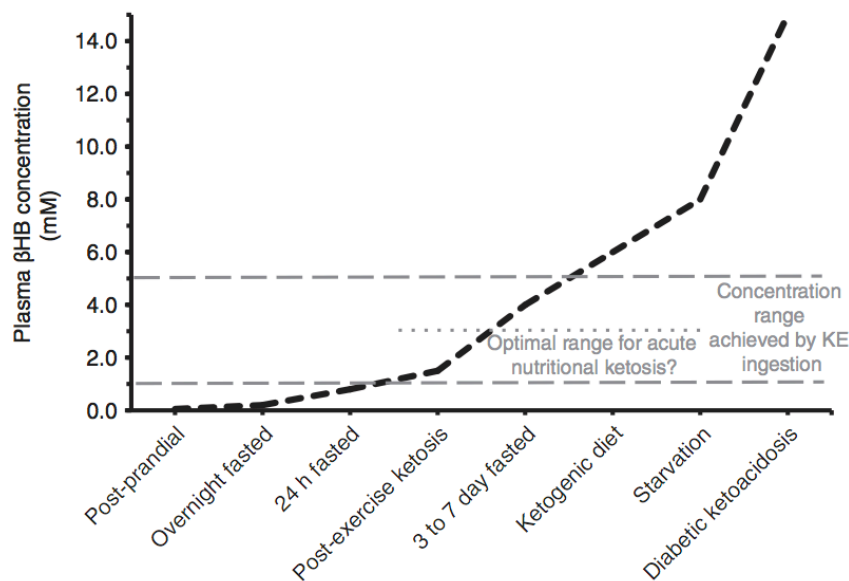


Figure 2.4 Changes in β HB concentrations under various physiological states.

Plasma [KB] is <0.1 mM in the postprandial state when consuming high CHO or high protein meals, and rises upward after an overnight fast, and with ketogenic dieting, prolonged fasting, starvation, and pathological states of ketoacidosis.

Ketogenesis

The primary substrate for ketogenesis are FFA liberated from adipose tissue. Ketogenic amino acids, namely leucine, lysine, phenylalanine, isoleucine, tryptophan, and tyrosine also serve ketogenesis, but are likely to contribute to less than 5% of circulating KB (Thomas et al., 1982). The rise in FFA is consequent to the stimulation of lipolysis as a result of declines in plasma glucose and insulin that are characteristic of reduced CHO availability. Factors stimulating ketogenesis include an elevated glucagon-to-insulin ratio and decline in hepatic glycogen concentration, while reduced blood flow to the liver or elevations in [KB] suppress ketogenesis (Robinson & Williamson, 1980; Laffel, 1999). Ketogenesis involves a series of sequential reactions beginning with acetyl CoA (Ac-CoA) and acetoacetyl CoA (AcAc-CoA), and ending with the liberation of AcAc (Figure 2.5). Some AcAc is exported, but the majority is reduced to β HB in an NAD^+/NADH -coupled near equilibrium reaction catalysed by 3-hydroxybutyrate dehydrogenase (BDH), in which the equilibrium constant

favours β HB formation. These KB are transported into the circulation via the solute ligand carrier (SLC) protein 16A (SLC16A) family of monocarboxylate transporters (MCTs) in mitochondrial and sarcolemmal membranes.

Ketolysis in extra-hepatic tissues

In peripheral tissues, KB, primarily in the form of β HB, enter the mitochondrial matrix again via MCT1-mediated transport. β HB is re-oxidised to AcAc via BDH after which sequential reactions result in the generation of two molecules of Ac-CoA (Figure 2.5). These are incorporated into the TCA cycle via citrate synthase for terminal oxidation and production of ATP, which in skeletal muscle contributes to fuelling muscular work (Fery & Balasse, 1986, 1988). Succinyl-CoA:3-oxoacid CoA transferase (OXCT) is essential for ketolysis in extra-hepatic tissues, with very low abundance in hepatocytes explaining the lack of ketolytic activity in these cells (Robinson & Williamson, 1980).

Activity of OXCT is highest in heart and kidney, followed by skeletal muscle and the brain (Robinson & Williamson, 1980), but because skeletal muscle accounts for ~40% of body mass in adult humans, this organ accounts for the highest fraction of total KB metabolism at rest (Balasse & Fery, 1989; Laffel, 1999). Beginning almost 50 years ago, models using various durations of fasting, and combined with primed constant infusion of radiolabelled either AcAc or β HB tracers and arteriovenous difference measures to quantify KB turnover, established that skeletal muscle is a major site of ketolysis at rest (Hagenfeldt & Wahren, 1968; Owen & Reichard, 1971; Wahren et al., 1984; Elia et al., 1990; Mikkelsen et al., 2015). Skeletal muscle has a high affinity to KB, but because of low circulating concentrations under normal conditions, the contribution to energy provision in muscle is less than 5%, and FFA are the main source of energy provision in the post-absorptive state. The relationship between ketone oxidation and KB concentrations is curvilinear such that

contribution to energy provision in skeletal muscle rises to ~10% after an overnight fast (Hagenfeldt & Wahren, 1968; Owen & Reichard, 1971), 20% to 50% after 72 h of fasting (Owen & Reichard, 1971; Elia et al., 1990), but declines to ~15% after 24 days of starvation (Owen & Reichard, 1971). Thus, skeletal muscle demonstrates saturation kinetics for the KB concentration-oxidation relationship, with saturation likely between 1 and 2 mM as demonstrated by fasting of various durations (compiled in (Balasse & Fery, 1989)) or step-wise β HB infusion (Mikkelsen et al., 2015).

Measurement of β HB

The majority of studies on acute exogenous ketone ingestion use handheld point-of-care meters with appropriate reagent strips to measure whole blood β HB concentrations (Precision Neo handheld monitor, Freestyle Optium handheld monitor, Precision Xtra handheld monitor Abbott Laboratories, Witney, UK; Glucomen Lx plus-meter, Menarini Diagnostics, Firenze, Italy) (O'Malley et al., 2017; Rodger et al., 2017; Stubbs et al., 2017; Myette-Côté et al., 2018; Stubbs et al., 2018; Waldman et al., 2018; Leckey et al., 2017). Ketone body reagent strips use a β HB dehydrogenase enzyme-based amperometric strip to establish whole blood β HB (Ceriotti et al., 2014). The use of handheld monitors is preferable to urinary ketone measurement due to the inability of nitroprusside, present in the urinary sticks, to react with β HB, the predominant ketone body in circulation (Guimont et al., 2015). Other studies have used laboratory methods, including reagent/colorimetric kits for measurement of plasma β HB (Randox, Daytona; Sigma Aldrich; ABX Pentra, France) (Cox et al., 2016; Leckey et al., 2017; Vandoorne et al., 2017). The Optium Freestyle handheld monitor (Abbott Laboratories, Berkshire, UK) overestimates β HB concentrations but falls within the coefficient of variation (10%) with a laboratory method (Guimont et al., 2015), but this overestimation increases with higher β HB concentrations (>5.0mM) (Ceriotti et al.,

2014). Confounding factors for β HB measurement with handheld monitors include high haematocrit and ascorbic acid concentrations, both of which cause overestimation of β HB with the Freestyle Optium monitors (Ceriotti et al., 2014; Dhatariya, 2016). Furthermore, β HB concentrations were overestimated by ~ 0.4 - 0.5 mM during exercise using a handheld monitors compared to direct assay-based laboratory methods (Leckey et al., 2017). These reports suggest elevations in β HB concentrations via exogenous ketone ingestion must be critically evaluated based on the measurement method and any confounding factors that could affect such methods.

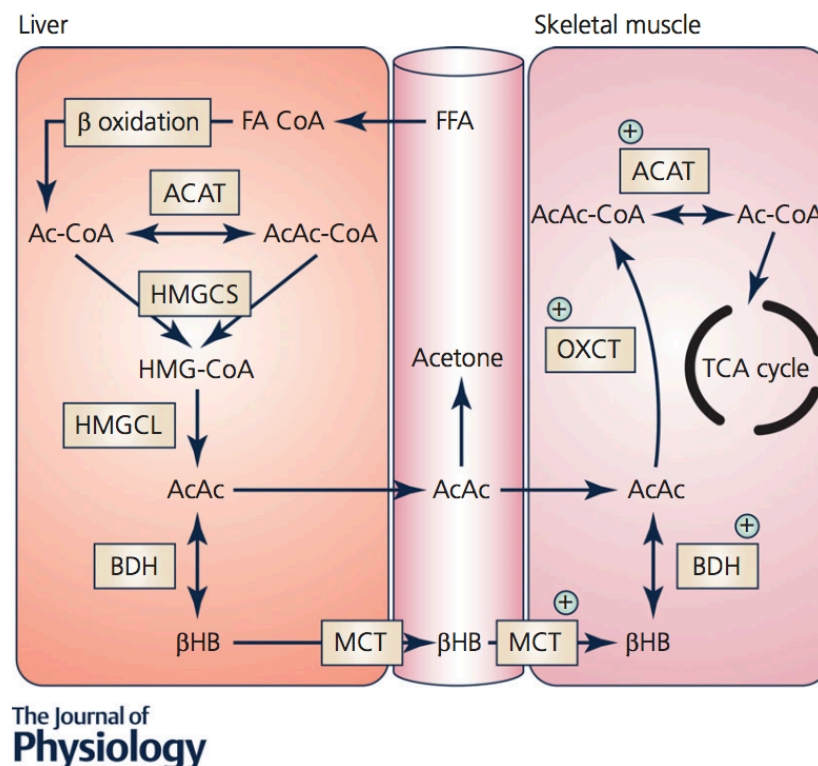


Figure 2.5 Metabolic pathways of ketone body metabolism in liver and skeletal muscle

During ketogenesis, FFA are converted to fatty acyl CoA, enter hepatic mitochondria via MCT transport and undergo β -oxidation to acetyl CoA. Sequential reactions lead to the reduction of the central ketone body, AcAc, to β HB. In the muscle β HB is reoxidised to AcAc. Two molecules of acetyl-CoA are formed during the process of ketolysis

Effect of aerobic exercise training on enzymes of ketogenesis and ketolysis

Adaptations to exercise training reduce perturbations to homeostasis during subsequent bouts of exercise, and thereby enhance resistance to fatigue. Central to these effects are enhanced respiratory capacity and contractile parameters, and importantly adaptations that contribute towards maximising delivery and utilisation of circulating substrates (Egan & Zierath, 2013). Therefore, if KB make a meaningful contribution to energy provision during exercise, it is pertinent to explore analogous regulation in skeletal muscle. Training-induced changes in expression and activities of enzymes of ketolysis in skeletal muscle have not been described in humans, but differences in KB metabolism during and after exercise between trained and untrained individuals have been reported (Johnson et al., 1969; Johnson & Walton, 1972; Rennie et al., 1974; Rennie & Johnson, 1974). The general pattern is for attenuation in trained individuals of the post-exercise rise in KB concentrations, but this is influenced by nutritional manipulation and relative exercise intensity, the latter of which has often been poorly controlled (see later sections).

Nevertheless, circulating concentrations reflect the balance between ketogenesis and ketolysis, these differences may be explained by the factors influencing one or both. For ketogenesis, data are limited but suggest that in exercise-trained rodents enzymatic activity of BDH or ACAT (Winder et al., 1974), or HMGCS (Askew et al., 1975) is unaltered in liver, and, in fact, the overall activity of the ketogenic pathway may be lower (El Midaoui et al., 2006) compared to untrained rodents. In these rodent models of intense aerobic exercise training, the activities of the ketolytic enzymes BDH, OXCT and ACAT are higher in trained skeletal muscle (Winder et al., 1974; Askew et al., 1975; Winder et al., 1975; Beattie & Winder, 1984). This coincides with two- to three-fold higher ex vivo rates of β HB and AcAc oxidation in gastrocnemius muscle homogenates presented with concentrations of both β HB and AcAc at 0.1 and 0.5 mM (Winder et al., 1973; Winder et al., 1975).

In terms of muscle fibre type, enzymatic activities of BDH, OXCT and ACAT are all highest in type I fibres, intermediate in type IIA fibres, and lowest in type IIB fibres of rats (Winder et al., 1974). BDH is essentially undetectable in type IIB muscle fibres, and across the fibre types BDH activity is much lower than activities of OXCT and ACAT (Winder et al., 1974). Although OXCT is essential for ketolysis, BDH activity is, therefore, potentially rate-limiting in skeletal muscle. When rats performed 12 weeks of treadmill running, compared to sedentary rats BDH activity was almost three-fold higher in type I fibres, but six-fold higher in type IIA fibres of trained skeletal muscle, resulting in concentrations comparable to the type I fibres (Winder et al., 1974). OXCT activity was 26% higher in type I, and approximately two-fold higher in type IIA and IIB fibres, whereas ACAT activity was 40% to 45% higher in all three fibre types in trained skeletal muscle (Winder et al., 1974). Similarly, in skeletal muscle from mice with 8 weeks of access to running wheels, the difference compared to sedentary mice was greater for BDH mRNA expression (~two-fold higher than sedentary) compared to differences in OXCT and ACAT mRNA expression (~30% to 50% higher) (Svensson et al., 2016). These changes in ketolytic enzymes are localised to the working muscle given the absence of change after training in the heart (Askew et al., 1975), kidney and brain (Winder et al., 1974).

In terms of KB transport into skeletal muscle, similarly to the ketolytic enzymes, MCT1 protein expression is highest in type I fibres, lowly expressed in type II fibres, and correlates well with muscle oxidative capacity (Bonen, 2001). Elevated MCT1 protein expression after exercise training is well-established for human skeletal muscle, and increases occur in an intensity-dependent manner (Thomas et al., 2012). Using a rodent perfused hindlimb model, the capacity for uptake of KB in skeletal muscle at 1 mM each of β HB and AcAc was higher in an aerobically-trained group of rats, with uptake of total KB, AcAc and β HB 33%, 27% and 53% higher, respectively, compared to untrained rats (Ohmori et al.,

1990). Similarly, β HB clearance during a β HB tolerance test is higher in mice given 8 weeks of running wheel access, or with enhanced oxidative capacity consequent to skeletal muscle overexpression of PGC-1 α , a transcriptional co-activator and master regulator of mitochondrial biogenesis in adaptive responses such as exercise training (Svensson et al., 2016). In both conditions, this coincides with elevated expression of MCT1 and the ketolytic enzymes in skeletal muscle. Therefore, the uptake and utilisation of KB in skeletal muscle is likely to be greatest in those individuals that are highly trained with a high proportion of type I muscle fibres and a high oxidative capacity in skeletal muscle.

Ketone body metabolism during exercise

The existing literature on fuel selection during exercise has focused almost exclusively on utilisation of CHO and fat, but skeletal muscle has the ability to resynthesize ATP from other substrates including protein, lactate, and KB (Fery & Balasse, 1986; Mazzeo et al., 1986; Fery & Balasse, 1988; Wagenmakers et al., 1991). With increasing exercise intensity, the contribution of substrates to energy provisions shifts from blood-borne FFA and glucose to increased reliance on intramuscular fuel stores, namely intramuscular triglyceride and muscle glycogen, such that at moderate to high intensities ($>75\%VO_{peak}$) of exercise, muscle glycogen is the main source of energy provision (van Loon et al., 2001). This pattern is readily altered by nutritional manipulation such as CHO loading and acute CHO ingestion resulting in increased CHO utilisation (Bosch et al., 1996), glycogen depletion resulting in increased contribution of protein to energy provision (Wagenmakers et al., 1991), and habitual high fat consumption resulting in increased contribution of fat to energy provision (Volek et al., 2016). Clearly, skeletal muscle is a major site of ketolysis under fasting conditions, but central to the rationale for exogenous ketone supplementation must be that

ketolysis increases during exercise, makes a meaningful contribution to energy provision, and can alter patterns of substrate utilisation.

The pioneering work of Hagenfeldt, Wahren and colleagues (Hagenfeldt & Wahren, 1968; Hagenfeldt & Wahren, 1971; Wahren et al., 1984) and Fery, Balasse and colleagues (Balasse et al., 1978; Fery & Balasse, 1983; Fery & Balasse, 1986; Fery & Balasse, 1988) established that KB disposal into human skeletal muscle is elevated as much as five-fold during exercise. This is generally reflected by a drop in KB concentrations soon after the onset of exercise, primarily β HB, concomitant with increases in KB oxidation in skeletal muscle and elevated metabolic clearance rate (MCR). MCR is a measure of the ability of tissues to remove ketones from the blood, analogous to arteriovenous difference per unit time, but measured during exercise is taken to represent an index of the ability of exercise to stimulate the capacity of working muscles to extract and utilise ketones (Fery & Balasse, 1983; Balasse & Fery, 1989). Because the stoichiometry of KB oxidation yields respiratory quotients of 1.00 and 0.89 for AcAc and β HB, respectively (Frayn, 1983), calculation of oxidation rates for KB from whole-body gas exchange data has not been routinely performed using methods that determine the relative contribution of CHO and fat oxidation. However, a recent attempt has been made (Cox et al., 2016) based on methods and assumptions described for KB utilisation during ketogenesis (Frayn, 1983). Previous to this, oxidation rates for KB have historically been derived from arteriovenous differences of radiolabelled KB across working muscles with rates calculated as a fraction of O_2 consumption or CO_2 production (Hagenfeldt & Wahren, 1968; Balasse et al., 1978).

Like CHO and fat utilisation, KB metabolism during exercise is influenced by a variety of factors including metabolic status (Wahren et al., 1984; Fery & Balasse, 1986), training status (Johnson & Walton, 1972; Rennie et al., 1974; Beattie & Winder, 1985), and the intensity of exercise (Cox et al., 2016). Given the aforementioned fibre type-specific

differences for activities of ketolytic enzymes, the muscle fibre type profile of the working muscle is also likely to be an important determinant of ketolysis during exercise. However, the most important determinant of KB metabolism during exercise is the degree of ketonemia, and the method by which this is achieved i.e. of endogenous or exogenous origin.

Ketone body metabolism during exercise under conditions of *endogenous* ketosis

Like KB metabolism in resting skeletal muscle, the relationship between concentration and oxidation or MCR is curvilinear (Balasse & Fery, 1989). At low ketonemia (<1.0 mM) such as that produced by an overnight fast, resting MCR is as much as four-fold greater than during prolonged fasting (Fery & Balasse, 1983). During prolonged exercise of low-to-moderate intensity after an overnight fast, MCR increases by 50% to 75% (Fery & Balasse, 1983; Fery & Balasse, 1986), which indicates that working muscle has an increased capacity to extract ketones from blood compared to rest. However, when ketonemia exceeds 2.5 mM such as that achieved by greater than 72 h of fasting, the exercise-induced rise in MCR is abolished (Fery & Balasse, 1986). Therefore, when ketosis is achieved by prolonged (>72 h) fasting there is a negligible contribution of KB oxidation to energy provision (Hagenfeldt & Wahren, 1971; Fery & Balasse, 1986), but after an overnight fast, the contribution ranges from 2 to 10% (Balasse et al., 1978; Fery & Balasse, 1983; Wahren et al., 1984). Under these conditions, the majority of energy provision in working muscle is from CHO and fat as classically described (van Loon et al., 2001). Moreover, unlike CHO and fat, there is progressive attenuation of the oxidation of KB with rising ketonemia, and thus the mobilisation of KB is not the factor limiting oxidation in skeletal muscle. This attenuation of exercise-stimulated MCR suggests either that above a threshold concentration the capacity for skeletal muscle to oxidise KB becomes saturated, and/or that hyperketonemia itself is a self-inhibitory factor (Balasse & Fery, 1989). Mechanistically, this is likely mediated either

through the inhibition of OXCT by elevated AcAc, and/or via FFA-mediated inhibition of ketolysis (Robinson & Williamson, 1980). This regulation is critical in the starvation response because the capacity of the liver to produce KB closely matches the requirements of the brain to utilise KB as an energy source (Robinson & Williamson, 1980). Therefore, excessive oxidation by working muscle would threaten survival, whereas its inhibition spares circulating substrate for the brain (Hagenfeldt & Wahren, 1971; Fery & Balasse, 1983).

Ketone body metabolism during exercise under conditions of *exogenous* ketosis

The aforementioned self-inhibitory effect of rising ketonemia underscores a key methodological issue when considering KB metabolism in skeletal muscle, namely the method of achieving ketosis. While fasting of various durations is a widely used model of ketosis, acute nutritional ketosis relevant to sports performance would be achieved with replete glycogen stores, and in the absence of prolonged elevations in FFA and KB concentrations that would likely impair KB oxidation rates through these mechanisms. To our knowledge, only two studies have addressed this convincingly by examining effects of exercise on KB metabolism without interference from the various hormonal and metabolic perturbations associated with prolonged fasting or diabetes (Fery & Balasse, 1988; Cox et al., 2016).

In the former study (Fery & Balasse, 1988), infusion of sodium AcAc after an overnight fast achieved KB concentrations of ~6 mM (β HB ~3.5 mM, AcAc ~2.5 mM) at the onset of 2 h of exercise at ~52% $\dot{V}O_{2\text{peak}}$. Notably, AcAc did not change during exercise whereas β HB declined throughout exercise to be reduced by ~2 mM at the end of exercise. This coincided with a progressive rise in MCR throughout exercise peaking at ~75% higher than rest at the end of exercise. In contrast, this effect was abolished with similar ketonemia in three to five day fasted participants. Importantly, although the inhibition of KB oxidation

by hyperketonemia is present during exogenous ketosis, an “auto-amplification” was noted that is not present in fasting ketosis i.e. the initial rise in MCR induced by exercise causes a reduction in concentration which, in turn, provokes a further rise in MCR and so on. Additionally, the threshold concentration at which hyperketonemia inhibits MCR was higher in exogenous ketosis than in fasting ketosis. However, in terms of contribution to energy provision, this ultimately only resulted in a 2% contribution over the 2 h exercise bout. Nevertheless, plasma lactate concentrations did not rise during exercise after AcAc infusion compared to a ~1 mM rise in the fasted participants, which suggests that despite a modest contribution to energy provision, exogenous ketosis can impact on metabolic processes during exercise.

Metabolic effects of ingesting ketone salts and ketone esters at rest and during exercise

Ketone salts at rest

Ingestion of 1.6 and 3.2 mM.kg⁻¹ body mass in the form of a sodium/potassium racemic KS elevates whole blood β HB concentrations to ~0.8 mM and 1.0 \pm 0.1 mM, respectively with concentrations peaking between 60-90 min (Stubbs et al., 2017). Similarly, ingestion of 0.5 g.kg⁻¹ body mass sodium/calcium KS, providing ~0.31 g.kg⁻¹ body mass β HB elevates plasma β HB concentrations to 0.60 \pm 0.30 mM after 2.5 h, after which concentrations returned to baseline over 3 h (Fischer et al., 2018). Elevations in β HB concentrations via KS ingestion coincide with decreases in plasma FFA, triglycerides and glucose, and an elevation in plasma insulin, which may be accounted for by the small amount of carbohydrate used to sweeten the KS (Stubbs et al., 2017). KS ingestion does not alter blood pH or plasma lactate, but reduces plasma potassium concentrations and increases sodium and chloride concentrations (Fischer et al., 2018). L- β HB concentrations are elevated

to ~2.0 mM with the ingestion of KS (Stubbs et al., 2017), suggesting racemic KS products are more effective at raising the L-enantiomer than the D-enantiomer of β HB.

Ketone salts during exercise

Acute ingestion of one serving KS elevates whole blood β HB concentrations between 0.5-0.8 mM (O'Malley et al., 2017; Rodger et al., 2017; Waldman et al., 2018). Given the lower dosage employed and the discrepancy in β HB measurement between handheld monitors and laboratory methods, it is a salient question whether these measurements in whole blood are accurate. Serving sizes have ranged from 11.38 g β HB to 23.4 g D/L- β HB. This modest elevation in β HB concentrations is accompanied by an ~10% decrease in plasma glucose during sub maximal exercise (O'Malley et al., 2017), however, this decrease is not always observed (Rodger et al., 2017; Waldman et al., 2018). During submaximal exercise, plasma lactate is not affected by acute KS ingestion (Rodger et al., 2017; O'Malley et al., 2017; Waldman et al., 2018). Ingestion of KS lowers RER during exercise (O'Malley et al., 2017) given the respiratory quotient of AcAc is 1.00, similar to that of CHO it would be expected to rise when compared to a non-caloric placebo, so more work is needed as to whether training status or a larger dose may have a further effect on RER during exercise. Studies using the manufacturers recommended dose of KS do not report incidences of gastrointestinal distress. However, given the modest elevation in β HB concentrations during exercise with these dosages, increasing the dose is required to achieve β HB >1.0 mM, which may cause gastrointestinal distress, possibly due to acute hyperosmotic load.

Increases in heart rate (HR) have been observed with β HB infusions (Gormsen et al., 2017; Svart et al., 2018) but this is not observed with KS ingestion (Waldman et al., 2018). It is likely the sodium load in KS is causing this elevation in HR as it is not observed with either acute KME or KDE supplementation. Acute sodium ingestion can elevate blood

pressure (Farquhar et al., 2005), and a similar sodium load delivered as sodium bicarbonate results in an elevation in HR of ~10 bpm during moderate intensity activity (Kahle et al., 2013).

Ketone esters at rest

In the fasted state, ingestion of 573 mg.kg⁻¹ body mass KME elevates plasma β HB concentrations to 3.5 ± 0.3 mM at 10 min, reaching ~6.0 and ~6.5 mM after 40 and 70 min, respectively (Cox et al., 2016). Ingestion of 482 mg.kg⁻¹ KME elevates blood β HB concentrations to ~3.2 mM after 30 min, thereafter concentrations decrease during a 2 h oral glucose tolerance test (Myette-Côté et al., 2018). Whole blood β HB concentrations peaked at 2.8 ± 0.2 mM 60 min following the ingestion of 282 mg.kg⁻¹ KME and at ~1.5 mM 20 min following ingestion of 141 mg.kg⁻¹ (Stubbs et al., 2017). These results demonstrate a dose response in β HB concentrations following KME ingestion, with elevating concentrations following increasing dosages in the fasted state. Feeding status alters the β HB response to KME ingestion, as a prior breakfast (600 kcal; macronutrient ratio carbohydrate: protein: fat of 2:1:1) attenuates the elevation in whole blood β HB by ~1.0 mM (Fed: ~2.0 mM; Fasted ~3.0 mM), reducing the β HB area under the curve by 27% following 395 mg.kg⁻¹ body mass KME (Stubbs et al., 2017). Conversely, ingestion of 500 mg.kg⁻¹ body mass KDE elevates serum β HB to ~0.4 mM 60 min after ingestion of the initial bolus (Leckey et al., 2017), a major difference between the two ketone esters. The AcAc response to KME and KDE ingestion are similar, with concentrations elevated to ~0.3 mM 60 min after KDE ingestion (Leckey et al., 2017) and ~0.7 mM after KME ingestion (Stubbs et al., 2017). The elevation in β HB concentrations with KME coincides with decreases in plasma glucose, FFA and triglycerides (Cox et al., 2016; Stubbs et al., 2017; Myette-Côté et al., 2018).

A secondary mechanism for the glucose lowering effect of ketone esters has been proposed (Myette-Côté et al., 2018); ingestion of KME resulted in a 44% reduction in FFA AUC over the course of a 2 h oral glucose tolerance test, with the authors suggesting the anti-lipolytic action of KB may be responsible for this increase insulin sensitivity. The insulintropic effects of KB in humans are evident (Balasse et al., 1970; Miles et al., 1981) but is frequently not observed (Balasse & Ooms, 1968; Balasse & Neef, 1975; Balasse, 1979; Fery & Balasse, 1980; Beylot et al., 1986; Nair et al., 1988; Mikkelsen et al., 2015); it is suggested that the insulintropic action of ketone bodies occur under conditions where the ketone body concentration is raised above 2.0 mM, and when this rise occurs abruptly (Balasse & Fery, 1989). Given the elevation in circulating β HB observed with KME ingestion, it may have an insulintropic effect. In the context of ketone ester ingestion, insulintropic is defined as an increase in circulating insulin concentrations with exogenous ketone supplement plus nutrient condition being greater than nutrient alone. In the fasted state, ingestion of KME causes a small elevation in insulin (Stubbs et al., 2017; Stubbs et al., 2018), but is ~20% to 40% of the response to isocaloric carbohydrate ingestion. When KME is coingested with carbohydrate, there is no difference in circulating insulin concentrations compared to carbohydrate alone (Stubbs et al., 2017; Myette-Côté et al., 2018; Cox et al., 2016).

Ingestion of 1.9 kcal.kg^{-1} body mass of KME (102-210 kcal; 21-43 g β HB) elevated whole blood β HB concentrations to $3.3 \pm 0.2 \text{ mM}$ after 1 h while attenuating the rise in circulating insulin and ghrelin compared to an isocaloric dextrose drink. This attenuation in plasma ghrelin resulted in a decrease in feelings of hunger, desire to eat and increased levels of fullness, suggesting KME may aid dietary strategies aimed at weight loss (Stubbs et al., 2018)

Ketone esters during exercise

Many of the responses seen at rest with exogenous ketone ingestion are prevalent during exercise. In one of a series of experiments, ingestion of 573 mg.kg⁻¹ body mass KME prior to 45 min cycling at 40% and 75% peak power output (W_{\max}) elevated β HB concentrations to ~4.0 mM and ~3.0 mM respectively and β HB contributed 18% and 16% towards total oxygen consumption. The exercise induced rise in plasma lactate was attenuated by ~50% during 1 h cycling at 75% W_{\max} compared to isocaloric carbohydrate ingestion (Cox et al., 2016).

The glucose lowering effect of ketone esters during exercise is well-demonstrated (Cox et al., 2016; Leckey et al., 2017; Vandoorne et al., 2017; Myette-Côté et al., 2018). This effect is caused by reduced hepatic gluconeogenesis (Mikkelsen et al., 2015) and occurs independently of differences in circulating insulin between conditions (Vandoorne et al., 2017; Myette-Côté et al., 2018).

Ketone ester ingestion prior to exercise attenuates the exercise induced increase in plasma lactate (Cox et al., 2016; Leckey et al., 2017). One of the main proposed benefits of exogenous ketones is a 'glycogen sparing' mechanism that will confer an advantage during periods of competition that are high intensity in nature and carbohydrate dependent (Pinckaers et al., 2016). Carbohydrate utilisation is reduced during submaximal exercise with KME ingestion in the fasted state (Cox et al., 2016), and it remains to be seen whether this sparing of carbohydrate can be overcome later in exercise or whether it will impair performance if the exercise duration is sufficiently extended. When compared to the effect of KS ingestion on plasma lactate, β HB concentrations between ~1.0 to 2.0 mM may be required to elicit an attenuation in plasma lactate concentrations.

The exercise-induced rise in FFA and glycerol is blunted with KME ingestion during 60 min cycling at 75% W_{\max} compared to isocaloric fat and carbohydrate ingestion. Plasma

insulin is elevated to $\sim 7 \text{ mU.L}^{-1}$ following KME ingestion but remains lower than isocaloric carbohydrate ingestion. KME increases the contribution of IMTG to total energy provision during exercise and results in a preservation of muscle glycogen (Cox et al., 2016).

Ketone body metabolism after exercise: post-exercise ketosis

Despite the aforementioned decline in KB concentrations at the onset of exercise, this pertains to situations where exercise has begun during hyperketonemia (Balasse et al., 1978; Fery & Balasse, 1983; Fery & Balasse, 1988; Cox et al., 2016). In the post-absorptive state, the pattern generally observed is for KB concentrations to rise gradually during prolonged exercise up to 0.2 to 0.4 mM, after which time post-exercise ketosis (PEK) of 0.3 to 2.0 mM is observed for several hours into recovery (Koeslag, 1982). Explained in terms of plasma kinetics, at cessation of exercise, the rate of appearance of KB increases coincident with a decrease in MCR relative to rates present during exercise. MCR remains above resting values for several hours after exercise, but ketogenesis exceeds ketolysis during this period.

On a mechanistic level, regulation likely resides at several sites including malonyl CoA-mediated regulation of fat transport into hepatocytes via CPT-1 in addition to availability of Ac-CoA for ketogenesis, and oxaloacetate for the TCA cycle as classically described for ketogenic regulation. Because oxaloacetate is a product of pyruvate formed during glycolysis, reductions in glycolytic flux with low glycogen content after intense exercise result in oxaloacetate moving to cytoplasm for preferential use in gluconeogenesis, which allows diversion of Ac-CoA towards ketogenesis during the post-exercise period rather than citrate synthesis for the TCA cycle. Additionally, the actions of insulin and glucagon exert a strong influence through activation and inhibition, respectively, of Ac-CoA carboxylase (ACC), which catalyses the synthesis of malonyl CoA from Ac-CoA. When liver glycogen becomes depleted and glucagon:insulin ratio is elevated, the synthesis of malonyl

CoA is reduced, thereby relieving the inhibition of fat transport into hepatocytes, and resulting in elevated concentrations of Ac-CoA. These regulatory mechanisms are acutely sensitive to nutrient manipulations before and after exercise and to aerobic exercise training, given their respective influences on substrate availability and utilisation during exercise.

Modulation of post-exercise ketosis by aerobic exercise training and nutrition intervention

An attenuation of, or abolished, post-exercise ketosis has been consistently observed in rodents and humans in response to aerobic exercise in trained versus untrained individuals (Johnson et al., 1969; Johnson & Walton, 1972; Rennie et al., 1974), or after a period of exercise training (Rennie & Johnson, 1974; Beattie & Winder, 1984, 1985; Adams & Koeslag, 1988; Adams & Koeslag, 1989; Ohmori et al., 1990). The aforementioned enhanced ketolytic capacity and downregulation of ketogenic capacity by training may play a role in these observations, but the majority of this work has been performed in comparisons with the absolute exercise intensity and duration being the same for comparisons (reviewed in (Koeslag, 1982)). This is problematic because the relative exercise intensity is the key determinant of the metabolic and hormonal response to acute exercise e.g. catecholamine responses, FFA mobilisation, glycogen utilisation among others. When trained and untrained participants have performed exercise at a similar relative intensity, PEK is blunted but not abolished in trained individuals (Rennie et al., 1974). Moreover, in rodents when exercise is completed to exhaustion, i.e. the trained rats exercise for longer than untrained, β HB concentrations are ~two-fold higher at the exercise cessation in the trained group (Askew et al., 1975). These divergent findings are likely due to the degree of liver glycogen depletion that occurs (Adams & Koeslag, 1988), inasmuch as higher concentrations of resting liver glycogen and attenuated rates of depletion are a consequence of training (Baldwin et al., 1975).

Therefore, PEK is strongly influenced by nutrition manipulation. High CHO feeding prior to exercise attenuates PEK regardless of training status (Rennie & Johnson, 1974; Askew et al., 1975; Koeslag et al., 1980), and CHO restriction increases PEK (Impey et al., 2016). Glucose ingestion at 2 h into recovery (Koeslag et al., 1982; Carlin et al., 1987) and alanine during recovery (Koeslag et al., 1980; Koeslag et al., 1985; Carlin et al., 1987) attenuate PEK

, but the glucose effect is not seen when glucose is ingested immediately after exercise. Alanine ingestion increases mitochondrial oxaloacetate concentrations in liver, thereby allowing condensation with Ac-CoA and diversion away from ketogenesis. This suggests that the early PEK response is determined by the extent of liver glycogen depletion and reduced glycolytic flux, whereas several hours into recovery is under regulation by insulin and FFA concentrations related to nutrition intake.

Metabolic consequences of post-exercise ketosis during recovery: a role for exogenous ketones as a recovery aid?

The physiological role for PEK is likely to favour the replenishment of muscle glycogen, consistent with classically described metabolic actions of ketosis in the sparing of protein and CHO stores during times of low CHO availability. During the post-exercise recovery period, in contrast to the reliance on CHO metabolism during exercise, muscle glycogen resynthesis has a high metabolic priority and is facilitated by an increase in fat oxidation and sparing of CHO sources for energy provision (Kiens & Richter, 1998). A priority for muscle glycogen resynthesis over liver glycogen resynthesis is suggested to occur because in ancestral terms, a depleted liver is less of hindrance to intense exertion than depleted muscle (Adams & Koeslag, 1988). To this end, the priority for muscle glycogen resynthesis is observed even during CHO restriction (Adams & Koeslag, 1989), and is

achieved through non-CHO sources such as lactate and alanine being used for hepatic gluconeogenesis and redistribution to skeletal muscle (Fournier et al., 2002). The contribution of PEK may be via the ability of KB to inhibit glycolysis and increase the conversion of glucose to glycogen as demonstrated in rat skeletal muscle in vitro (Maizels et al., 1977), and a perfused heart model in dogs (Laughlin et al., 1994). This effect is likely mediated by inhibition of PDH and phosphofructokinase (PFK) by elevations in Ac-CoA and citrate formation, respectively, as a consequence of metabolism of AcAc in mitochondria (Randle et al., 1964; Maizels et al., 1977; Laughlin et al., 1994; Kashiwaya et al., 1997).

This raises the possibility that an optimal post-exercise recovery milieu exists that includes both CHO and ketones to enhance recovery of muscle glycogen. This is not possible by conventional nutrition strategies because elevations in glucose, lactate and alanine ultimately limit ketogenesis and PEK. The suggestion is that the co-ingestion of KME and CHO in a recovery protocol can confer a metabolic advantage.

Following exercise, glycogen re-synthesis and stimulation of muscle protein synthesis (MPS) are of high importance to improve recovery. Athletes are recommended to follow optimal carbohydrate and protein-based fuelling related to the type, timing and amount of carbohydrate and protein to facilitate this recovery process and prepare the next bout of exercise. Ingestion of carbohydrate at a rate of $1.2 \text{ g.kg}^{-1} \text{ body mass. h}^{-1}$ during the first 4 h of recovery maximises the rate of muscle glycogen resynthesis; addition of $0.4 \text{ g.kg}^{-1} \text{ body mass. h}^{-1}$ of protein added to suboptimal carbohydrate doses during recovery increases the rate of glycogen resynthesis compared to suboptimal carbohydrate alone (Beelen et al., 2010). When the time between exercise bouts is limited ($< 8 \text{ h}$), nutritional strategies to augment glycogen resynthesis for subsequent exercise performance are a priority (Burke et al., 2017). During recovery from endurance exercise, the general pattern is a gradual rise in ketosis for several hours in the range of 0.3 to 2.0 mM, termed post exercise ketosis (Koeslag, 1982).

This rise in PEK is attenuated in aerobically-trained individuals when exercise is performed at the same absolute intensity, but not when performed at the same relative intensity (Johnson et al., 1969; Rennie and Johnson, 1974). Carbohydrate availability abolishes the rise in PEK during 3 h recovery following a mixture of high intensity and steady-state exercise, demonstrating the influence of circulating nutrients on ketosis (Impey et al., 2016). Similarly, alanine ingestion during recovery attenuates PEK (Koeslag et al., 1980; Carlin et al., 1987).

The physiological role for PEK is likely to favour the replenishment of muscle glycogen and enhancing MPS, consistent with the metabolic action of KB as a survival mechanism during periods of low carbohydrate availability. Resting muscle glycogen concentrations are not lower in 'fat adapted' ultra-endurance athletes following a high fat (70%) diet for 20 months; during and after 3 h submaximal running, the pattern of glycogen utilisation and recovery respectively was not different to a matched group of athletes following a high carbohydrate (55%) diet despite a larger contribution of fat towards total energy consumption (Volek et al., 2016). This is a pattern repeated in Alaskan sled dogs, who despite following a high fat/low carbohydrate diet show no cumulative signs of muscle glycogen depletion over several days of exercise or inability to oxidise carbohydrate (McKenzie et al., 2005; McKenzie et al., 2008). Glycogen replenishment following endurance exercise is facilitated by non-carbohydrate gluconeogenic sources, of which KB may be one. In a postabsorptive state, over 90% of gluconeogenesis is accounted for by gluconeogenic amino acids (alanine, glutamine), lactate and glycerol (Gerich et al., 2001), suggesting the preservation of muscle glycogen stores observed in fat-adapted humans and dogs is due to the increase in gluconeogenic substrate availability to the liver, preserving circulating glucose concentrations. KB act to inhibit glycolysis in rat skeletal muscle in vitro (Maizels et al., 1977), and a perfused heart model in dogs (Laughlin et al., 1994) and infusion of β HB stimulates muscle protein synthesis by 10% (Nair et al., 1988). These data would

suggest exogenous ketones could play in enhancing muscle glycogen re-synthesis and MPS in the recovery period after exercise.

In the fasted state, ingestion of 573 mg.kg⁻¹ body mass KME immediately after glycogen-depleting exercise alongside a 2 h hyperglycaemic clamp reportedly increases muscle glucose uptake by 32%, insulin concentrations two-fold and muscle glycogen concentrations by 50% compared to an isocaloric control (glucose only) drink (Holdsworth et al., 2017). Participants completed a glycogen-depleting bout of interval cycling lasting 115 ± 2 min. A 10 mM hyperglycaemic clamp was chosen to replicate circulating glucose concentrations during a practically-relevant nutrition intervention and KME or control drinks were provided immediately after exercise. Blood βHB concentrations peaked at 5.3 ± 0.5 mM during recovery and were 3.3 ± 0.2 mM at the end of the clamp. The stated 50% increase in muscle glycogen in the KME condition represents the absolute increase muscle glycogen after the clamp regardless of immediate post-exercise muscle glycogen concentrations (KET: 114 ± 23 mM.kg⁻¹ dry weight (DW); CON: 70 ± 23 mM.kg⁻¹ DW) i.e. muscle glycogen was decreased by 30% more in the KET condition than in the CON condition. At the end of the 2 h recovery phase, muscle glycogen concentrations during KME (132 ± 20 mM.kg⁻¹ DW) increased by 86% and during CON (94 ± 15 mM.kg⁻¹ DW) by 74%, representing a 12% increase in muscle glycogen re-synthesis with acute KME ingestion, which may be accounted for by the increase in caloric intake with KME (5 kcal. g⁻¹). The proposed mechanism for the enhanced muscle glycogen re-synthesis is stated to be the doubling of insulin concentrations with KME (KME: 31±5 mU.L⁻¹; CON: 16±3 mU.L⁻¹). However, early post-exercise muscle glycogen resynthesis appears to independent of circulating insulin, lasting 30-60 min when muscle glycogen concentrations are depleted below 150-200 mM.kg⁻¹ DW (Beelen et al., 2010), as is the case in this study.

Conversely, a separate study demonstrated that there is no effect on muscle glycogen repletion when 0.50 g.kg^{-1} body mass KME is ingested immediately after exercise, followed by 0.25 g.kg^{-1} body mass alongside $1.00 \text{ g.kg}^{-1} \text{ body mass.h}^{-1}$ carbohydrate and $0.30 \text{ g.kg}^{-1} \text{ body mass.h}^{-1}$ whey protein in the form of a recovery drink during a 5 h hour recovery (Vandoorne et al., 2017). This feeding regimen was chosen to elicit maximal rates of MPS (Morton et al., 2015) and muscle glycogen re-synthesis during the post-exercise recovery period (Burke et al., 2017). Participants were provided with a high carbohydrate (70%) breakfast and completed a glycogen-depleting bout of unilateral knee extensions. Whole blood βHB concentrations peaked at $4.3 \pm 0.5 \text{ mM}$ at 4 h of recovery. Unilateral exercise depleted muscle glycogen concentrations by $\sim 60\%$ during both PLA and KME, reaching $170 \pm 22 \text{ mM.kg}^{-1} \text{ DW}$ and $183 \pm 21 \text{ mM.kg}^{-1} \text{ DW}$, respectively. No difference in plasma insulin concentrations was observed during the 5 h recovery period, reaching $\sim 25 \text{ mU.L}^{-1}$ at 90 and 300 min of recovery in KME and PLA conditions and blood glucose was $\sim 1.0 \text{ mM}$ lower during KME during the final 4 h of recovery. Differences in study design may explain the contrasting findings of these two studies, e.g. different exercise modalities in cycling vs. unilateral exercise, the different method of elevating plasma glucose availability (hyperglycaemic clamp vs. powdered drinks), the different lengths of recovery period, and the magnitude of glycogen depletion.

Regarding anabolic signalling in skeletal muscle, ingestion of KME alongside optimal post-exercise carbohydrate and protein fuelling increased the phosphorylation of proteins involved in the mammalian target of rapamycin (mTORC1)-regulated muscle protein synthesis cascade. In vivo phosphorylation of S6K1 increased \sim two-fold and 4E-BP1 increased 60% with in the KME condition compared to placebo ingestion. In vitro, the addition of $\beta\text{HB} + \text{AcAc}$ ($4.0 + 1.4 \text{ mM}$, respectively) to 1.5 mM leucine increased S6k1 phosphorylation \sim three-fold and 4E-BP1 by 51% in skeletal muscle cells. Moreover, the

addition of β HB + AcAc to 1.5 mM leucine increased the rate of MPS by two-fold compared to leucine alone. However, KB were not added to the higher leucine concentration of 5.0 mM, which alone stimulated MPS to a higher extent than a combination of β HB, AcAc and 1.5 mM leucine (Vandoorne et al., 2017). Therefore, it remains to be explored whether AcAc and β HB have anabolic effects similar to leucine alone, despite earlier reports of a 10% in MPS after β HB infusion in humans (Nair et al., 1988).

To examine the anticatabolic effect of KB, infusion of $2.4 \text{ mL} \cdot \text{kg}^{-1} \text{ body mass} \cdot \text{h}^{-1}$ D/L- β HB increased plasma β HB concentrations to 3.5 mM in a LPS stimulated inflammatory model in humans (Thomsen et al., 2018). Compared to FFA, infusion of D/L- β HB reduced forearm phenylalanine release by ~70%, reflecting a net decrease in muscle protein loss and whole body phenylalanine-to-tyrosine degradation was reduced. The insulin-Akt-mTOR and p70 S6K phosphorylation was unaffected by D/L- β HB infusion. These results suggest an anti-catabolic effect of β HB at a muscle and whole body level, rather than a direct anabolic effect. Direct comparisons between β HB and amino acids and simultaneous administration of both substrates is required before any statement can be made about the efficacy of β HB as a treatment in muscle wasting/catabolic models.

These preliminary data suggest that when undertaking best practice recovery guidelines, the addition KME confers no benefit to glycogen re-synthesis, but may enhance MPS. However, it remains to be seen whether addition of KS, KDE or KME to suboptimal delivery of carbohydrate post exercise may, like protein, aid with glycogen re-synthesis.

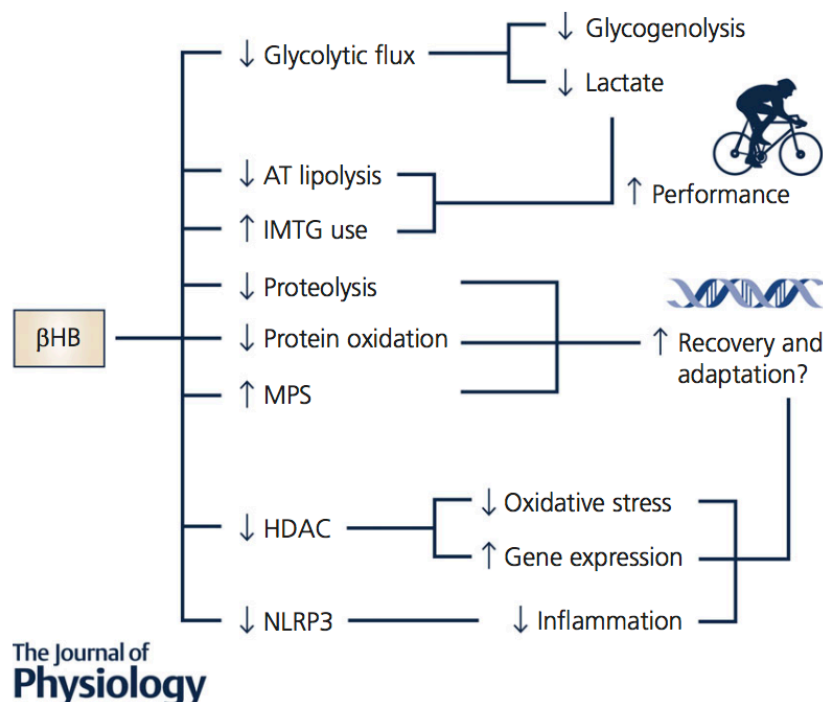


Figure 2.6 βHB as a metabolic regulator and signalling metabolite

Effects of elevating βHB through acute nutritional ketosis may be mediated by acute regulation of substrate utilisation that may enhance performance, and /or possibly through regulation of recovery and adaptive processes related to inflammation, oxidative stress and changes in gene expression.

Effects beyond fuelling: βHB as a HDAC inhibitor

As investigative techniques in molecular biology evolve, so too does our appreciation of how complex integrative signalling networks regulate skeletal muscle adaptation in response to stimuli such as nutrient manipulation and exercise training (Egan & Zierath, 2013). Previously considered relatively inert outside their primary metabolic function, numerous substrates and metabolites are emerging as important regulators of intracellular signalling and tissue adaptation (Hashimoto et al., 2007; Gao et al., 2009; Morton et al., 2009; Roberts et al., 2014). Noteworthy for the present review is the recent identification of AcAc as a regulator of skeletal muscle satellite cell proliferation and muscle regeneration (Zou et al., 2016), and βHB as an inhibitor of HDACs (Shimazu et al., 2013) and the NLRP3 inflammasome (Youm et al., 2015). The latter observations are a consequence of βHB, in

essence, acting as a signalling metabolite to regulate gene expression and metabolic processes (Figure 2.6).

Histone acetyltransferases (HATs) and HDACs are enzymes that facilitate the addition or removal, respectively, of acetyl moieties from specific lysine residues on histones and target proteins (McKinsey et al., 2001). In general, hyperacetylation of histone tails induces transcriptional activation while hypoacetylation is associated with transcriptional repression. Class IIa HDACs (HDAC4, -5, -7 and -9) are highly expressed in skeletal muscle (McKinsey et al., 2001) and their function is responsive to both aerobic endurance exercise in humans (McGee et al., 2009; Egan et al., 2010) and nutritional intervention in rodents (Gao et al., 2009; Shimazu et al., 2013). An acute bout of aerobic exercise increases class IIa HDAC phosphorylation and subsequent nuclear exclusion, thus inhibiting HDAC-mediated repression of specific exercise-responsive genes such as GLUT4 and PGC-1 α (McGee & Hargreaves, 2004; McGee et al., 2009; Egan et al., 2010). This suggests that compounds that inhibit or disrupt HDAC inhibition could be used to mimic or enhance adaptations to exercise.

Regulation of HDAC activity by nutrients including butyrate and β HB has also been established (Gao et al., 2009; Shimazu et al., 2013). Butyrate, a short chain fatty acid formed via the fermentation of indigestible dietary fibres by microbial species in the gut, is a potent inhibitor of HDAC activity (Gao et al., 2009). Mice supplemented with sodium butyrate are resistant to diet-induced obesity, and have elevations in markers of skeletal muscle mitochondrial biogenesis analogous to exercise effects (Gao et al., 2009). β HB is structurally similar to butyrate, and although not as potent as butyrate, also inhibits HDAC class I and II activity in a dose-dependent manner and suppressed oxidative stress responses (Shimazu et al., 2013). Importantly, HDAC inhibition by β HB both in vitro and in vivo is evident at physiologically-relevant concentrations of β HB i.e. 1 to 4 mM, which is similar to those attained during fasting, PEK and exogenous ketone ingestion (Figure 2.6) (Clarke et al., 2012;

Kesl et al., 2016). However, although the inhibitory effects were observed in multiple tissues, they remain to be confirmed in skeletal muscle. If confirmed, it will be intriguing to explore whether, apart from the aforementioned ergogenic effects, exogenous ketone supplementation complements exercise-mediated adaptive changes associated with modulating HDAC function.

Effects on physical performance

Ketone esters

The aforementioned work by Cox and colleagues at the University of Oxford remains the seminal study demonstrating the potential benefits of exogenous ketones on endurance performance (Cox et al., 2016). Ingestion of 573 mg.kg^{-1} body mass KME improved 30 min max distance cycling time trial performance by $\sim 2\%$ ($411 \pm 162 \text{ m}$), following a 1 h pre-load at $75\% W_{\text{max}}$. This improvement coincided with a $\sim 1.5\text{-}2.0 \text{ mM}$ reduction in plasma lactate and a reduction in plasma glucose. The drinks consumed in these trials were isocaloric, meaning performance improved despite receiving less carbohydrate throughout the KME arm of the trial (Cox et al., 2016). Since then, several reports on the physical performance effects of acute exogenous ketones ingestion have emerged (O'Malley et al., 2017; Rodger et al., 2017; Leckey et al., 2017; Waldman et al., 2018). These studies all use exogenous ketone supplementation, but in different forms, and it is important to distinguish between the different forms given that each have different metabolic effects that can potentially alter performance.

Ingestion of 500 mg.kg^{-1} body mass KDE impaired performance during an ecologically-valid 31.2 km time trial in professional cyclists (Leckey et al., 2017). Participants ingested the bolus as two 250 mg.kg^{-1} body mass aliquots 50 and 30 min prior to the time trial, having consumed a high carbohydrate breakfast (2 g.kg^{-1} body mass) on the

morning of each trial. Participants received 200 mg caffeine, a 6% carbohydrate sports drink and a caffeine gel during the trial to mimic the real-world practice in professional cycling. Performance was impaired by $2 \pm 1\%$, (58.2 s; ES -0.42 small), explained by a 3.7% reduction in power output and was accompanied by a high prevalence of gastrointestinal distress among participants in the KDE condition. The lower plasma β HB concentrations in the KDE study makes it difficult to compare the performance findings with the KME work (Cox et al., 2016). Plasma β HB reached ~ 0.4 mM pre-time trial, suggesting KDE at this dose is currently ineffective at achieving nutritional ketosis (>0.5 mM β HB), a problem complicated by the reports of high prevalence of gastrointestinal distress.

To date, ketone ester work has focused on endurance athletes due to the proposed glycogen sparing mechanism of elevated circulating β HB concentrations (Cox et al., 2016). Sport nutrition guidelines for team sports promote high dietary carbohydrate intake prior to and during competition to maximise muscle glycogen stores with a view to enhancing performance (Burke et al., 2006). Therefore, nutritional strategies that could attenuate the reduction of muscle glycogen during match play (Mohr et al., 2003) is of value to scientists and practitioners. Whether ketone ester ingestion benefits team sport activity, and indeed most other sporting contexts, remains to be confirmed.

Ketone salts

There are three reports on exercise performance effects of acute KS ingestion (O'Malley et al., 2017; Rodger et al., 2017; Waldman et al., 2018). These studies use a variety of doses of KS. Ingestion of two servings of a commercially available KS (KetoForce), providing 11.7 g D/L- β HB per serving elevated whole blood β HB concentrations to 0.6 ± 0.2 mM but did not improve mean power output during 4 min maximal performance test in highly trained cyclists (KS: 364 ± 56 W; PLA: 355 ± 46 W) (Rodger et al., 2017). Similarly,

ingestion of one serving of KS (PerfectKeto) elevated whole blood β HB concentrations to 0.53 ± 0.19 mM but had no effect on mean power output (KS: 715 ± 94 W; PLA: 714 ± 93 W) or peak power output (KS: 969 ± 157 W; PLA: 955 ± 151 W) during 4x15 s maximal sprints on a cycle ergometer (Waldman et al., 2018). Ingestion of 0.3 g.kg^{-1} body mass KS (KetoForce) 50 min prior to exercise elevated blood β HB concentrations to ~ 0.8 mM and lowered mean power output during a 150 kJ time trial (~ 10 km) (-7%, -16W) and resulted in a longer time to finish (KS: 711 ± 137 s; PLA: 665 ± 120 s) (O'Malley et al., 2017).

Overall, KS only modestly elevate β HB concentrations into the range of 0.3-0.8 mM during exercise, all measured by handheld monitors. Given the described overestimation of β HB concentrations by handheld monitors compared to gold-standard laboratory-based assays, it is likely β HB was lower than described in these works on KS ingestion and exercise performance. Secondly, elevation of β HB concentrations is postulated to impair high intensity exercise performance through an inhibition of glycolytic flux via inhibition of PDH and phosphofructokinase (PFK) by increases in NADH:NAD⁺, acetyl-CoA:CoA or citrate. The performance measures used in these studies were high intensity in nature, which are unlikely to benefit from exogenous ketone supplementation.

Effects on cognitive performance

The primary physiological role of KB is to provide glucose-dependent organs with a usable source of carbon during periods of low carbohydrate availability. FFA acids are unable to cross the blood-brain barrier and must be converted to KB in order to provide energy to the brain under these circumstances (Owen et al., 1967). KB account for $\sim 60\%$ of the brain's energy demands after 5-6 weeks of starvation in obese patients (Owen et al., 1967). After short term (3.5 d), brain glucose metabolism is reduced to $\sim 70\%$ of control values, measured

by positron emission tomography, and coincides with an increase in KB oxidation in compensation as an alternative fuel source to glucose (Hasselbalch et al., 1994).

Elevations in circulating KB concentrations have marked effects on brain energy metabolism. Elevating β HB concentrations to 4.0-8.0 mM by means of chronic fasting causes a 50% reduction in brain glucose MCR (Zhang et al., 2013). Moreover, infusion of D/L- β HB increased plasma D- β HB concentrations to 5.5 ± 0.4 mM (similar to that achieved by prolonged fasting) and this coincided with a 14% reduction in cerebral glucose utilisation, a 30% increase in cerebral blood flow and unchanged oxygen consumption (Svart et al., 2018). The authors suggest may reduce oxidative stress, explaining the well-established neuroprotective properties of KB (Svart et al., 2018). This work was performed in the elderly, with similar results observed in young subjects (Hasselbalch et al., 1996) and is not related to the age-related decline in glucose utilisation.

Exogenous ketones may also play a role in improving cognition in Alzheimer's Disease (AD) and during mild cognitive impairment (MCI) by helping to overcome the characteristic brain energy deficit (Veech, 2004). During AD and MCI, brain glucose uptake is impaired while uptake of KB is unchanged and the elevation of KB through oral medium chain triglyceride supplementation improves cognitive outcomes in these populations (Reger et al., 2004; Krikorian et al., 2012; Cunnane et al., 2016).

Similarly, glucose uptake and oxidation in the brain is reduced in an exercise intensity-dependent manner, as evidenced by a reduction of $\sim 30\%$ when exercising for 35 min at $\sim 75\%$ compared to $30\% \dot{V}O_{2\text{peak}}$ (Kemppainen et al., 2005). Conversely, the uptake and oxidation of circulating lactate by the brain increases with increasing exercise intensities paralleling the downregulation of carbohydrate oxidation. Under resting conditions the uptake and oxidation of lactate is minimal but >4.0 mM the brain goes from a state of lactate release to uptake, related to an increase in arterial concentrations (Rasmussen et al., 2011). At

higher lactate concentrations, lactate may replace glucose as a source for the brain by up to 25% (Quistroff et al., 2008; Rasmussen et al., 2011). The point to be made here is that the brain can oxidise circulating substrates in an arterial concentration-dependent manner. AD, MCI and intense exercise are all characterised by a decrease in brain glucose oxidation that is compensated for by other substrates capable of crossing the blood brain barrier. Elevations in β HB concentrations via ketogenic supplementation can compensate for the deficit in glucose oxidation in AD and MCI (Cunnane et al., 2016), so it is tempting to speculate whether exogenous ketones may play a similar role in exercise contexts.

These works suggest that elevating circulating KB concentrations reduces the brain's reliance on glucose for fuel provision. Exercise places a high demand on whole body carbohydrate stores, and nutritional interventions that ensure normal brain energy homeostasis warrant investigation. There is no clear relationship between anaerobic maximal bouts of exercise and cognitive processes, while aerobic exercise lasting <90 min exerts a selective influence on cognition, including improvements in complex problem solving and attentional processes that are involved in response inhibition (Tomprowski, 2003). There is a facilitating effect of physical activity and cognitive performance, with the majority of work on arousal and reaction time, possibly due to elevated levels of cortical catecholamines. Much of this work has been performed using low intensity exercise at 40% to 60% $\dot{V}O_{2\text{peak}}$ (Davranche et al., 2006), while both acute and chronic physical activity improves executive function (Best, 2012). It is important to be cognisant of the selective effects of exercise on different aspects on cognitive function being measured in nutritional supplement trials. Submaximal exercise of >2 h in duration that depletes energy (carbohydrate) stores and dehydrates the athlete is more likely to negatively impact cognitive function (Cian et al., 2000; Cian et al., 2001). If there is a central effect of KB during exercise, it may be that KB

could help to conserve cognitive function during endurance exercise by alleviating the reliance on glucose as a fuel source, and attenuating the decline in brain energy stores.

Work on ketone ester ingestion, cognitive performance and brain metabolism is focused in rodents using KME (Murray et al., 2016; Pawlosky et al., 2017) and KDE (Ciarlone et al., 2016; Ari et al., 2016; Kovács et al., 2017). Chronic feeding of 2.6 g.day^{-1} KME for 5 days improved decision-making and reduced time to completion of a radial maze task by 38% versus rats fed an isocaloric high carbohydrate diet (Murray et al., 2016). Concentrations of cortical and hippocampal glycolytic intermediates is lower following chronic treatment of KME and cortical citrate synthase concentrations are significantly higher, while citrate synthase, isocitrate, alpha-ketoglutarate, succinate, fumarate and acetyl Co-A were all elevated after chronic ingestion of KME (Pawlosky et al., 2017). These changes coincided with a significantly more energetic ΔG in the hippocampus of KME-fed rats. Chronic KDE ingestion decreases audiogenic and chemically-induced seizure activity in mice when βHB concentrations are elevated to between 0.8-1.0 mM during 8 wk of treatment. (Ciarlone et al., 2016). Furthermore, chronic KDE administration by intra-gastric gavage decreases absence epileptic activity when βHB concentrations are elevated to $\sim 1.8 \text{ mM}$ at the end of 1 wk of treatment (Kovács et al., 2017). These results are explained by an increase in GAD65/67 hippocampal protein expression, which code for GABA, with this elevation being antithetic to the reduction in GABA that contributes to seizure activity.

The cognitive effects of exogenous ketones are underexplored in humans with only one study incorporating cognitive tasks (Waldman et al., 2018). Following acute ingestion of KS, participants completed a 5-min reaction test during their warm up and immediately following 4x15 s anaerobic Wingate sprints on a cycle ergometer. No effect on reaction time was observed after the anaerobic activity despite a higher fatigue index in the KS condition (KET: $32.3 \pm 13.9 \text{ W/s}$; PLA: $29.2 \pm 12.6 \text{ W/s}$). However, a learning/order effect is noted by

the authors for hits and misses on the same test with repeated testing (Waldman et al., 2018). This protocol may also not have been long enough to cause a reduction in whole body energy stores or dehydration to cause substantial decline in cognitive function (Tomprowski, 2003), and would seemingly agree that short duration exercise has little impact on cognitive function. Like physical performance, this work suggests low concentrations of β HB (<1.0 mM) may not effect cognitive performance in this high intensity exercise context. Whether ketone ester ingestion has an effect on cognitive performance in endurance and high intensity exercise challenges remains to be studied.

State-of-the-art in exogenous ketone supplementation and exercise performance

Exogenous ketone supplements come in multiple forms; racemic ketone salts, a (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester, and a R,S-1,3-butanediol acetoacetate diester. Each of these supplements elevates β HB concentrations but differ in their effect on the metabolic response to exercise, and possibly exercise performance. There is a strong rationale for the performance-enhancing benefits of exogenous ketone in sport and the same is true for performance impairment, much of which would depend on the supplement being consumed and the nature of the exercise challenge. The (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester is the only supplement shown to be beneficial to endurance performance in cyclists, hypothesised to result from a ‘glycogen sparing’ effect. Whether this ergogenic effect will be observed in other sporting contexts, namely high intensity intermittent team sports, that rely on a high rate of carbohydrate oxidation for sustaining performance, and other modalities of endurance exercise (i.e. running) remains to be determined. Given the role of KB in fuel provision to the brain during short and long term starvation, the effect of exogenous ketone supplements on aspects of cognitive performance, including reaction time, sustained attention, and decision making is of interest.

Chapter 3

Evans M, Patchett E, Nally R, Kearns R, Larney M, Egan B (2018) Effect of acute ingestion of β -hydroxybutyrate salts on the response to graded exercise in trained cyclists. *European Journal of Sport Science* 18(3):376–86.

Abstract

Introduction: Acute ingestion of ketone salts induces nutritional ketosis by elevating β -hydroxybutyrate (β HB), but few studies have examined the metabolic effects of ingestion prior to exercise. **Methods:** Nineteen trained cyclists (12 male, 7 female) undertook graded exercise (8 min each at ~30%, 40%, 50%, 60%, 70%, and 80% $\dot{V}O_{2\text{peak}}$) on a cycle ergometer on two occasions separated by either 7 or 14 days. Trials included ingestion of boluses of either (i) plain water (3.8 mL kg body mass⁻¹) (CON) or (ii) β HB salts (0.38 g kg body mass⁻¹) in plain water (3.8 mL kg body mass⁻¹) (KET), at both 60 min and 15 min prior to exercise. **Results:** During KET, plasma β HB concentrations increased to 0.33 ± 0.16 mM prior to exercise and 0.44 ± 0.15 mM at the end of exercise (both $p < .05$). Plasma glucose was 0.44 ± 0.27 mM lower ($p < .01$) 30 min after ingestion of KET and remained ~0.2mM lower throughout exercise compared to CON ($p < .001$). Respiratory exchange ratio (RER) was higher during KET compared to CON ($p < .001$) and 0.03–0.04 higher from 30% $\dot{V}O_{2\text{peak}}$ to 60% $\dot{V}O_{2\text{peak}}$ (all $p < .05$). No differences in plasma lactate, rating of perceived exertion, or gross or delta efficiency were observed between trials. Gastrointestinal symptoms were reported in 13 out of 19 participants during KET. **Discussion:** Acute ingestion of β HB salts induces nutritional ketosis and alters the metabolic response to exercise in trained cyclists. Elevated RER during KET may be indicative of increased ketone body oxidation during exercise, but at the plasma β HB concentrations achieved, ingestion of β HB salts does not affect lactate appearance, perceived exertion, or muscular efficiency.

Introduction

The relationship between energy provision and factors intrinsic and extrinsic to exercise is complex, but has traditionally focused on the relative contribution of carbohydrate and fat being regulated by the intensity and duration of an exercise challenge (Egan & Zierath, 2013). Many sports nutrition strategies are based around optimizing carbohydrate provision before and during performance (Burke, 2015), but there is increasing interest in alternative fuelling strategies, particularly in endurance sports, including low carbohydrate and ketogenic diets, and the use of exogenous ketone supplements (Burke, 2015; Evans, Cogan, & Egan, 2017).

Ketone bodies [namely β -hydroxybutyrate (β HB) and acetoacetate (AcAc)] are produced in the liver during periods of low glucose availability such as during fasting, starvation and ketogenic diets (Robinson & Williamson, 1980; Balasse & Fery, 1989; Laffel, 1999). Although principally acting as an alternative fuel source for the brain when glucose concentrations are diminished, ketone bodies are also used by skeletal muscle to provide up to 10% of energy during exercise in the fasted state (Franken, Neef & Balasse, 1974; Balasse, Fery, & Neef, 1978; Fery & Balasse, 1983; Fery, Wahren, Sato, Ostman, Hagenfeldt, & Felig, 1984). However, the direct contribution to energy provision may be secondary to the potential metabolic action of supplemental ketones. For instance, ketone bodies have wide-ranging metabolic effects on peripheral tissues such as glucose sparing, anti-lipolytic effects and stimulation of muscle protein synthesis (Maizels et al., 1977; Nair et al., 1988; Mikkelsen et al., 2015). During moderate intensity exercise, infusion of sodium AcAc after an overnight fast attenuates the rise in plasma lactate (Fery & Balasse, 1988), whereas sodium β HB infusion similarly alters the metabolic response to very intense exercise in rats (Kamysheva & Ostrovskaya, 1980), and ischemic forearm exercise in humans (Lestan et al., 1994).

Despite these observations, the potential performance benefits of ketone bodies have been unexplored until the recent emergence of exogenous ketone supplements in the form of ketone esters and ketone salts. For instance, acute ingestion of the (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester produced plasma β HB concentrations of ~ 3 mM during exercise, and improved 30 min time-trial performance by 2% in elite cyclists (Cox et al., 2016). Ketone esters are not commercially-available to date, but ketone salts represent a cheaper, more readily-available exogenous ketone supplement. These salts comprise of the free acid form of β HB buffered with sodium, potassium, and/or calcium salts, but are less effective at elevating plasma β HB concentrations compared to the ketone monoester (Stubbs et al., 2017). The effects of acute ketone salt ingestion on short-duration, high intensity exercise performance in human has been the subject of two recent reports (O'Malley et al., 2017; Rodger et al., 2017), both of which did not observe the performance benefits associated with the ketone monoester. Given that this is an emerging field of research, and to better understand the impact of ketone salt ingestion on responses across a range of exercise intensities, the purpose of the present study is investigate the effect of acute ingestion of a commercially-available β HB salt formulation on the metabolic and physiological responses to a graded submaximal exercise session in young, trained male and female cyclists. We hypothesise that ingestion of the β HB salt formulation will alter the metabolic response to exercise based on our pilot data collection.

Methods

Participants

Nineteen trained cyclists [12 male, 7 female (Table 3.1)] gave written informed consent to participate after written and verbal explanation of the procedures. Ethical approval (permit number: LS-15-82-Evans-Egan) was obtained from the University College Dublin Research Ethics Committee. All participants were active in regular cycling training (≥ 6

sessions per week) and competition in road, time-trial, and/or triathlon disciplines, and had been competing in their respective discipline for at least one calendar year.

Table 3.1 Participant anthropometrics and fitness profile

	Whole cohort (<i>n</i> = 19)	Males (<i>n</i> = 12)	Females (<i>n</i> = 7)	Males vs. Females <i>P</i> value
Age (y)	26.8 ± 7.6	25.1 ± 6.2	30.3 ± 8.8	0.148
Height (m)	174.3 ± 8.9	178.7 ± 7.4	166.7 ± 5.7	0.002
Body mass (kg)	69.0 ± 9.7	73.8 ± 6.8	60.9 ± 8.5	<0.001
Body fat (%)	17.6 ± 6.8	13.7 ± 3.9	24.3 ± 5.0	<0.001
FFM (kg)	57.7 ± 10.5	64.3 ± 5.9	46.4 ± 5.3	<0.001
W _{max} (W)	325 ± 67	368 ± 40	251 ± 23	<0.001
LT (W)	245 ± 59	278 ± 41	187 ± 36	<0.001
VO _{2peak} (L·min ⁻¹)	4.3 ± 8.5	4.8 ± 4.3	3.3 ± 4.3	<0.001
VO _{2peak} (mL·kg ⁻¹ ·min ⁻¹)	61.1 ± 7.1	65.5 ± 5.6	54.9 ± 3.6	<0.001
VO _{2peak} (mL·kg FFM ⁻¹ ·min ⁻¹)	74.0 ± 5.1	75.2 ± 5.1	71.8 ± 4.7	0.171

Data are presented as mean ± SD. LT, power output at 4 mM lactate threshold; FFM, fat-free mass

Experimental design

Participants visited the laboratory for exercise tests on three separate occasions. All tests were performed on the same electronically-braked stationary cycle ergometer (Lode Excalibur Sport, Netherlands). Saddle height and handlebar position were adjusted to each participant's preference, but kept consistent for the three visits. Participants performed the exercise tests in their own cycling shoes with appropriate pedals provided by the laboratory.

Body mass and height were measured using digital scales (SECA, Germany) and a wall-mounted stadiometer (Holtain, UK), respectively. Body composition was measured using dual-energy x-ray absorptiometry (Lunar iDXA, GE Healthcare, UK).

During their first visit to the lab, participants performed a submaximal incremental exercise test to establish their lactate threshold, after which they performed an incremental exercise test to volitional exhaustion to establish their peak oxygen uptake ($\dot{V}O_{2\text{peak}}$). Two experimental trials, each comprised of a graded exercise test of six stages (at power outputs corresponding to approximately 30%, 40%, 50%, 60%, 70% and 80% $\dot{V}O_{2\text{peak}}$), with each stage lasting 8 min (Figure 3.1), were performed during subsequent visits in a randomized cross-over design. Each experimental trial was identical with the exception of a drink consumed in the hour prior to each exercise test, namely plain water (CON), or β HB salts (KET).

Incremental exercise tests

Assessment of lactate threshold and $\dot{V}O_{2\text{peak}}$ was performed in accordance guidelines from the British Association of Sport and Exercise Sciences (BASES) (Davison & Wooles, 2007; Spurway & Jones, 2007). Briefly, for determination of lactate threshold, participants completed 4 min stages (3 min of cycling and 1 min of rest), starting at 50 W. The power output was increased by 50 W for the next two stages, and 30 W thereafter until a blood lactate concentration (Lactate Pro 2, Japan) of 4 mM was exceeded. After a 15 min rest, $\dot{V}O_{2\text{peak}}$ was determined via an incremental test to exhaustion. Participants began cycling at a pre-determined power output based on body mass as per the BASES guidelines, and power output was progressively increased by was increased by 20 W.min⁻¹ for males and 15 W.min⁻¹ for females thereafter until volitional exhaustion.

Pre-trial preparation

All experimental trials were performed between 07:00 and 10:00, but on an individual basis, participants performed their second trial at the same time ± 1 h as their first trial. Pre-trial preparation was the same for each experimental trial. Participants were asked to abstain from alcohol for 48 h and caffeine for at least 12 h, and refrain from strenuous exercise training for the day prior to each trial. Each trial took place after a standardized 10 h overnight fast. Participants were asked to keep a one-day portion estimate food diary for the day corresponding to two days prior to the first trial. They were instructed to repeat this pattern of intake before their second trial. On the day immediately prior to both experimental trials, participants were provided with a standardized diet (Gourmet Fuel, Ireland), which provided 40 kcal.kg body mass⁻¹ at a macronutrient ratio of 40% carbohydrate, 30% protein and 30% fat. Male participants performed the two experimental trials separated by 7 or 14 days. Because the phase of the menstrual cycle influences fuel utilization during exercise (Oosthuyse & Bosch, 2010), female participants performed the two experimental trials separated by 7 days, but within the early to mid-luteal phase of their menstrual cycle.

Experimental trials

Experimental trials were performed in a randomized cross-over open-label design, and were identical with the exception of the drink consumed in the hour prior to each exercise test. The open-label design was chosen because of the difficulty in masking the pungent taste of the β HB salts, and considered acceptable because there was no performance element to the experimental design. Neither the study participants nor research personnel were blinded to allocation of condition, with the exception of the laboratory technician who did undertake analysis of the blood samples in a blinded manner.

During each trial, a bolus of a given drink was ingested at both 60 min and 15 min

prior to the commencement of exercise (Figure 3.1). Each bolus consisted of either (i) plain water provided at 3.8 mL.kg body mass⁻¹ (CON), or (ii) β HB salts (KetoCaNa, Prototype Nutrition, IL USA) provided at 0.38 g.kg⁻¹ body mass dissolved in 3.8 mL.kg body mass⁻¹ plain water (KET). Each bolus serving of KET provided ~18.5 g β HB, 2.1 g sodium and 1.8 g calcium, which is approximately 60% more β HB than the manufacturer's guidelines of 11.7 g β HB per serving. This timing and dosing strategy was based on our own pilot experiments (unpublished data) showing that plasma β HB concentration peaked at 60 min after ingestion of a single bolus, and that a greater elevation in plasma β HB concentration could be achieved with two smaller doses of β HB salts compared to a single larger dose equivalent to the same total amount of β HB salts.

Upon arrival at the laboratory, an indwelling catheter was introduced into an antecubital vein for serial blood sampling at rest (-60, -30 and 0 min) and during exercise (last 30 s of each 8 min stage) (Figure 3.1). The catheter was maintained patent between samples with saline (0.9% sodium chloride). The exercise test was graded and consisted of six stages at power outputs corresponding to approximately 30%, 40%, 50%, 60%, 70% and 80% $\dot{V}O_{2\text{peak}}$, with each stage lasting 8 min (Figure 3.1). Expired air was collected continuously throughout each exercise test on a breath-by-breath basis (COSMED Quark b2, Italy). During the last 30 s of each 8 min stage, HR (Polar, Finland) and RPE (Borg scale) were recorded, and a blood sample was collected for measurement of plasma β HB, lactate, and glucose concentrations.

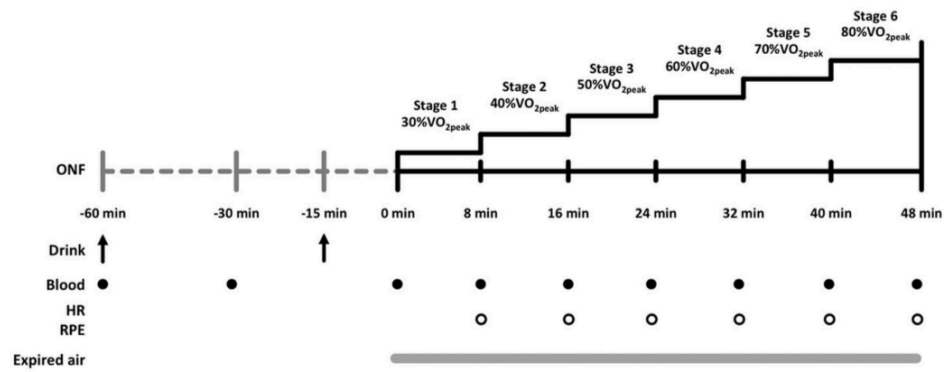


Figure 3.1 Experimental design schematic

Blood analysis

Blood samples (4 mL) were collected in plastic tubes containing sodium fluoride/potassium oxalate (Vacuette Glucose tubes, Greiner-Bio-One, Germany) for subsequent analysis. Samples were stored on ice before centrifugation at 3000 g for 10 min at 4°C, after which three aliquots of plasma were separated for storage at -80°C until later analysis of plasma β HB, lactate, and glucose (RX Daytona, Randox Laboratories, UK; assay codes RB1007, LC2389 and GL364 respectively).

Data analysis

Cardiopulmonary and metabolic parameters. Minute ventilation (\dot{V}_E), $\dot{V}O_2$, carbon dioxide production ($\dot{V}CO_2$) and RER were calculated from an average of breath-by-breath measurements during the last 30 s of each stage in the incremental exercise tests, and during the last 2 min of each stage in main experimental trials. Oxygen pulse (O_2 pulse), defined as oxygen uptake per heartbeat and expressed in $mL \cdot beat^{-1}$, was calculated by dividing $\dot{V}O_2$ ($L \cdot min^{-1}$) by HR ($beats \cdot min^{-1}$) during the last 30 s of each stage.

Substrate utilization. The rate of energy expenditure ($kcal \cdot min^{-1}$) during each stage was calculated from the average $\dot{V}O_2$ and $\dot{V}CO_2$ values during the last 30 s of each stage using equations applied on intensity-dependent basis (Jeukendrup & Wallis, 2005). Rates of

carbohydrate and fat oxidation are not reported because of the likely error introduced into these calculations by the oxidation of β HB and AcAc, which yield respiratory quotient values of 0.89 and 1.00 respectively (Frayn, 1983). Reporting oxidation rates based on RER is inaccurate during periods of nutritional ketosis unless appropriate correction factors for CO_2 displacement, and excretion of ketone bodies in urine and expired air are employed (Frayn, 1983), which were beyond the scope of the current work.

Mechanical efficiency. Gross efficiency (GE) was calculated as the ratio of the work performed per minute (W converted to $\text{kJ}\cdot\text{min}^{-1}$) to the energy expended per minute ($\text{kJ}\cdot\text{min}^{-1}$) at each stage, expressed as a percentage. Delta efficiency (DE) was calculated as the ratio of the change in work performed per minute to the change in energy expended per minute between each stage, expressed as a percentage (Gaesser & Brooks, 1975).

Statistical analysis

Data were evaluated using GraphPad Prism 6 (GraphPad Software, Inc., CA USA), and are presented as mean \pm SD, with the exception of Figure 3.2 where error bars represent 95% confidence intervals. The experiment was powered based on change in RER as the primary outcome, which was chosen as a measure of an altered metabolic response. Based on the aforementioned pilot data where a 0.034 ± 0.015 difference in RER between KET and CON was observed, $n=13$ participants would have been required given $\alpha=0.05$ and $1-\beta=0.8$ (GPower v3.1). Independent samples t-tests were used to determine differences between male and female participants for baseline characteristics. Two-way (condition \times intensity) repeated measures analysis of variance (ANOVA) was used to determine differences between the two experimental trials for variables with serial measurements. When a main effect of condition, or an interaction effect between condition and intensity, was indicated, post-hoc testing was performed using Holm-Sidak's multiple comparisons test with multiplicity-adjusted P values

to compare KET to CON at respective time points. The data were tested for normality and sphericity prior to proceeding with the tests described. In addition, standardised differences in the mean were used to assess magnitudes of effects between conditions at respective time points. These were calculated using Cohen's d effect size (ES) and interpreted using thresholds of <0.25, >0.25, >0.5 and >1.0 for trivial, small, moderate, and large, respectively (Rhea, 2004). Pearson's product-moment correlation coefficient (r) was used to explore correlations between variables. No differences were observed between male and female participants for the effect of KET on the metabolic response to exercise compared to water, so male and female data are presented as combined (n = 19) data unless otherwise stated. The significance level was set at $\alpha = 0.05$ for all statistical tests.

Results

Plasma β HB, glucose and lactate. Fasting plasma β HB (KET, 0.13 ± 0.10 mM; CON, 0.12 ± 0.09 mM; ES=.05) and glucose (KET, 4.82 ± 0.46 mM; CON, 4.79 ± 0.40 mM; ES=.06) concentrations did not differ between trials (Figure 3.2). Ingestion of KET resulted in a rise in plasma β HB concentration to 0.28 ± 0.13 mM ($P < .001$) 30 min after ingestion, and remained elevated throughout exercise ($P < .001$) (Figure 3.2a). The highest plasma β HB concentration during KET was observed in the final stage of exercise at 0.44 ± 0.15 mM ($P < .001$). Plasma glucose concentration averaged 0.44 ± 0.27 mM lower 30 min after ingestion of KET compared to CON ($P = .008$; ES=.96). An inverse correlation ($r = -0.647$, $P = .004$) was observed for the change in plasma β HB and glucose concentrations at this time point. Plasma glucose concentrations remained lower at all stages throughout exercise, with effect sizes indicating small to moderate effects i.e. 30% $\dot{V}O_{2peak}$, -0.19 ± 0.36 mM, ES=.39; 40% $\dot{V}O_{2peak}$, -0.21 ± 0.43 mM, ES=.44; 50% $\dot{V}O_{2peak}$, -0.27 ± 0.40 mM, ES=.66; 60% $\dot{V}O_{2peak}$, -0.21 ± 0.39 mM, ES=.62;

70% $\dot{V}O_{2peak}$, -0.17 ± 0.54 mM, ES=.33; and 80% $\dot{V}O_{2peak}$, -0.39 ± 1.24 mM, ES=.34 (Figure 3.2b). Plasma lactate concentrations were elevated above resting values during the final two stages of exercise, but no differences between KET and CON were observed for plasma lactate concentrations at any time point (Figure 3.2c).

Cardiorespiratory responses to graded exercise and KET ingestion. All cardiorespiratory parameters exhibited main effects for exercise intensity (all $P < .001$). No differences in % $\dot{V}O_{2peak}$, $\dot{V}O_2$, $\dot{V}CO_2$, or VE were observed between conditions (Table 3.2). RER was elevated by KET ($P < .001$ for condition), and was ~ 0.03 higher for intensities up to 60% $\dot{V}O_{2peak}$ (all $P < .05$), with effect sizes indicating moderate effects at these intensities i.e. 30% $\dot{V}O_{2peak}$, 0.038 ± 0.030 , $P = 0.003$, ES=.90; 40% $\dot{V}O_{2peak}$, 0.035 ± 0.036 , $P = 0.007$, ES=.92; 50% $\dot{V}O_{2peak}$, 0.028 ± 0.031 , $P = 0.025$, ES=.81; and 60% $\dot{V}O_{2peak}$, 0.027 ± 0.037 , $P = 0.031$, ES=.78 (Figure 3.2d). The effect of KET on RER was small at 70% $\dot{V}O_{2peak}$ (0.018 ± 0.030 , $P = 0.16$, ES=.50) and 80% $\dot{V}O_{2peak}$ (0.012 ± 0.045 , $P = 0.37$, ES=.28). HR was also elevated by KET ($P = .003$ for condition), wherein HR averaged ~ 4 to 8 bpm higher during KET and effect sizes indicated small to moderate effects i.e. 30% $\dot{V}O_{2peak}$, 5.6 ± 4.5 bpm, ES=.48; 40% $\dot{V}O_{2peak}$, 8.5 ± 7.1 bpm, ES=.66; 50% $\dot{V}O_{2peak}$, 7.8 ± 7.1 bpm, ES=.55; 60% $\dot{V}O_{2peak}$, 3.9 ± 8.6 , ES=.26; 70% $\dot{V}O_{2peak}$, 4.9 ± 8.2 bpm, ES=.29; and 80% $\dot{V}O_{2peak}$, 4.4 ± 7.0 bpm, ES=.34 (Figure 3.2e). No differences in oxygen pulse, RPE, gross efficiency or delta efficiency were observed between conditions (Table 3.2).

Gastrointestinal responses. Thirteen out of nineteen (68%) participants reported symptoms of gastrointestinal distress in response to KET ingestion. These comprised of seven (37%), three (16%), two (11%), and one (5%) of the participants reporting nausea, diarrhoea, vomiting and light-headedness, respectively. These symptoms manifested in the latter stages of and immediately after the cessation of exercise. No symptoms were reported during CON

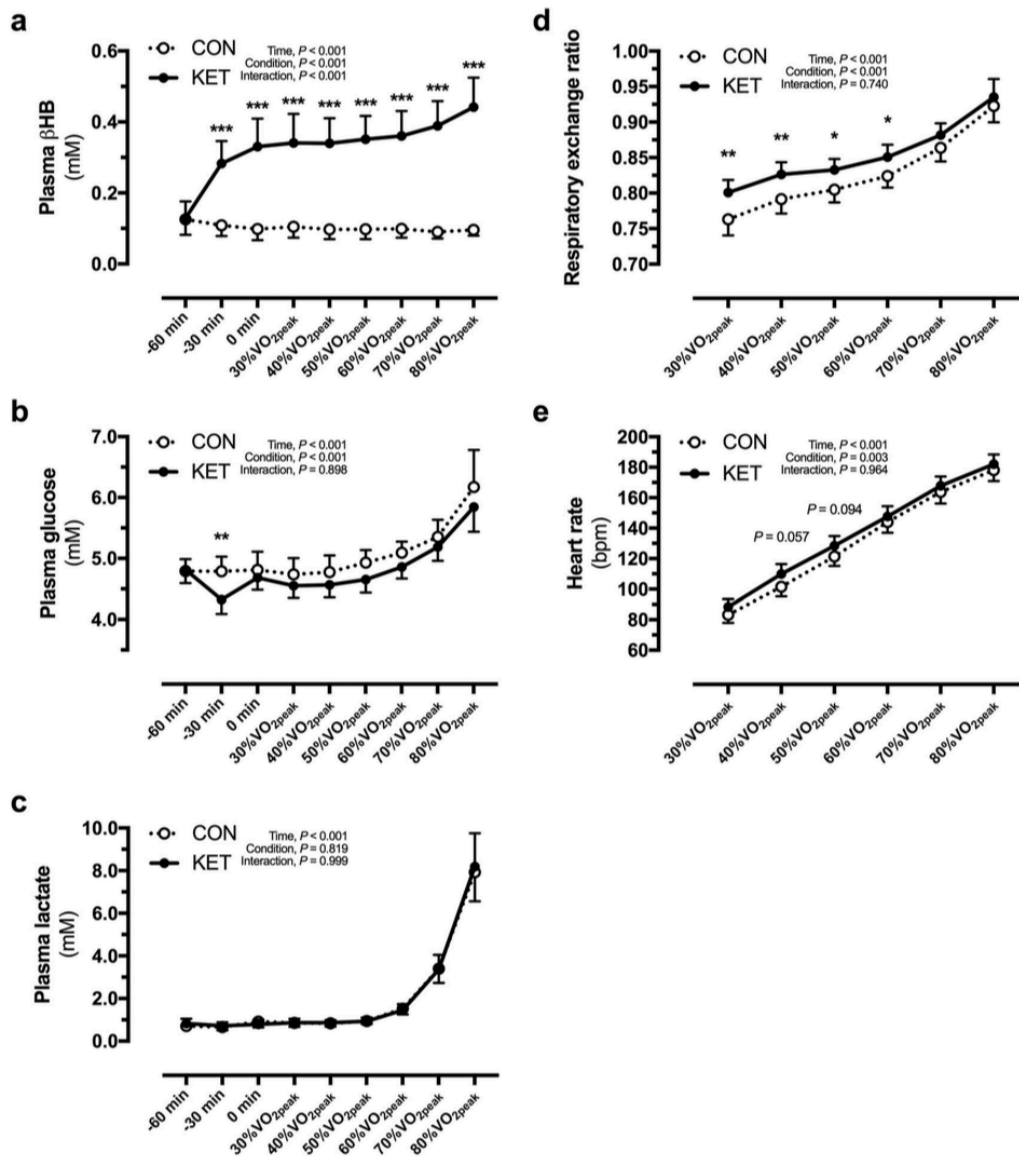


Figure 3.2 Plasma β HB (a), glucose (b) and lactate (c), respiratory exchange ratio (d), and HR (e) responses after β HB salt ingestion and water ingestion.

Data are presented as mean values, with error bars representing 95% confidence intervals. * $p < .05$ KET vs. CON; ** $p < .01$ KET vs. CON; *** $p < .001$ KET vs. CON; KET, β HB salt ingestion; CON, water ingestion; HR, heart rate; RER, respiratory exchange ratio.

Table 3.2 Cardiorespiratory responses during graded exercise in CON or KET.

	Stage 1	Stage 2	Stage 3	Stage 4	Stage 5	Stage 6
%W_{max}						
KET	16±5	28±4	40±4	52±4	64±4	75±4
CON	16±5	28±4	40±4	52±4	64±4	75±4
%$\dot{V}O_{2peak}$						
KET	29±3	39±2	50±3	61±3	74±4	85±4
CON	29±3	39±2	50±2	61±3	74±4	86±5
VE (L·min⁻¹)						
KET	30.7±5.5	40.8±5.3	52.6±5.3	66.2±7.7	89.9±10.8	127.6±16.1
CON	30.0±4.6	40.0±5.0	52.2±5.0	66.3±6.2	89.1±9.6	124.5±23.5
$\dot{V}O_2$ (L·min⁻¹)						
KET	1.42±216	1.89±151	2.42±152	2.96±210	3.55±257	4.09±299

CON	1.40±183	1.87±201	2.41±206	2.96±270	3.58±322	4.11±444
$\dot{V}CO_2$ (L·min⁻¹)						
KET	1.15±195	1.57±172	2.04±183	2.53±243	3.16±293	3.90±360
CON	1.09±163	1.49±175	1.95±179	2.44±222	3.10±305	3.80±424
RPE						
KET	7±1	9±1	11±1	13±2	15±1	18±2
CON	6±1	8±1	11±1	13±1	15±1	17±2
O₂ pulse (mL·beat⁻¹)						
KET	13±3	15±3	16±3	17±3	19±4	19±4
CON	14±4	16±4	18±4	18±4	19±4	21±5

Data are presented as mean ± SD. %Wmax, percentage of maximum power output; % $\dot{V}O_{2peak}$, percentage or peak oxygen uptake; VE, minute ventilation; $\dot{V}O_2$, rate of oxygen uptake; $\dot{V}CO_2$, rate of carbon dioxide production; RPE, rating of perceived exertion.

Discussion

The aim of the present study was to investigate the effect, if any, of acute ingestion of β HB salts on metabolic and physiological responses to a graded exercise session in trained cyclists. Ingestion of commercially-available β HB salts resulted in elevated plasma β HB concentrations (>0.3 mM) at rest and during exercise. This coincided with elevated RER (moderate effects) and HR (small to moderate effects) during submaximal exercise intensities, and a lowering of plasma glucose concentrations (small to moderate effects), compared to the ingestion of water. However, a range of other parameters including plasma lactate, rating of perceived exertion, gross efficiency, and delta efficiency were unaffected by the acute ingestion of β HB salts.

Exogenous ketone supplements, such as β HB salts, represent a novel method to increase the concentration of circulating ketone bodies without implementing restrictive dietary practices such as fasting or low carbohydrate, ketogenic diets (Cox & Clarke, 2014; Evans et al., 2017). Despite the increasing commercial availability of β HB salts, to date there is a paucity of data from humans on the metabolic response to ingestion at rest or during exercise. The sodium/potassium β HB mineral salt ingested in the present study resulted in a modest elevation (~ 0.3 to 0.4 mM) in plasma β HB concentrations. These values are similar to those observed after a 24 h fast (Balasse & Fery, 1989; Laffel, 1999), and can be considered to have produced nutritional ketosis (i.e. >0.2 mM (Robinson & Williamson, 1980)). The dosing strategy employed involved a bolus ingested both at 60 min and 15 min prior to exercise, but plasma β HB concentrations peaked during the last stage of exercise at 0.44 ± 0.15 mM. This suggests that the supplement was still being released into circulation approximately one hour after the ingestion of the second bolus, a time course consistent with several recent reports describing β HB salt ingestion at rest (Stubbs et al., 2017), and prior to exercise (O'Malley et al., 2017; Rodger et al., 2017). However, these studies reported

somewhat higher blood β HB concentrations (~ 0.6 to 1.0 mM) after ingestion at doses providing of two boluses of 11.7 g of β HB (Rodger et al., 2017), ~ 21 to 27 g of β HB (O'Malley et al., 2017), and ~ 12 g or ~ 25 g of β HB (Stubbs et al., 2017), compared to the two doses of ~ 18.5 g in the present study. However, unlike the present study, these studies measured β HB concentrations in whole blood from finger-prick sampling using handheld monitors, which are known to over-estimate blood β HB concentration ranging from 50% to three-fold relative to lab-based measures performed on serum (Guimont et al., 2015; Leckey et al., 2017).

The aim of ingestion of exogenous ketone supplements is to achieve acute nutritional ketosis (Cox & Clarke, 2014), and this is readily-achieved by the (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester (Cox et al., 2016). Ingestion of $573 \text{ mg} \cdot \text{kg}^{-1}$ body mass of that supplement raises plasma β HB concentrations to ~ 3 mM 10 min after ingestion, which rise further to ~ 6 mM within the next 60 min at rest (Cox et al., 2016). Clearly the β HB salts ingested in the present study produce plasma β HB concentrations that are ~ 10 -fold less than this. Despite the modest change in plasma β HB concentrations, the acute ingestion of β HB salts does exert some metabolic action at rest and during exercise. For instance, a $\sim 10\%$ decline in plasma glucose was observed 30 min after the ingestion of β HB salts, with an inverse correlation observed between the respective changes in plasma β HB and glucose concentrations at this time. This is consistent with the acute infusion of ketone bodies producing β HB concentrations of ~ 0.5 to 1 mM resulting in a decline in plasma glucose of $\sim 10\%$ (Mikkelsen et al., 2015; Sherwin et al., 1975), and similar results associated with β HB salt ingestion (Stubbs et al., 2017). Moreover, a slightly lower plasma glucose concentration (~ 0.2 mM; small to moderate effects) was evident throughout exercise in the present study, which confirms other recent reports (Leckey et al., 2017; O'Malley et al., 2017; Rodger et al., 2017).

Other effects observed during exercise in the present study include elevations in RER (moderate effects) and HR (small to moderate effects) during the low-to-moderate intensities of exercise. The elevation in RER may be indicative of oxidation of ketone bodies during exercise based on the stoichiometry of oxidation of AcAc. Before being oxidized as a fuel source in skeletal muscle, β HB is re-oxidized to AcAc through the action of 3-hydroxybutyrate dehydrogenase (BDH). The respiratory quotient for oxidation of AcAc is similar to that glucose at a value of 1.0 (Frayn, 1983). Therefore, a contribution of ketone oxidation to energy provision likely explains the elevation in RER during exercise after ingestion of β HB salts in the present study. The elevated RER is consistent with a recent report of prolonged submaximal exercise in trained male cyclists (Rodger et al., 2017), but the opposite of what was reported during graded exercise in recreationally-active men (O'Malley et al., 2017). Like the former study, we studied trained cyclists, so whether training status is the only explanation for the divergent findings remains to be confirmed. However, this would be consistent with our previous suggestion that oxidation of ketone bodies during exercise is likely to be greatest in trained participants with a high proportion of type I muscle fibres and/or a high oxidative capacity in skeletal muscle (Evans et al., 2017).

Calculations of arteriovenous differences of radiolabelled ketone bodies across working muscles estimate the contribution of ketone bodies to energy provision of 2% to 10% during exercise in the fasted state (Balasse et al., 1978; Fery & Balasse, 1983; Wahren et al., 1984). This contribution is unlikely to be >10% unless plasma β HB concentrations are elevated above 1 mM and exercise is being performed by trained participants (Evans et al., 2017). In well-trained participants consuming exogenous ketones as a ketone monoester, the contribution of ketone bodies to energy provision is greater i.e. 16 to 18% of total oxygen consumption (Cox et al., 2016). Therefore, although the elevation in plasma β HB concentration in the present study was modest, it is likely that this did result in an increased

contribution of ketone bodies to energy provision during exercise.

Apart from a contribution to energy provision, the principal efficacy of supplemental ketones as a performance aid is likely to be secondary effects on metabolism and alterations in fuel selection (Evans et al., 2017). For instance, acute infusion of sodium AcAc (Fery & Balasse, 1988) or sodium β HB (Lestan et al., 1994) attenuates the exercise-induced rise in plasma lactate, an effect also observed after ingestion of the aforementioned ketone monoester (Cox et al., 2016). In the latter work, reduced glycolytic flux, glycogen sparing, and increased contribution of intramuscular triglyceride to energy provision were observed during 2 h of cycling at $\sim 70\% \dot{V}O_{2\text{peak}}$. However, an attenuation of the rise in plasma lactate was not observed in the present study, or in other recent studies examining acute ingestion of β HB salts (O'Malley et al., 2017; Rodger et al., 2017). Again, this might be explained by the relatively lower increase in plasma β HB concentration produced by the β HB salts compared to the ketone monoester.

An important methodological note is that the β HB salts used in the present study provide a racemic mixture of β HB i.e. containing both the D- and L- enantiomers of β HB (also designated R- and S- respectively), whereas the β HB assay employed determines the concentration of D- β HB. D- β HB is the circulating and primary form of β HB (Tsai et al., 2006), but intracellular concentrations of L- β HB are sensitive to factors such as aging and metabolic health (Hsu et al., 2011). The D- and L- enantiomers of β HB exert divergent physiological effects on glucose metabolism in the heart (Tsai et al., 2006) and skeletal muscle (Yamada, Zhang, Westerblad, & Katz, 2010), and on longevity (Edwards et al., 2014). Recent work has demonstrated that racemic β HB ingested as β HB salts results in elevations in L- β HB concentrations of ~ 2 mM (Stubbs et al., 2017). However, it is doubtful that a change in circulating L- β HB concentration, if provided by an exogenous ketone supplement,

would have any direct effect on substrate metabolism in skeletal muscle. For instance, L- β HB is not a substrate for mitochondrial BDH and thus is not metabolized to AcAc (Scofield et al., 1982), and its physiological role is most likely in the synthesis of sterols and fatty acids in non-muscle tissues (Webber & Edmond, 1977).

The small to moderate effects observed for an elevated heart rate of 4 to 8 bpm after ingestion of β HB salts compared to water may warrant future investigation. Heart rate during exercise was not reported in previous work with β HB infusion, or ketone monoester or β HB salt ingestion, but was elevated by 25% under resting conditions after sodium β HB infusion compared to saline infusion (Gormsen et al., 2017). This indicates an effect of β HB itself rather than sodium load, but occurred at a plasma β HB concentration of \sim 4 mM in contrast to \sim 0.4 mM in the present study. Alternatively, the sodium load delivered by the β HB salts may exert some hemodynamic effects. Acute sodium ingestion can transiently elevate blood pressure (Farquhar et al., 2005), and sodium bicarbonate ingestion providing a similar dose of sodium to the present study results in an elevation in heart rate of \sim 10 bpm during moderate intensity exercise (Kahle et al., 2013).

Also notable in the present study was that thirteen out of nineteen (68%) participants reported symptoms of gastrointestinal distress after exercise in the β HB salt condition. The hypertonic nature of the β HB salts ingested likely caused an intraluminal osmotic load and water shift into the intestinal lumen resulting in osmotic diarrhoea. However, gastrointestinal distress also is a potential side effect of acute ingestion of ketone esters, with high prevalence noted after the ingestion of the ketone diester by elite cyclists (Leckey et al., 2017), and increasing incidences occurring with increasing dosages of the ketone monoester, however, this may be confounded by the increase in litres per day of the milk based meal replacement shake (Clarke et al., 2012). Clearly such issues would be deleterious to exercise performance,

and therefore require further exploration, either in terms of optimal dosing strategies, or whether repeat exposure to exogenous ketone supplements reduces these symptoms.

In conclusion, acute ingestion of a commercially-available β HB salt formulation by trained cyclists resulted in a modest increase in plasma β HB concentrations before and during graded exercise to concentrations that can be considered acute nutritional ketosis. This resulted in alterations in the metabolic and physiological response to exercise as evidenced by lowering of plasma glucose concentrations, and elevated RER and heart rate values at low-to-moderate exercise intensities compared to ingestion of water. However, no effect was observed on perceived exertion or muscular efficiency, or on plasma lactate concentrations. This is in contrast to previous work using β HB infusion or ingestion of a ketone monoester supplement, both of which achieve markedly higher plasma β HB concentrations during exercise. This suggests the likelihood that a dose-response effect exists for exogenous ketone supplements on metabolic responses and exercise performance. Given the gastrointestinal issues observed with the present β HB salts, further work is needed with other methods of increasing circulating ketone concentrations including improved free acid or mineral salt formulations, before the merit, if any, of ketone salts for performance enhancement in athletes is likely to be realised.

Chapter 4

Evans M, Egan B (2018). Intermittent running and cognitive performance after ketone ester ingestion. *Medicine & Science in Sports & Exercise* 50(11):2330-2338.

Abstract

Introduction: Ingestion of exogenous ketones alters the metabolic response to exercise and may improve exercise performance, but has not been explored in variable intensity team sport activity, or for effects on cognitive function. **Methods:** On two occasions in a double-blind, randomised crossover design, eleven male team sport athletes performed the Loughborough Intermittent Shuttle Test (Part A, 5x15 min intermittent running; Part B, shuttle run to exhaustion), with a cognitive test battery before and after. A 6.4% carbohydrate-electrolyte solution was consumed before and during exercise either alone (PLA), or with 750 mg.kg⁻¹ of a ketone ester supplement (KE). Heart rate (HR), rating of perceived exertion (RPE), and 15 m sprint times were recorded throughout, and serial venous blood samples were assayed for plasma glucose, lactate and β -hydroxybutyrate (β HB). **Results:** KE resulted in plasma β HB concentrations of ~1.5 to 2.6 mM during exercise ($P < 0.001$). Plasma glucose and lactate concentrations were lower during KE compared to PLA (moderate-to-large effect sizes). HR, RPE and 15 m sprint times did not differ between trials. Run time to exhaustion was not different ($P = 0.126$, $d = 0.45$) between PLA [(mean (95% CI); 268, (199, 336) sec) and KE (229, (178, 280) sec). Incorrect responses in a multi-tasking test increased from pre- to post-exercise in PLA [1.8 (-0.6, 4.1)] but not KE [0.0 (-1.8, 1.8)] ($P = 0.017$; $d = 0.70$). **Discussion:** Compared to carbohydrate alone, co-ingestion of a ketone ester by team sport athletes attenuated the rise in plasma lactate concentrations, but did not improve shuttle run time to exhaustion or 15 m sprint times during intermittent running. An attenuation of the decline in executive function after exhausting exercise suggests a cognitive benefit after KE ingestion.

Introduction

Ketone bodies, namely β -hydroxybutyrate (β HB), acetoacetate (AcAc) and acetone, are fatty acid metabolites whose production markedly increases in physiological states characterised by reduced glucose availability, such as starvation and ketogenic diets (Robinson and Williamson, 1980; Balasse & Fery, 1989). Ketone bodies are principally produced as a survival mechanism to provide a substrate for the brain, but are also oxidised by skeletal muscle and provide up to 10% of energy during exercise in a fasted state (Fery & Balasse, 1986). Infusion of ketone bodies exerts a range of metabolic actions, such as attenuation of hepatic glucose output, anti-lipolytic effects in adipose tissue, and in skeletal muscle, glucose ‘sparing’ and stimulation of protein synthesis (Maizels et al., 1977; Nair et al., 1988; Mikkelsen et al., 2015).

The effects of ketone bodies on substrate utilisation during exercise, and consequently athletic performance, is of increasing interest due to the development of exogenous ketone supplements, namely ketone salts and ketone esters (Clarke et al., 2012; Kesl et al., 2016; Stubbs et al., 2017). These formulations represent a method of acutely inducing nutritional ketosis (plasma β HB >0.5 mM) resulting in a variety of effects on exercise metabolism, performance and recovery (Cox et al., 2016; Holdsworth et al., 2017; O'Malley et al., 2017; Vandoorne et al., 2017; Rodger et al., 2017; Leckey et al., 2017; Evans et al., 2018; Waldman et al., 2018). Ketone salts in their presently-available racemic form produce only modest changes (<1.0 mM) in plasma β HB concentrations (Stubbs et al., 2017; O'Malley et al., 2017; Rodger et al., 2017; Evans et al., 2018; Waldman et al., 2018). While their pre-exercise ingestion can alter the metabolic response to exercise (O'Malley et al., 2017; Evans et al., 2018), there is no evidence of an ergogenic effect (O'Malley et al., 2017; Rodger et al., 2017). Alternatively, exogenous ketone supplements in the form of ketone esters produce markedly

greater changes in plasma β HB concentrations than ketone salts in humans (Stubbs et al., 2017) and rats (Kesi et al., 2016).

Two ketone esters have been reported in the recent literature: a R,S-1,3-butanediol acetoacetate diester (KDE) (Kesi et al., 2016; Leckey et al., 2017) and a (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester (KME) (Clarke et al., 2012; Stubbs et al., 2017; Cox et al., 2016). Both esters were tested in elite endurance athletes with divergent findings (Cox et al., 2016; Leckey et al., 2017). Acute ingestion of KME produced plasma β HB concentrations of \sim 3.0 mM after 20 min, and improved 30 min time-trial performance by 2% (Cox et al., 2016)). In contrast, acute ingestion of KDE was less effective at raising serum β HB concentrations (\sim 0.4 mM) and impaired 31.2 km time-trial performance by 2% (Leckey et al., 2017). Consumption of KME increased the estimated contribution of ketone bodies to fuel provision during exercise to 16-18% of total energy provision, in addition to marked metabolic effects including the attenuation of blood lactate concentrations, 'sparing' of muscle glycogen, and increased intramuscular triglyceride utilisation (Cox et al., 2016). The reduction in glycolytic flux may reflect an impairment of glycogen utilisation rather than glycogen sparing (Burke, 2015; Pinckaers et al., 2017; Evans, Cogan and Egan, 2017), a key question that requires further investigation. The former is likely to impair performance in high intensity sports that demand a high rate of ATP provision from carbohydrate sources (Burke, 2015).

Team sports such as Australian football, soccer, Gaelic games, rugby union, lacrosse and field hockey are high-intensity and intermittent in nature, consisting of repeated periods of high intensity activity (sprinting) interspersed with exercise at low-to-moderate intensities (walking, jogging) (Spencer et al., 2005; Cummins et al., 2013). Nutrition guidelines for soccer, for instance, recommend high intakes of carbohydrate prior to and during competition to maximise muscle glycogen stores with the view to enhancing performance (Burke et al.,

2006). Soccer match play results in a marked reduction in muscle glycogen, and high intensity running is attenuated in the last 15 minutes of play (Jacobs et al., 1982; Mohr et al., 2003). Therefore, nutrition strategies that could spare glycogen and maintain high intensity running in the latter parts of matches are of interest to scientists and practitioners, but research on exogenous ketone supplements to date has mostly focused on athletes from endurance sports (Cox et al., 2016; Rodger et al., 2017; Leckey et al., 2017; Evans et al., 2017). Moreover, because ketone bodies are the dominant fuel source for the brain in ketogenic states (Owen et al., 1967), there is potential for central and/or cognitive effects of exogenous ketone supplements, but to date cognitive effects were only explored in short-term feeding trials in rats (Ari et al., 2016; Kovacs et al., 2017; Murray et al., 2016). Therefore, the aim of the present study was to investigate the effects of acute ingestion of a ketone ester on metabolic responses, physical and cognitive performance in team sport athletes in response to an intermittent running protocol that simulated soccer match play. We hypothesise that ingestion of a ketone ester will have no effect on physical or cognitive performance in response to the intermittent running protocol.

Methods

Participants

Eleven male team sport athletes (mean±SD: age, 25.4±4.6 y; height 1.80±0.05 cm; body mass, 78.6±5.3 kg; $\dot{V}O_{2peak}$ 53.9±2.2 mL·kg⁻¹·min⁻¹) gave written informed consent to participate after written and verbal explanation of the procedures. Ethical approval (permit number: DCUREC2017_130) was obtained from the Dublin City University Research Ethics Committee in accordance with the Declaration of Helsinki. All participants were actively training and competing in high-intensity field-based team sports.

Experimental design

Participants visited the laboratory for exercise tests on three separate occasions over a 14 to 21 day period. During their first visit to the lab, each participant's maximal oxygen consumption ($\dot{V}O_{2peak}$) and speed at $\dot{V}O_{2peak}$ were determined using a progressive multistage shuttle run test (Yo-Yo intermittent recovery test level 1; Yo-Yo IR1) (Bangsbo et al., 2008). These data were used to determine jogging (55% $\dot{V}O_{2peak}$) and cruising (95% $\dot{V}O_{2peak}$) speeds for use during the Loughborough Intermittent Shuttle Test (LIST). The LIST is a validated simulation of the physiological and metabolic responses during soccer match play and consists of two parts: Part A comprises a fixed period of variable intensity shuttle running over 20 m; Part B consists of continuous running, alternating every 20 m between 55% and 95% $\dot{V}O_{2peak}$ until volitional fatigue (Nicholas et al., 2000). After a 15 min rest after completion of the Yo-Yo IR1, participants were familiarised with the LIST protocol by completing one 15 min block at their personalised running speeds. Cognitive tests were performed before the Yo-Yo IR1 and after familiarisation with the LIST in order to familiarise participants with the cognitive test battery.

Two main experimental trials, each comprising of the LIST (Parts A and B) with cognitive tests before and after, were performed during subsequent visits in a double-blinded, randomised cross-over design. Both experimental trials included a standardised diet for ~36 h prior to the exercise test, and were identical except for the drinks consumed before and during the LIST, namely a 6.4% carbohydrate-electrolyte solution, which was either flavoured (Symrise, UK) and acted as the control/placebo condition (PLA), or included a ketone ester (KE) (Figure 3.1). The primary outcome was endurance capacity measured by run time to exhaustion in the LIST Part B, with secondary outcomes including 15 m sprint times during the LIST Part A, heart rate (HR), rating of perceived exertion (RPE) and plasma glucose, lactate and β HB concentrations.

Incremental exercise test and familiarisation

For determination of $\dot{V}O_{2peak}$, jogging (55% $\dot{V}O_{2peak}$) and cruising (95% $\dot{V}O_{2peak}$) during the LIST, participants completed the Yo-Yo IR1. All participants completed a standardised 5 min warm-up consisting of progressive shuttle runs at 20%, 40%, 60% and 80% $\dot{V}O_{2peak}$ and dynamic stretching (high knees, heel kicks, groin bridges), followed by a period of self-selected stretching. The Yo-Yo IR1 consists of 40 m shuttle runs (2x20 m) between two sets of cones set 20 m apart. Shuttles progressively increase in speed that is dictated by an audio signal (Teambeep Software, UK). Each 40 m shuttle is separated by a 10 s rest period. The test was terminated when participants failed to complete the second 20 m shuttle on two consecutive occasions or if they reached volitional fatigue. $\dot{V}O_{2peak}$ was calculated as

$$\dot{V}O_{2peak} (\text{mL} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}) = \text{Yo-Yo IR1 distance (m)} \times 0.0084 + 36.4.$$

After a 15 min break, participants were familiarised with the LIST by performing one block of intermittent activity i.e. 15 min of Part A, were allowed 3 min of rest, and then completed the Part B run to exhaustion. Participants completed a battery of cognitive tests before the Yo-Yo IR1 and after the intermittent run to fatigue.

Cognitive test battery

The battery of cognitive tests (CANTAB Cognition, UK) was administered via a touch screen tablet lasting ~25 min. An identical test battery was administered before and after each trial. Technical issues, namely with loss of wireless internet access during test administration, resulted in the data for the cognitive test battery comprising of $n=8$ participants.

During the reaction time (RTI) task, participants select and hold a button at the bottom of the screen and five circles are presented above. In each case, a yellow dot appears in one of the five circles, and the participant must react as soon as possible, releasing the

button at the bottom of the screen, and selecting the circle in which the dot appeared. Release time (msec), reaction time (msec), and number of errors were recorded.

The multi-tasking test (MTT) is a test of executive function that measures the participant's ability to switch attention between stimuli, and ignore task-irrelevant information. White arrows are displayed on a black background, with the arrows located on either the left or right side of the screen, and pointing either to the left or to the right. A cue is displayed at the same time as the arrows, reading either "SIDE" or "DIRECTION." When the "SIDE" cue is presented, the participant is required to press a button on the left or right of the screen corresponding to the side of the screen where the arrow is presented, regardless of the direction the arrow is pointing. Conversely, when the "DIRECTION" cue is presented, the participant is required to touch a button on the left or right of the screen corresponding to the direction the arrow is pointing, regardless of which side of the screen the arrow is presented. Reaction time (msec), and number of correct and incorrect responses were recorded.

The rapid visual information processing task (RVP) is a test of sustained attention. The participant is presented with a white box in the center of the screen. Single digits ranging from 2 to 9 are presented one at a time in a pseudo-random order inside the box, appearing at a rate of 100 digits per minute. The participant is required to detect specific 3-digit sequences, including 2-4-6, 4-6-8, and 3-5-7. As soon as a target sequence is detected, the participant is required to touch a button on the screen. Response latency (msec), correct responses and false alarms were recorded.

Pre-trial preparation

All experimental trials were performed between 15:30 and 20:00, but on an individual basis, participants performed their second trial at the same time ± 1 h as their first trial. Pre-trial preparation was the same for each experimental trial. Participants were asked to abstain from alcohol for 48 h and caffeine for 24 h, and refrain from strenuous exercise training the

day prior to each trial. The day prior to experimental trials, participants were provided with a standardised diet (Gourmet Fuel, Ireland), which provided 40 kcal·kg body mass⁻¹ at a macronutrient ratio of 60% carbohydrate, 20% protein and 20% fat. On the day of experimental trials, participants consumed two meals providing 3 g·kg body mass⁻¹ of carbohydrate before arriving at the lab. The second meal was consumed 3 h before the initiation of the LIST. In addition to the energy content and macronutrient ratio, the food itself was identical for both trials. Participants performed the two experimental trials separated by either 7 or 14 days.

Experimental trials

Experimental trials were performed in a double-blinded, randomised cross-over design, and were identical except for the drinks consumed. During each trial, a bolus of a given drink was ingested 20 min prior to exercise (drink 1), and during each 3 min seated break during Part A (drinks 2 to 6) (Figure 4.1). During PLA, a 6.4% carbohydrate-electrolyte solution (Lucozade Sport, Lucozade Ribena Suntory Ltd., UK) was provided at a rate of ~1.2 g·min⁻¹ of exercise. During KE, a 6.4% carbohydrate-electrolyte solution was provided at a rate of ~1.2 g·min⁻¹ combined with 750 mg·kg⁻¹ body mass of a R-βHB (R)1,3-butanediol ketone ester (KE4, KetoneAid, USA). The ketone ester was mixed directly with the carbohydrate-electrolyte solution for ingestion in three boluses (50:25:25) i.e. at 20 min prior to exercise (drink 1), and after 30 (drink 3) and 60 min (drink 5) of exercise, respectively (Figure 4.1). During PLA, drinks 1, 3 and 5, were flavoured with a bitter additive (Symrise, UK) to taste-match with KE, and in both trials, drinks 2, 4, and 6 were provided as the unadulterated carbohydrate-electrolyte solution. All drinks were administered in opaque drinks bottles.

Upon arrival at the laboratory, participants provided a urine sample for assessment of hydration status (PalOSMO, VITECH Scientific, Japan), and then proceeded to complete the

described battery of cognitive tests. Thereafter, an indwelling catheter (21G Insyte Autoguard; Becton Dickinson, USA) was introduced into an antecubital vein for serial venous blood sampling at rest (-20 and 0 min), during each 3 min seated rest period between the 15 min blocks in Part A, and immediately after the run to exhaustion. Participants were fitted with a Bluetooth heart rate monitor (Polar V7, Polar Electro Oy, Finland) for continuous recording of HR, and then performed the standardised 10 min warm up followed by self-selected stretching. Participants then performed the LIST protocol (Part A: 5x15 min intermittent activity; Part B: run to exhaustion) (Nicholas, Nuttall and Williams, 2000). All exercise intensities were based on percentages of $\dot{V}O_{2peak}$ determined during the Yo-Yo IR1. The repeating order of activity in Part A, which occurs in a continuous manner for each 15 min block, comprises of 3 x 20 m at walking speed, 1 x maximal 15 m sprint, 4 sec recovery, 3 x 20 m jogging speed (55% $\dot{V}O_{2peak}$) and 3 x 20 m at cruising speed (95% $\dot{V}O_{2peak}$). Sprint times were measured by two sets of wireless infrared photoelectric cells (TC Timing System; Brower Timing, USA).

Part B consists of single 20 m shuttles alternating between the jogging (55% $\dot{V}O_{2peak}$) and cruising (95% $\dot{V}O_{2peak}$) speeds. The shuttle run to exhaustion, measured in sec, continued until participants were unable to complete two consecutive shuttles at cruising speeds, or until volitional fatigue. All speeds were dictated using audio software (Team Beep Software, UK). All participants received consistent encouragement during the maximal sprinting of Part A, and the run to exhaustion of Part B.

Venous blood samples were collected during the 3 min break between each 15 min block of Part A, and RPE (Borg scale) was recorded during the same time period. Incidences of gastrointestinal (GI) symptoms were recorded by interview after each trial after completion of the cognitive test battery. After completion of both experimental trials participants

completed an exit interview in which they were asked whether they could identify the KE condition, and which trial did they believe that they performed their longest run to exhaustion.

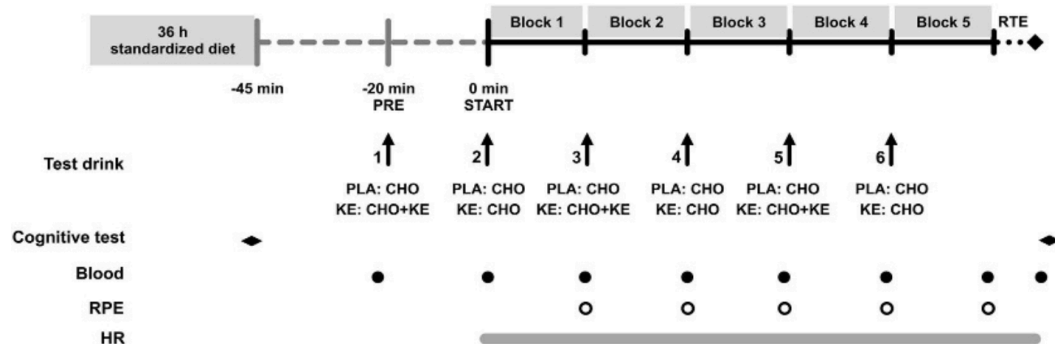


Figure 4.1 Schematic of the study protocol. CHO, carbohydrate-electrolyte solution; PLA, placebo; RTE, shuttle run to exhaustion

Blood analysis

Blood was collected in plastic tubes (2 mL) containing sodium heparin (Plus Blood Collection Tubes; Becton Dickinson, USA) for subsequent analysis of β HB). A second blood sample was collected in plastic tubes (4 mL) containing sodium fluoride (Plus Blood Collection Tubes; Becton Dickinson, USA). Samples were stored on ice before centrifugation at 3000 g for 10 min at 4 C, after which three aliquots of plasma were separated for storage at -80°C until later analysis of plasma β HB, lactate and glucose (RX Daytona, Randox Laboratories, UK: assay codes RB1007, LC2389 and GL364 respectively).

Statistical analysis

Data were evaluated using Prism 7.0 (GraphPad Software, Inc., CA, USA) and are presented as mean (lower, upper 95% confidence interval of the mean), with the exception of the participant characteristics, which are described as mean \pm SD. A paired samples t-test was used to determine differences between trials in run time to exhaustion and average HR during Part B. Two-way (time x condition) repeated measures analysis of variance (ANOVA) was used to determine differences between the two experimental trials for all other with variables

with serial measurements. When a main effect of condition, or an interaction effect between condition and time was indicated, *post-hoc* testing was performed with Bonferroni's correction with multiplicity-adjusted *P* values applied to compare KE to PLA at the respective time points. The data were tested for normality using the Shapiro-Wilk test prior to proceeding with the parametric tests described. For null hypothesis statistical testing, the significance level was set at $\alpha = 0.05$ for all tests. Apart from and independent of the outcome of the repeated measures ANOVA, standardized differences in the mean were used to assess magnitudes of effects between conditions at respective time points. These effect sizes were calculated using Cohen's *d*, and interpreted using thresholds of <0.25, >0.25, >0.5, and >1.0 for trivial, small, moderate, and large, respectively (Rhea, 2004).

Results

Pre-exercise hydration status

Hydration status, measured as urine osmolality prior to each trial, did not differ between trials [KE, 420 (259, 581) mOsm·kg⁻¹ vs. PLA, 460 (189, 732) mOsm·kg⁻¹; *P*=0.645, *d*=0.18].

Plasma β HB, glucose and lactate concentrations

Fasting plasma concentrations of β HB [KE, 0.11 (0.09, 0.13) mM; PLA, 0.11 (0.09, 0.13) mM; *P*>0.99], glucose [KE, 4.81 (4.62, 5.00) mM; PLA, 4.84 (4.57, 5.01) mM; *P*>0.99] and lactate [KE, 0.86 (0.71, 1.02) mM; PLA 0.96 (0.82, 1.11) mM; *P*>0.99] concentrations did not differ between trials (Figure 4.2). A main effect of time and condition (both *P*<0.001), and time x condition interaction effect (*P*<0.001) were observed for plasma β HB concentrations (Figure 4.2A). Ingestion of KE resulted in a rise in plasma β HB concentrations to 1.05 (0.83, 1.26) mM (*P*<0.001) by the start of exercise. Concentrations

continued to rise throughout exercise with the highest concentrations during KE observed at cessation of shuttle run to exhaustion at 2.61 (2.03, 3.10) mM ($P<0.001$).

A main effect of time ($P<0.001$) and condition ($P=0.020$) were observed for plasma glucose concentrations (Figure 4.2B). Plasma glucose concentrations were lower during KE compared to PLA at each time point with the exception of block 3, but *post-hoc* pairwise comparisons did not reveal significant differences between conditions at any time point. However, standardized differences in the mean indicated moderate effect sizes at each of these time points. Specifically, plasma glucose concentrations were lower during KE compared to PLA by 10.9% [-0.56 (-1.48, 0.35) mM; $d=0.52$] after block 1, 7.5% [-0.48 (-1.40, 0.44) mM; $d=0.56$] after block 2, 11.6% [-0.76 (-1.68, 0.15) mM; $d=0.80$] after block 4, and 8.4% [-0.55 (-1.46, 0.37) mM; $d=0.56$] after block 5. There was no difference in plasma glucose concentration at the end of the shuttle run to exhaustion [KE, 6.49 (6.01, 6.90) mM; PLA, 6.46 (5.64, 7.23) mM; $P>0.99$; $d=0.04$].

A main effect of time and condition (both $P<0.001$), and time x condition interaction effect ($P=0.009$) were observed for plasma lactate concentrations (Figure 4.2C). Plasma lactate concentrations were elevated from block 1 onwards during both conditions but were lower during KE compared to PLA at each time point. *Post-hoc* pairwise comparisons revealed significant differences between conditions at block 4 ($P=0.037$), block 5 ($P=0.042$) and the end of the shuttle run to exhaustion ($P<0.001$), and effect sizes indicating small, moderate and large effects across all time points i.e. block 1: -11.9%, -0.48 mM (-1.46, 0.50), $d=0.27$; block 2: -13.8%, -0.56 (-1.54, 0.42) mM, $d=0.33$; block 3: -20.2%, -0.73 (-1.71, 0.25) mM, $d=0.45$; block 4: -29.3%, -1.02 (-2.00, -0.04) mM, $d=0.58$; block 5: -30.1%, -1.00 (-1.98, 0.02) mM, $d=0.63$; end of the shuttle run to exhaustion: -21.5%, -1.85 (-2.83, -0.87) mM, $d=1.00$).

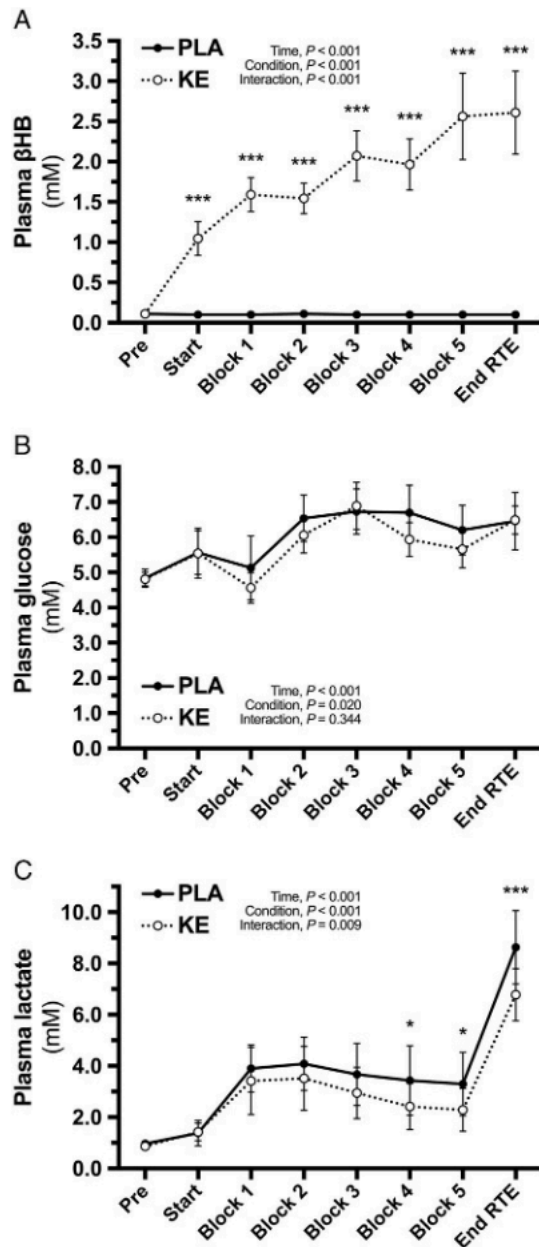


Figure 4.2 Plasma β HB (A), glucose (B), and lactate (C) responses to ketone ester and placebo ingestion.

Data are presented as mean values, with error bars representing 95% confidence intervals. * $P < 0.05$ KE vs. PLA; *** $P < 0.001$ KE vs. PLA; KE, ketone ester; PLA; placebo

For both HR and RPE, main effects of time were observed (both $P < 0.001$), but the absence of main effects of condition or time x condition interaction effects indicates that ingestion of KE did not alter the HR or RPE response during any block of Part A of the LIST protocol (Figure 4.3). However, the average HR during the shuttle run to exhaustion was

lower [-3.9 (-6.4, -1.4) bpm; $P=0.007$; $d=0.42$] during KE [170.7 (163.4, 177.9) bpm] compared to PLA [174.6 (168.3, 180.8) bpm].

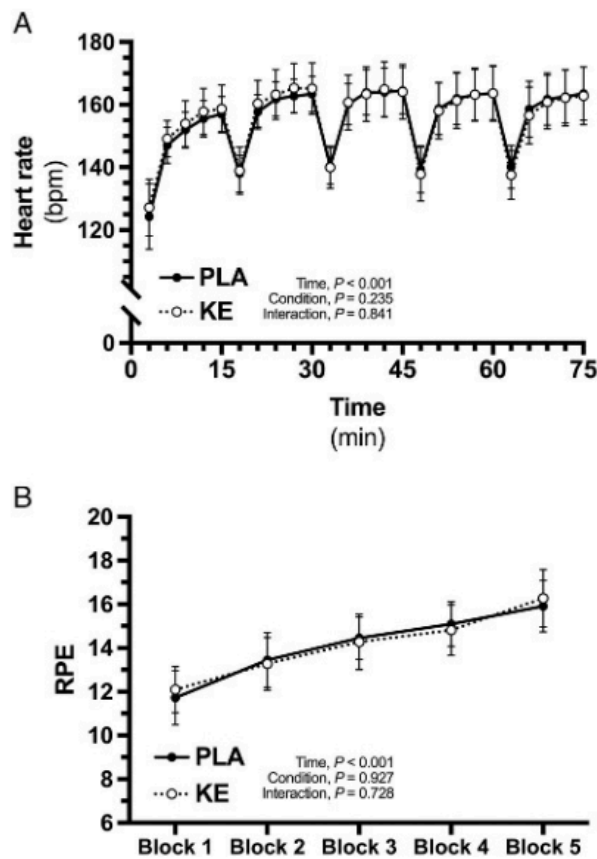


Figure 4.3 HR (A) and RPE (B) responses to ketone ester and placebo ingestion

Data are presented as mean values, with error bars representing 95% confidence intervals. KE, ketone ester; PLA; placebo.

15 m sprint times and shuttle run time to exhaustion

A main effect of time was observed for 15 m sprint times during Part A ($P < 0.001$), but no main effect of condition or time x condition interaction effect were observed (Figure 4.4A). There was no statistically significant difference in the shuttle run time to exhaustion [KE, 229 (178, 280) sec; PLA, 267 (199, 336) sec; $P=0.126$] but standardized differences in the mean indicated a small effect size for this difference [-38 (-89, 13) sec; $d=0.45$].

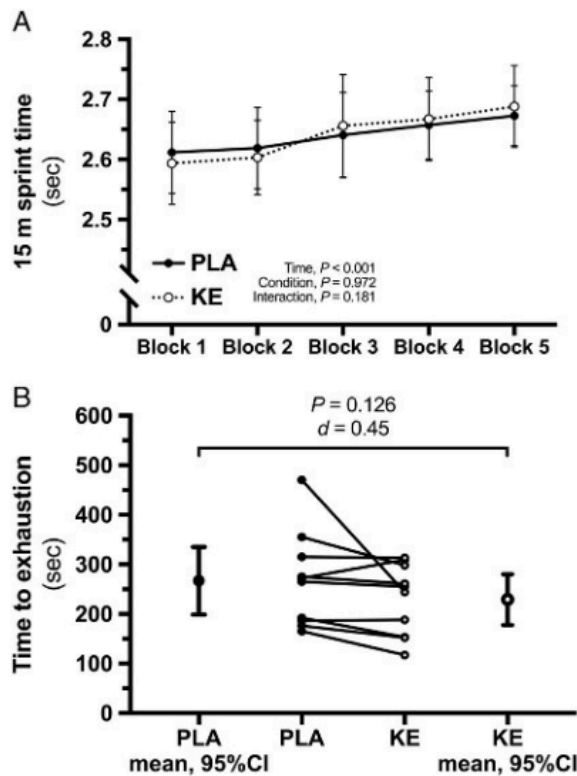


Figure 4.4 Fifteen-meter sprint times (A) and shuttle run time to exhaustion (B) during ketone ester and placebo trials

Data are presented as mean values, with error bars representing 95% CI. KE; ketone ester; PLA; placebo.

Cognitive performance

A time x condition interaction effect ($P=0.021$) was observed for the number of incorrect responses in the executive function multi-tasking test, which increased from pre- to post-exercise in PLA [1.8 (-0.6, 4.1)], but not in KE [0.0 (-1.8, 1.8)] ($P=0.017$; $d=0.70$) (Table 4.1). The absence of main effects for time or condition, and time x condition interaction effects indicates that there was no difference between conditions in either reaction time, or rapid visual information processing assessed by a sustained attention task (Table 4.1).

Gastrointestinal symptoms

Four out of eleven (36%) participants reported symptoms of GI distress during PLA and comprised of four (36%), three (27%), three (27%), one (9%) and one (9%) reports of belching, cramps, flatulence, boating and nausea, respectively. Nine out of eleven (82%) participants reported symptoms of GI distress during KE. These comprised of seven (64%), six (55%), four (36%), three (27%), three (27%) and one (9%) of the participants reporting nausea, cramps, belching, heartburn, flatulence and vomiting, respectively.

Identification of KE and best performance trials

Eight out of eleven (73%) participants correctly identified the trial in which they received KE, identifying KE by the awareness of taste and GI symptoms. However, only five (45%) of the participants correctly identified the trial in which they performed better in the Part B run to exhaustion. Only three participants (27%) stated that they believed KE ingestion improved their performance, and two out of those three participants correctly identified their KE trial and their best performance.

Table 4.1 Cognitive performance measures assessed before and after each trial

Reaction time test (RTI)									
	Release time (msec)			Reaction time (msec)			Errors		
	Pre	Post	Post-Pre	Pre	Post	Post-Pre	Pre	Post	Post-Pre
KE	393 (360, 425)	394 (369, 419)	1 (16, 18)	222 (181, 263)	218 (177, 259)	-4 (-39, 31)	0.4 (-0.1, 0.8)	0.8 (0.0, 1.5)	0.4 (-0.6, 1.4)
PLA	404 (371, 436)	397 (368, 427)	-6 (-31, 19)	225 (172, 278)	237 (205, 269)	12 (-10, 34)	0.8 (-0.2, 1.7)	0.6 (-0.6, 1.8)	-0.1 (-1.0, 0.7)
<i>d</i>			-0.28			0.45			-0.46
Multi-tasking test (MTT)									
	Response latency ^{##} (msec)			Correct responses [§]			Incorrect responses [§]		
	Pre	Post	Post-Pre	Pre	Post	Post-Pre*	Pre	Post	Post-Pre*
KE	590 (510, 669)	550 (483, 616)	-40 (-78, -2)	157.9 (156.4, 159.3)	157.9 (156.0, 159.7)	0.0 (-1.8, 1.8)	2.1 (0.7, 3.6)	2.1 (0.3, 4.0)	0.0 (-1.8, 1.8)
PLA	589 (526, 652)	543 (499, 587)	-46 (-74, -19)	157.8 (155.7, 159.8)	156.0 (152.8, 159.2) [†]	-1.8 (-4.1, 0.6)	2.3 (0.2, 4.3)	4.0 (0.8, 7.2) [†]	1.8 (-0.6, 4.1)
<i>d</i>			-0.16			-0.70			0.70
Rapid visual information processing test (RVP)									
	Response latency (ms)			Correct responses			False alarms		
	Pre	Post	Post-Pre	Pre	Post	Post-Pre	Pre	Post	Post-Pre
KE	449 (389, 509)	430 (365, 495)	-19 (-65, 26)	44.0 (35.7, 52.3)	44.6 (36.9, 52.3)	0.6 (-2.7, 4.0)	2.0 (0.9, 3.1)	2.3 (0.5, 4.0)	0.3 (-0.9, 1.4)
PLA	460 (395, 425)	446 (383, 510)	-14 (-43, 16)	45.3 (39.0, 51.5)	44.8 (37.4, 52.1)	-0.5 (-5.7, 4.7)	2.4 (1.1, 3.6)	2.5 (0.8, 4.2)	0. (-1.2, 1.4)
<i>d</i>			0.12			-0.21			-0.08

Data are presented as mean (95%CI), $n=8$. Effect size calculated as Cohen's d . Symbols are ^{##} $P < 0.01$ for main effect of Time; [§] $P < 0.05$ for Condition x Time interaction effect; [†] $P < 0.05$ for Post vs. Pre; * $P < 0.05$ for KE vs. PLA.

Discussion

The aim of the present study was to investigate the effect, if any, of the acute ingestion of a ketone ester on metabolic responses, physical and cognitive performance in team sport athletes in response to an intermittent running protocol that simulated soccer play. Compared to carbohydrate ingestion alone (PLA), ingestion of the ketone ester with carbohydrate (KE) resulted in an elevation in plasma β HB concentration to >1.5 mM after 15 min of exercise and reached ~ 2.6 mM by the end of exercise. Metabolic consequences included reductions in plasma glucose and lactate concentrations compared to PLA, but no differences in HR or RPE were observed between conditions. KE was without benefit to 15 m sprint times throughout the simulated protocol, or endurance capacity measured by shuttle run to exhaustion. However, cognitive performance in a multi-tasking executive function test was preserved with KE, but declined during PLA.

Recent reports have investigated the effect of acute ingestion of exogenous ketone supplements on physical performance in endurance athletes (Cox et al., 2016; Rodger et al., 2017; Leckey et al., 2017). Ingestion of a β HB ketone monoester (KME) increased distance covered in a 30 min cycling time-trial by $\sim 2\%$ (411 ± 162 m) (Cox et al., 2016), whereas in contrast, ingestion of an acetoacetate ketone diester (KDE) impaired performance in a 31.2 km cycling time trial by $2 \pm 1\%$ (58.2 sec) (Leckey et al., 2017). The latter effect was explained by a reduction in average power output by 3.7%, and coincided with a high prevalence of GI distress. Additionally, ingestion of a β HB salt formulation had no effect on average power output during a 4 min maximal performance cycling test (Rodger et al., 2017). A key distinction between these studies is the form of exogenous ketone supplement ingested. Acute ingestion of KME produces plasma β HB concentrations of ~ 3.0 mM after 20 min, but the KDE and racemic ketone salts result in blood β HB concentrations only in the 0.3 to 0.6 mM range (Rodger et al., 2017; Leckey et al., 2017; Evans et al., 2017). While this

concentration range constitutes acute nutritional ketosis and is sufficient to impact the metabolic response to exercise (O'Malley et al., 2017; Evans et al., 2018), performance is unlikely to be impacted unless circulating β HB concentrations exceed 1.0 mM (Evans, Cogan and Egan, 2017). In the present study, the KE condition resulted in an elevation of plasma β HB concentrations to >1.5 mM after 15 min of exercise and reached ~2.6 mM by the end of exercise, which is broadly similar to previous work (Cox et al., 2016; Stubbs et al., 2017). Our participants ingested 750 mg.kg⁻¹ body mass split across three boluses with 50% ingested 20 min prior to commencing exercise, and the remainder split into aliquots of 25% ingested at 30 and 60 min of exercise, respectively. This feeding strategy mimics the previous work with KME ingestion and exercise performance, which resulted in plasma β HB concentrations of ~2 mM 20 min after ingestion, and ranged from ~2.0 to 3.0 mM throughout 90 min of exercise (Cox et al., 2016). In contrast, our participants only achieved these concentrations 65 min after ingestion of the initial bolus of KE, but these participants ingested KE in the postprandial state as opposed to the fasted state in the KME work. Ingestion of KME in the post-prandial state can attenuate the C_{max} of blood β HB concentrations by 33%, and the 4 h β HB AUC by 27% (Stubbs et al., 2017).

Accordingly, despite the similar changes in circulating β HB concentrations to the previous investigation of performance effects using the KME (Cox et al., 2016), the KE condition in the present study was without benefit to 15 m sprint times throughout the 75 min intermittent running protocol (LIST Part A), or on shuttle run time to exhaustion (LIST Part B). The metabolic consequences of KME ingestion were recapitulated herein, namely lower plasma glucose and lactate concentrations during KE compared to PLA. A 10% reduction in plasma glucose was observed 35 min after ingestion of the initial KE bolus, and was 8 to 12% lower (moderate effects) during blocks 1, 2, 4 and 5 of Part A. The glucose-lowering effect of exogenous ketones is well-documented whether ingested alone (Cox et al., 2016; Stubbs et al.,

2017; O'Malley et al., 2017; Evans et al., 2018; Stubbs et al., 2018) or in combination with carbohydrate and/or protein (Stubbs et al., 2017; Cox et al., 2016; Vandoorne et al., 2017; Leckey et al., 2017; Myette-Côté et al., 2018). While the insulinotropic action of ketone bodies is not always observed (Nair et al., 1988; Mikkelsen et al., 2015), it can occur under certain conditions (Balasse & Fery, 1989), including when KME is ingested in the fasted state (Cox et al., 2016; Stubbs et al., 2017; Stubbs et al., 2018). However, when co-ingested with carbohydrate and/or protein, the glucose-lowering effect of exogenous ketones occurs despite similar circulating insulin concentrations in response to carbohydrate and/or protein alone at rest (Myette-Côté et al., 2018), during exercise (Cox et al., 2016), and during recovery from exercise (Vandoorne et al., 2017). A β HB-mediated glucose-lowering effect is likely a result of an attenuation of hepatic gluconeogenesis and increase in hepatic glucose uptake (Mikkelsen et al., 2015). The rise in plasma lactate concentrations was attenuated during KE compared to carbohydrate ingestion alone, consistent with previous KME work (Cox et al., 2016). An attenuation in the exercise-induced rise in plasma lactate was previously explained by a reduction in glycolytic flux, sparing of muscle glycogen during exercise and an increased contribution of ketone bodies and intramuscular triglycerides to energy provision (Cox et al., 2016). Whereas a 50% reduction in the rise in plasma lactate was observed during a 60 min pre-load at 75% W_{\max} and 30 min time-trial in trained cyclists (Cox et al., 2016), we observed a reduction ranging from ~10 to 30%. Given the lower aerobic fitness in our team sport athletes, the trained cyclists may have had a greater capacity to extract ketones from circulation and oxidise them as a substrate, resulting in a larger contribution towards total energy production and a greater reduction in glycolytic flux. This is because ketone bodies are transported across the skeletal muscle membrane by monocarboxylate transporters (MCT), which are most highly expressed in type I muscle fibres, and are increased in response to endurance exercise training (Thomas et al., 2012).

For that reason, we previously hypothesised that performance benefits of exogenous ketones are most likely to be realised in those individuals with high levels of aerobic fitness and higher proportions of type I muscle fibres and/or MCT expression (Evans, Cogan and Egan, 2017). A lower level of aerobic fitness and training status, and therefore ability to oxidise circulating ketones may be one explanation for the lack of performance benefit in the present work. That notwithstanding, our performance test was shorter (~2 to 6 min) and intermittent in nature, which may be another factor contributing to the contrasting results. Another explanation may relate to the proposed benefits of exogenous ketone supplements being via their glycogen sparing effect. Given that we employed an optimal carbohydrate-based fuelling on the day prior to and the day of each trial, our athletes may not have experienced glycogen depletion to an extent that the purported glycogen sparing would have benefited performance in Part B. In fact, standardized differences in the mean used to assess magnitudes of effects between KE and PLA indicate a small effect size for a decrement in performance with KE, so it would be remiss not to consider that the effect of exogenous ketones in this instance may have been to impair carbohydrate utilisation.

Nutrition strategies such as high fat feeding, ketogenic diets and exogenous ketone ingestion alter substrate utilisation during exercise, which generally results in lower rates of carbohydrate utilisation at moderate-to-high exercise intensities (Spencer et al., 2005; Burke, 2015, Pinckaers et al., 2017). Whether this shift in substrate utilisation reflects a sparing of muscle glycogen, which can then be utilised later in an exercise challenge, or instead reflects an impairment of muscle glycogen utilisation during such exercise intensities is a salient issue for alternative fuelling strategies. The mechanistic basis for reduced carbohydrate utilisation in the presence of exogenous ketones is proposed as an attenuation of glycolytic flux via inhibition of pyruvate dehydrogenase (PDH) and phosphofructokinase by increases in NADH:NAD⁺, acetyl-CoA:CoA or citrate. A similar mechanism is likely to contribute to

the impaired performance during moderate-to-high intensity efforts observed under high fat feeding (Havemann et al., 2006; Burke et al., 2017). The attenuation of PDH activity under such conditions (Stellingwerff et al., 2006) could be problematic for intermittent activity sports that require high intensity efforts, which rely heavily on ATP provision from glycolytic pathways, performed on a moderate intensity background. Clearly this is the nature of the exercise challenge in the present study, but future work will require direct measurement of PDH activity and glycolytic flux in muscle biopsies in order to make definitive conclusions about the effects of exogenous ketones on utilisation of muscle glycogen in this model. Conversely, we observed no benefit or decrement on 15 m sprint times performed at a rate of approximately nine sprints per 15 min block across 75 min of intermittent activity. Maximal short duration sprints rely primarily on the ATP-phosphocreatine (PCr) system and anaerobic glycolysis for energy provision, but as the number of repeated sprints increases, the contribution of both decline and the contribution of aerobic glycolysis of circulating glucose and muscle glycogen increases over time (Gaitanos et al., 1993; Parolin et al., 1999). The lower plasma lactate concentrations in Part A during KE suggests a reduction in glycolytic flux, but the reduction ultimately did not impact performance in repeated sprints of <3 sec duration.

A higher incidence of gastrointestinal (GI) symptoms occurred during KE compared to PLA, although this did not affect the HR or RPE responses during exercise. GI symptoms are a common side effect of KME and KDE ingestion and more work is needed on the dose and timing of both these supplements to mitigate this response. KME ingested as part of meal replacement milkshake drink causes a step-wise increase in symptoms with increasing dosages (Clarke et al., 2012). Furthermore, ingestion of 500 mg.kg⁻¹ body mass of KDE split in two doses caused symptoms in all participants during a cycling time-trial (Leckey et al., 2017). These symptoms are likely to be a large contributor to the performance decrement in

that study given the participants' nomination of their symptoms as a distraction or interference to performance. The incidence of GI symptoms was higher in the present study than in previous work with KME (Cox et al., 2016), but the aforementioned commencement of exercise in a fed as opposed to fasted state, or this protocol involving running as opposed to cycling exercise, may be contributing factors.

A novel finding herein is the preservation of executive function during KE compared to PLA, measured by the number of incorrect responses to a multi-tasking test. Given that team sport athletes are presented with a multitude of decisions throughout match play, interventions that preserve or improve cognitive performance could positively influence performance outcomes. The primary physiological role of ketogenesis as a survival mechanism during low carbohydrate availability is providing a substrate to the brain in the presence of diminishing blood glucose concentrations (Owen et al., 1967). Cognitive benefits and a neuroprotective role are established for exogenous ketones in non-exercise contexts (Ari et al., 2016; Murray et al., 2016; Kovacs et al., 2017; Svart et al., 2018). Notably, in a short-term (5 day) feeding study, rats supplemented daily with KME were 38% faster at completing a radial maze task, and made more correct decisions before making a mistake during the test (Murray et al., 2016). This outcome is consistent with our findings and suggests that central effects may be relevant during exercise, although other tests of cognitive function, i.e. reaction time and sustained attention tasks, were unaffected.

In conclusion, in team sport athletes acute ingestion of a ketone ester elevated plasma β HB concentrations, but did not improve performance in a shuttle run to exhaustion performed after 75 min of intermittent running. Reductions in plasma glucose and attenuated increases plasma lactate during exercise demonstrate the obvious effects of exogenous ketone ingestion on carbohydrate metabolism during exercise. However, participants experienced incidences of GI symptoms. These results underscore the need for future work to explore

possible dose-response effects while minimising any GI distress to athletes. Despite the lack of benefit to physical performance, the novel finding of preserved executive function after exhausting exercise suggests that there remains a possibility that exogenous ketones could enhance sport-specific performance of team sport athletes via other mechanisms.

Chapter 5

Presented in manuscript format as submitted for review to Medicine and Science in Sports and Exercise in February 2019 (Appendix D).

Evans M, McSwiney F, Brady A, Egan B. No benefit of ingestion of a ketone monoester supplement on 10-km running performance (currently under review)

Abstract

Introduction: Pre-exercise ingestion of exogenous ketones alters the metabolic response to exercise, but effects on exercise performance have been equivocal. **Methods:** On two occasions in a double-blind, randomized crossover design, eight endurance-trained runners performed 1 h of submaximal exercise at $\sim 65\%$ $\dot{V}O_{2\text{peak}}$ immediately followed by a 10-km self-paced TT on a motorized treadmill. An 8% carbohydrate-electrolyte solution was consumed before and during exercise, either alone (CHO+PLA), or with 573 mL kg^{-1} of a ketone monoester supplement (CHO+KME). Expired air, heart rate (HR), and rating of perceived exertion (RPE) were monitored during submaximal exercise. Serial venous blood samples were assayed for plasma glucose, lactate and β -hydroxybutyrate concentrations. **Results:** CHO+KME produced plasma β -hydroxybutyrate concentrations of ~ 1.0 to 1.3 mM during exercise ($P < 0.001$), but plasma glucose and lactate concentrations were similar during exercise in both trials. $\dot{V}O_2$, running economy, respiratory exchange ratio, HR and RPE did not differ between trials. Performance in the 10-km TT was not different ($P = 0.483$) between CHO+KME (mean = 2402 s; 95% confidence interval [CI] = 2204, 2600 s) and CHO+PLA (mean = 2422 s; 95% CI = 2217, 2628 s). Cognitive performance, measured by reaction time and a multi-tasking test, did not differ between trials. **Conclusion:** Compared with carbohydrate alone, co-ingestion of KME by endurance-trained athletes elevated plasma β -hydroxybutyrate concentrations, but did not improve 10-km running TT or cognitive performance.

Introduction

The therapeutic and performance potential of exogenous ketone supplements has been the subject of increasing interest in recent years (Egan & D'Agostino, 2016; Koutnik et al., 2018). Metabolic effects the ketone bodies (KB), namely β -hydroxybutyrate (β HB), acetoacetate (AcAc), are well-established in many organs, including attenuation of glycolysis, hepatic glucose output and adipose tissue lipolysis (Robinson & Williamson 2018), but their potential role in modulating substrate utilization has garnered attention for athletic performance (Cox et al., 2016; Evans & Egan, 2018). In the fasted state, KB provide up to 10% of energy to skeletal muscle during exercise (Balasse & Fery, 1989), and after acute ingestion of exogenous ketone supplements, this contribution can increase to 16 to 18% when circulating β HB is elevated to the 3 to 4 mM range (Cox et al., 2016). Moreover, this increase in β HB oxidation coincides with a reduction in glycolytic flux, as evidenced by an attenuation in the exercise-induced rise in plasma lactate and glycolytic intermediates, and an increase in intramuscular triglyceride utilization during exercise (Cox et al., 2016).

Circulating KB concentrations are <0.1 mM in the postprandial state, whereas hyperketonaemia is accepted as KB concentrations exceeding 0.2 mM (Robinson & Williamson 1980). Ingestion of a variety of exogenous ketone supplements can acutely produce nutritional ketosis (Cox et al., 2016; Evans et al., 2018; Stubbs et al., 2018; Stubbs et al., 2017; Rodger et al., 2017; O'Malley et al., 2017; Leckey et al., 2017; Myette-Côté et al., 2018; Waldman et al., 2017; Vandoorne et al., 2017; Holdsworth et al., 2017; Fischer et al., 2018; Evans & Egan, 2018), which has been defined as circulating KB concentrations >0.5 mM (Volek et al., 2015). The most potent of these exogenous ketone supplements is the (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester (KME). When ingested at rest in the fasted state, KME produces a dose-dependent increase in circulating β HB concentrations of up to 6 mM 20 min after the ingestion of up to 573 mg.kg⁻¹ body mass (Cox et al., 2016;

Stubbs et al., 2018). This elevation in β HB concentration coincides with decreases in plasma glucose, free fatty acids, triglycerides and ghrelin concentrations (Cox et al., 2016; Stubbs et al., 2017; Stubbs et al., 2018 Myette-Côté et al., 2018). Exercise attenuates the rise in β HB concentrations, as ingestion of 573 mg.kg^{-1} body mass KME prior to 45 min cycling at 45% and 75% peak power output (W_{max}) resulted in circulating β HB of $\sim 4.0 \text{ mM}$ and $\sim 3.0 \text{ mM}$, respectively. As a consequence of the aforementioned effects on substrate utilization, acute ingestion of KME attenuates the rise in plasma glucose and lactate concentrations during exercise, whether in an endurance cycling or intermittent running context (Cox et al., 2016; Evans & Egan, 2018).

These metabolic consequences have been proposed to explain the observation that the co-ingestion of KME in addition to a carbohydrate-based fuelling strategy improved performance in a 30 min maximum distance cycling time-trial by 2% when preceded by 1 h of submaximal ‘pre-load’ exercise (Cox et al., 2016). In contrast, high-intensity shuttle running capacity (~ 4 to 6 min) performed after 75 min of intermittent running was not improved in team sport athletes with KME co-ingestion compared to carbohydrate alone (Evans and Egan, 2018). While the former study considering a ‘sparing’ of muscle glycogen to be major factor in the performance benefit (Cox et al., 2016), the latter study speculated that the attenuation of glycolytic flux in the presence of elevated circulating β HB may have been a factor in the lack of performance benefit in that exercise model (Evans & Egan, 2018). Performance in exercise of long duration that incorporates high intensity efforts (i.e. sprint finishes, climbs) is largely dependent on carbohydrate utilization (Hawley & Leckey, 2015). Therefore, nutrition strategies that could spare muscle glycogen and maintain high intensities in the latter parts of races are of interest to scientists and practitioners (Pinckaers et al., 2017). However, if glycogen sparing occurs via an attenuation of glycolytic flux that cannot be overcome when higher intensity efforts are required, this would instead be likely to impair

performance (Hawley & Leckey, 2017). Moreover, the recent observation that acute ingestion of KME prior to intermittent exercise in team sport athletes resulted in preserved executive function as measured by a decision-making task after volitional exhaustion (Evans & Egan, 2018), remains to be confirmed in other exercise settings.

Therefore, the aim of the present study was to investigate the effects of acute ingestion of an exogenous ketone supplement in the form of a commercially-available KME on physiological responses, physical and cognitive performance in endurance-trained runners in response to 1 h submaximal exercise immediately followed by a 10-km time trial. We hypothesise ingestion of KME will have no effect on physical or cognitive performance in endurance-trained runners.

Methods

Participants

Eight trained, middle and long distance runners (M/F, 7/1; age, 33.5 ± 7.3 y; height, 1.79 ± 0.07 m; body mass, 68.8 ± 9.7 kg; body fat, 8.0 ± 4.1 %; $\dot{V}O_{2peak}$, 62.0 ± 5.6 mL kg⁻¹ min⁻¹) gave written informed consent to participate after written and verbal explanations of the procedures. Ethical approval (permit number: DCUREC2018_039) was obtained from the Dublin City University Research Ethics Committee in accordance with the Declaration of Helsinki.

Experimental design

Participants visited the laboratory for exercise tests on four separate occasions over a 21 to 28 day period, comprising one baseline, one familiarization and two main experimental trials. During their first visit to the lab, each participant's maximal rate of oxygen consumption ($\dot{V}O_{2peak}$) was determined using an incremental treadmill test to volitional exhaustion. The exercise protocol performed in the familiarization visit (visit 2) and two main

experimental trials (visits 3 and 4) comprised of a pre-load of 1 h of treadmill running at 65% $\dot{V}O_{2peak}$ followed by a self-paced 10-km time-trial (TT) performance test performed on a motorized treadmill (Figure 5.1). A battery of cognitive tests were performed before and after the exercise protocol. The main experimental trials were performed in a double-blind, placebo-controlled, randomized crossover design. Visits 2, 3 and 4 were identical in terms of the pre-test preparation (standardized physical activity and diet for 24 h prior to each visit) and the exercise protocol. The visits differed only in the drinks consumed before and during exercise, namely an 8% carbohydrate-electrolyte solution, which was co-ingested with either a flavored placebo condition (CHO+PLA), or included the (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester (CHO+KME). The primary outcome was endurance performance measured by time to complete the self-paced 10-km TT, with secondary outcomes including cognitive performance, oxygen consumption ($\dot{V}O_2$), running economy, respiratory exchange ratio (RER), heart rate (HR), rating of perceived exertion (RPE), and plasma β HB, glucose, and lactate concentrations.

Assessment of $\dot{V}O_{2peak}$ and submaximal running speeds

Body mass was measured to the nearest 0.2 kg using a calibrated digital scales (SECA, Hamburg, Germany), and height was measured to the nearest 0.01 m using a wall-mounted stadiometer (Holtain, Crymych, UK). Body fat was determined by bioelectrical impedance analysis (DC-430U Dual Frequency Analyzer; Tanita, Arlington Heights, IL USA). All exercise testing and experimental trials were conducted on a motorized treadmill (T200; COSMED, Rome Italy). Initially, for the determination of the responses in $\dot{V}O_2$ and blood lactate concentration at submaximal running speeds, participants ran for 4 min stages at a progressively increasing speeds, interspersed with a 1 min rest interval for determination of blood lactate concentrations (Lactate Pro 2; Arkray, Kyoto, Japan), RPE (Borg scale) and HR (Polar H7; Polar, Kempele Finland). The first stage was 4 km h⁻¹ slower than the average

speed corresponding to each participant's personal best time for a 10-km race. For each subsequent stage, the running speed was increased by 1 km h^{-1} until the running speed exceeded the speed corresponding to their personal best 10-km race speed. After a 10 min rest, participants began running at a speed corresponding to the last completed speed of the preceding test. Treadmill speed was increased by 2.0 km h^{-1} every 2 min for two stages, after which treadmill gradient was increased by 1.0% every 1 min until volitional fatigue. Expired air was collected and analyzed throughout these tests using the Quark RMR metabolic cart (COSMED, Rome, Italy). $\dot{V}O_2$, carbon dioxide production ($\dot{V}CO_2$), and RER were calculated from an average of breath-by-breath measurements during the last 30 s of each stage during the submaximal running stages and the assessment of $\dot{V}O_{2\text{peak}}$. $\dot{V}O_{2\text{peak}}$ was considered to have been achieved if two of the following criteria were achieved: (1) plateauing of $\dot{V}O_2$ despite increasing treadmill speed (increase of less than $2.0 \text{ mL kg}^{-1} \text{ min}^{-1}$), (2) HR within 5% of the age-predicted HR_{max} ($208 - 0.7 \times \text{age in years}$), and (3) an $RER \geq 1.10$.

Cognitive test battery

The battery of cognitive tests (CANTAB Cognition, Cambridge, UK) was administered via a touch screen tablet lasting ~10 min. An identical test battery was administered before and after each trial in visits 2, 3 and 4.

During the reaction time (RTI) test, participants select and hold a button at the bottom of the screen and five circles are presented above. In each case, a yellow dot appears in one of the five circles, and the participants must react as soon as possible, releasing the button at the bottom of the screen, and selecting the circle in which the dot appeared. Release time (msec), reaction time (msec), and number of errors were recorded.

The multi-tasking test (MTT) is a test of executive function that measures the participant's ability to switch attention between stimuli, and ignore task-irrelevant information. White arrows are displayed on a black background, with the arrows located on

either the left or right side of the screen, and pointing either to the left or to the right. A cue is displayed at the same time as the arrows, reading either “SIDE” or “DIRECTION”. When the “SIDE” cue is presented, the participant is required to press a button on the left or right of the screen corresponding to the side of the screen where the arrow is presented, regardless of the direction the arrow is pointing. Conversely, when the “DIRECTION” cue is presented, the participants are required to touch a button on the left or right of the screen corresponding to the direction the arrow is pointing, regardless of which side of the screen the arrow is presented. Reaction time (msec), and number of correct and incorrect responses were recorded.

Pre-trial preparation

All experimental trials commenced between 0730 and 1130, and were completed within a period of 4.0-4.5 h (Figure 5.1). On an individual basis, participants performed their second main experimental trial at the same time ± 1 h as their first main trial. Pre-trial preparation was the same for the familiarization visit and each main experimental trial. Participants were asked to abstain from alcohol for 48 h and caffeine for 24 h, and refrain from strenuous exercise training on the day prior to each trial. For the day prior to experimental trials, participants were provided with a prescribed meal plan that provided ~ 2800 kcal (~ 41 kcal.kg⁻¹) at a macronutrient ratio of 60% carbohydrate (~ 6.2 g kg⁻¹), 20% protein and 20% fat. Participants performed the two main experimental trials separated by either 7 or 14 days.

Main experimental trials

The protocol for the familiarization and main experimental trials were identical except for the drinks consumed before and during exercise (Figure 5.1). Participants arrived to the laboratory in a fasted state 2 h prior to the commencement of exercise, and immediately consumed a standardized breakfast of quick-cook porridge oats and cereal bars providing

~300-400 kcal (~4.4-5.8 kcal.kg⁻¹) and ~1.0 g kg⁻¹ of carbohydrate, and 500 mL of water. Participants proceeded to complete the cognitive test battery 45 min after breakfast. Thereafter, an indwelling catheter (21G Insyte Autoguard; Becton Dickinson, Franklin Lakes, NJ USA) was introduced into an antecubital vein for serial blood sampling at rest (-30 and 0 min), during submaximal exercise (20, 40 and 60 min) and immediately after the 10-km TT.

For each trial, a bolus of a given drink was ingested 30 min prior to exercise (drink 1), at 20 min intervals during the 1 h of submaximal running (drinks 2 to 4), and at the 5-km mark of the 10-km TT (drink 5) (Figure 5.1). The carbohydrate-based fuelling strategy (CHO) consisted of a 6.4% carbohydrate-electrolyte solution (Lucozade Sport; Lucozade Ribena Suntory Ltd., Uxbridge, UK) with maltodextrin (Cargill Inc, Minneapolis, MN USA) added to make an 8.0% carbohydrate-electrolyte solution that was provided at a rate of ~1.0 g min⁻¹ of exercise. During CHO+PLA, CHO was supplemented with denatonium benzoate, malic acid and arrow root extract to mimic the bitter taste and mouth-feel of the KME. During CHO+KME, CHO was supplemented with 573 mg.kg⁻¹ body mass of a (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester (HVMNTM Ketone; HVMN, Inc., San Francisco, CA USA). The commercially-available ketone ester was mixed directly with the carbohydrate-electrolyte solution for ingestion, and the 573 mg.kg⁻¹ body mass dose was divided into three boluses at a ratio 50:25:25 ingested at -30 min (drink 1), 20 min (drink 2) and 60 min (drink 4), respectively (Figure 5.1). During CHO+PLA, drinks 1, 2 and 4 were flavored with the bitter additives to taste match with CHO+KME, and in both trials, drinks 3 and 5 were provided as the unadulterated 8% carbohydrate-electrolyte solution. All drinks were administered in opaque drinks bottles.

For the exercise protocol, participants first performed a standardized 5 min warm up on the motorized treadmill (8 km h⁻¹) followed by self-selected stretching. Participants then performed 1 h of treadmill running at a speed corresponding to ~65% $\dot{V}O_{2peak}$ (Table 5.1).

Immediately after completion of the 1 h pre-load, participants completed a 10-km TT. The pre-load followed by TT protocol was modeled on the previous work demonstrating a benefit of KME on cycling TT performance (Cox et al., 2016), and has been similarly applied to treadmill running in previous studies (Russell et al., 2004; Scott et al., 2019). Prior to each TT, participants were told to complete the distance as fast as possible and they accelerated from a standing start by manually-adjusting a mounted control panel on the side of the treadmill. Participants were blinded to the speed of the treadmill and the time elapsed at all times, but were aware of the distance covered throughout the TT, including the 5-km mark when drink 5 was provided. After completing the 10-km TT, participants completed the same cognitive test battery as completed prior to exercise.

Venous blood samples were collected at 30 min prior to exercise, at 20 min intervals during submaximal exercise, and immediately after the 10-km TT. HR and RPE were recorded at 20 min intervals during submaximal exercise. Expired air was collected during the first 10 min, 25 to 30 min, and 55 to 60 min of the submaximal exercise for the monitoring of exercise intensity, and calculation of RER and running economy. Running economy is expressed as the volume of oxygen required to run 1 km relative to body mass ($\text{mL kg}^{-1} \text{ km}^{-1}$) (Barnes & Kilding, 2015). Incidences of gastrointestinal (GI) symptoms were recorded by interview after each trial. At the end of visit 4, participants completed an exit interview in which they were asked whether they could identify the CHO+KME condition, and to identify which experimental trial they believed that they performed their best TT.

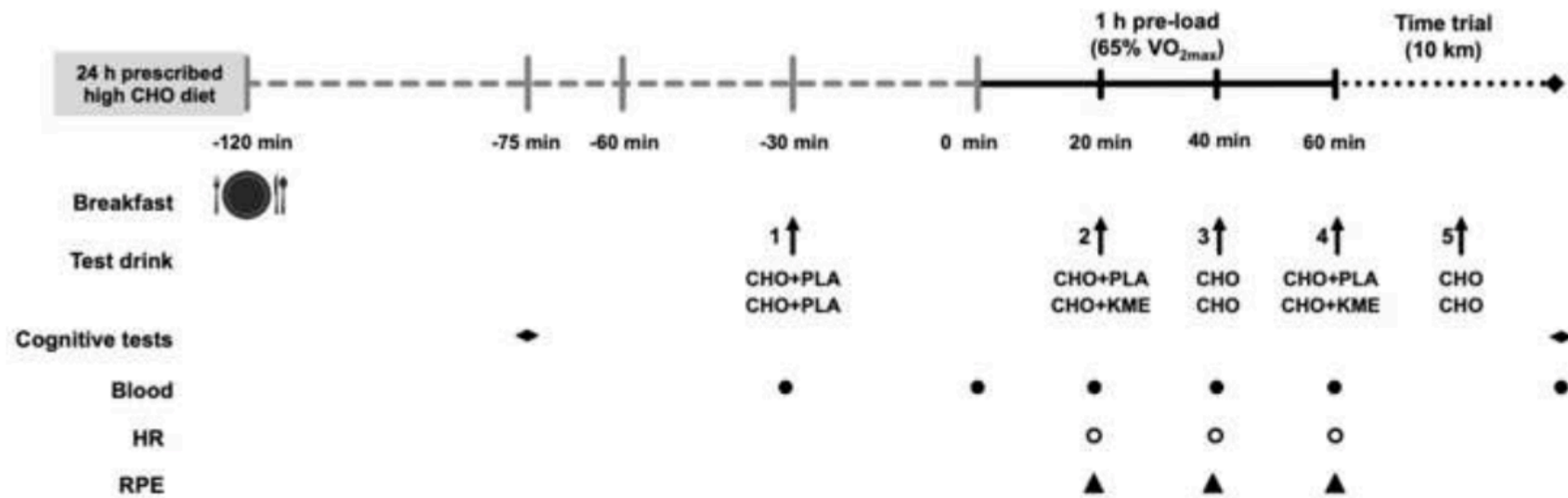


Figure 5.1 Schematic of the study protocol.

CHO, carbohydrate electrolyte solution; HR, heart rate; KME, ketone monoester; PLA, placebo; RPE, rating of perceived exertion

Blood analysis

Blood was collected in plastic tubes (2 mL) containing sodium heparin (Plus Blood Collection Tubes; Becton Dickinson, Franklin Lakes, NJ USA) for subsequent analysis of β HB. A second blood sample was collected in plastic tubes (4 mL) containing sodium fluoride (Plus Blood Collection Tubes; Becton Dickinson, Franklin Lakes, NJ USA) for subsequent analysis of glucose and lactate. All collection tubes were pre-chilled, and blood samples were stored on ice before centrifugation at 3000 g for 10 min at 4°C, after which aliquots of plasma were separated for storage at -80°C until later analysis. Plasma β HB was determined by colormetric assay as per the manufacturer's instructions (MAK041; Sigma-Aldrich, Arklow, Ireland). Plasma glucose and lactate were measured using the RX DaytonaTM chemical autoanalyser and appropriate reagents as per the manufacturer's instructions (Randox Laboratories, Crumlin, UK: assay codes GL3815 and LC3980, respectively).

Statistical analysis

Data were evaluated using Prism v8.0 (GraphPad Software, Inc., San Diego, CA USA) and are presented as mean [lower, upper 95% confidence interval (CI) of the mean], except for the participant characteristics, which are described as mean \pm SD. A paired samples t-test was used to determine differences between trials in time to complete the 10-km TT. The smallest worthwhile difference (SWD) was set at 0.2 between-subject SD, which is suggested to represent a practically-relevant change in performance in athletes (Hopkins et al., 2009). Thus, SWD corresponded to 48 sec, or 2.0%, for 10-km TT performance in this study. Two-way (time x condition) repeated measures of analysis variance (ANOVA) was used to determine differences between the two experimental trials for all variables with serial measurements. When a main effect of condition, or an interaction effect between condition and time was indicated, *post-hoc* testing was performed with Bonferroni's correction with

multiplicity-adjusted P values applied to compare CHO+KME to CHO+PLA at the respective time points. The data were tested for normality using Shapiro-Wilk test prior to proceeding with the parametric tests described. For null hypothesis statistical testing, the significance level was set at $\alpha = 0.05$ for all tests.

Results

Plasma β HB, glucose and lactate concentrations

Postprandial plasma concentrations of β HB (mean [95% CI]: CHO+KME, 0.27 [0.22-0.33] mM; CHO+PLA, 0.28 [0.14-0.43] mM), glucose (CHO+KME, 3.96 [3.22-4.70] mM; CHO+PLA, 3.70 [3.06-4.35] mM), and lactate (CHO+KME, 1.04 [0.79-1.29] mM; CHO+PLA, 1.02 [0.84-1.20] mM) did not differ between trials (all $P > 0.99$). A main effect of time and condition (both $P < 0.001$) and a time-condition interaction effect ($P < 0.001$) were observed for plasma β HB concentrations (Figure 5.2A). Ingestion of CHO+KME resulted in a rise in plasma β HB concentrations to 0.99 (0.85-1.14) mM at 0 min. β HB concentrations peaked at 1.33 (1.13-1.52) mM during submaximal exercise at 40 min, with similar concentrations observed at the cessation of the 10-km TT at 1.33 mM (0.95-1.70) mM.

A main effect of time ($P < 0.001$) and condition ($P = 0.027$) was observed for plasma glucose concentrations (Figure 5.2B). Plasma glucose concentrations were lower in CHO+KME at 0 min, i.e. 30 min after ingestion of the first bolus of either CHO+KME or CHO+PLA (CHO+KME, 3.87 [3.22-4.70] mM; CHO+PLA, 4.52 [3.91-5.13] mM; $P = 0.016$) (Figure 5.2B). Plasma glucose concentrations rose throughout submaximal exercise (Figure 5.2B) with the highest concentrations observed at cessation of the 10-km TT (CHO+KME, 6.94 [5.60-8.28] mM; CHO+PLA 7.24 [5.93-8.54] mM), with no difference between trials ($P > 0.99$).

A main effect of time ($P < 0.001$) was observed for plasma lactate concentrations, but were similar between trials at all time points (Figure 5.2C). Peak plasma lactate concentrations were observed at cessation of the 10-km TT (CHO+KME, 6.94 [4.15, 9.73] mM; CHO+PLA, 7.48 [5.46-9.51] mM; $P = 0.738$).

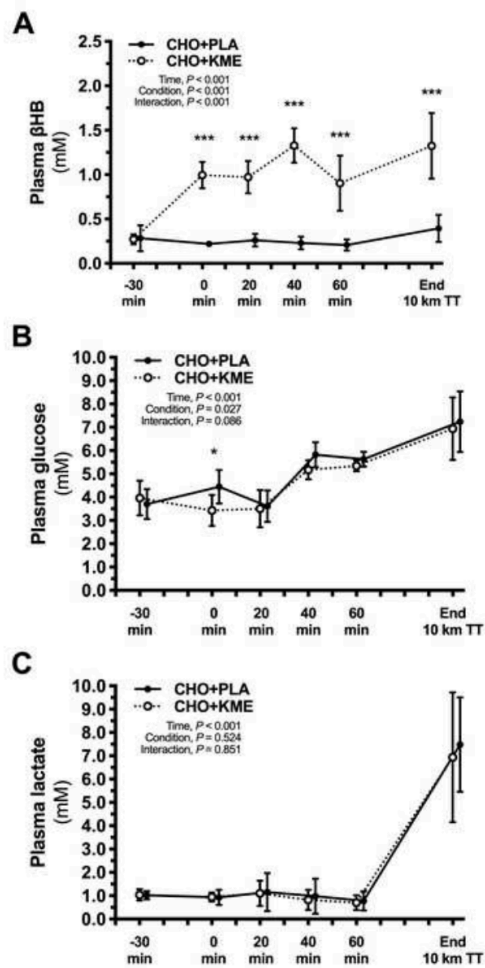


Figure 5.2 Plasma β HB (A), glucose (B), and lactate (C) concentrations during ketone monoester and placebo trials

Data are presented as mean values, with error bars representing 95% confidence intervals. * $P < 0.05$ for CHO+KME vs. CHO+PLA; *** $P < 0.001$ for CHO+KME vs. CHO+PLA. CHO+PLA, placebo; CHO+KME, ketone monoester

Submaximal exercise

Running speeds were identical between trials as per the study design. There was no difference in $\% \dot{V}O_{2\text{peak}}$, $\dot{V}O_2$, running economy, $\dot{V}CO_2$, RER, HR, and RPE between CHO+KME and CHO+PLA during the submaximal exercise period (Table 5.1). Main effects of time were observed for the decline in RER ($P < 0.001$), and the increase in RPE ($P < 0.001$) during the submaximal exercise bout (Table 5.1).

Table 5.1 Physiological responses to 1 h of treadmill running at ~65% $\dot{V}O_{2peak}$ with carbohydrate co-ingested with either placebo or a ketone monoester

	Time			<i>P</i> value
	0-10 min	10-30 min	30-60 min	
Running speed (km h ⁻¹)	12.4 (11.3, 13.5)	12.4 (11.3, 13.5)	12.3 (11.1, 13.5)	
$\dot{V}O_2$ (L min ⁻¹)				Time, <i>P</i> = 0.517
CHO+PLA	2.84 (2.52, 3.16)	2.84 (2.56, 3.12)	2.81 (2.53, 3.09)	Condition, <i>P</i> = 0.153
CHO+KME	2.78 (2.42, 3.13)	2.79 (2.42, 3.13)	2.72 (2.49, 2.95)	Interaction, <i>P</i> = 0.700
%$\dot{V}O_{2peak}$				Time, <i>P</i> = 0.576
CHO+PLA	67.0 (62.8, 71.2)	66.9 (64.5, 69.4)	66.2 (63.8, 69.4)	Condition, <i>P</i> = 0.170
CHO+KME	65.3 (60.9, 69.8)	65.8 (62.6, 69.8)	64.1 (63.2, 65.0)	Interaction, <i>P</i> = 0.710
Running economy (mL kg ⁻¹ km ⁻¹)				Time, <i>P</i> = 0.633
CHO+PLA	202 (184, 219)	203 (185, 220)	202 (185, 219)	Condition, <i>P</i> = 0.182
CHO+KME	196 (181, 212)	199 (181, 217)	196 (179, 213)	Interaction, <i>P</i> = 0.779
$\dot{V}CO_2$ (L min ⁻¹)				Time, <i>P</i> = 0.058
CHO+PLA	2.67 (2.36, 2.99)	2.60 (2.30, 2.90)	2.55 (2.26, 2.84)	Condition, <i>P</i> = 0.470
CHO+KME	2.63 (2.28, 2.98)	2.58 (2.28, 2.89)	2.50 (2.26, 2.74)	Interaction, <i>P</i> = 0.677
RER				Time, <i>P</i> < 0.001***

CHO+PLA	0.94 (0.92, 0.96)	0.91 (0.89, 0.94)	0.91 (0.88, 0.93)	Condition, $P = 0.315$
CHO+KME	0.95 (0.92, 0.97)	0.92 (0.89, 0.96)	0.92 (0.89, 0.95)	Interaction, $P = 0.478$
HR				
(bpm)				Time, $P = 0.121$
CHO+PLA	141 (133, 149)	146 (137, 155)	145 (137, 154)	Condition, $P = 0.359$
CHO+KME	140 (131, 150)	144 (134, 154)	143 (134, 152)	Interaction, $P = 0.747$
RPE				Time, $P < 0.001^{***}$
CHO+PLA	10 (9, 12)	11 (10, 13)	12 (10, 13)	Condition, $P = 0.903$
CHO+KME	10 (8, 12)	11 (9, 12)	11 (9, 13)	Interaction, $P = 0.656$

Data are presented as mean (95% CI), n = 8. *** $P < 0.001$.

10-km TT performance

There was no statistically significant difference in 10-km TT performance between trials (CHO+KME, 2402 [2204-2600] s; CHO+PLA, 2422 [2217-2628] s; $P = 0.483$) (Figure 5.3A). Compared to CHO+PLA, three participants demonstrated improvements in performance with CHO+KME that were greater than the SWD, and one participant demonstrated a decrement in performance with CHO+KME that was greater than the SWD (Figure 5.3B). The remaining participants' differences in performance between trials were less than the SWD. Running speeds for each 2 km split during the 10-km TT did not differ between trials, but did increase progressively throughout the TT (main effect of time, $P < 0.001$) (Figure 5.3C).

Cognitive performance

In the reaction time test (RTI), main effects of time ($P = 0.026$) and condition ($P = 0.026$) were observed for release time, but no interaction effect was present ($P = 0.535$), whereas an interaction effect was observed for reaction time ($P = 0.014$) (Table 5.2). In the multi-tasking test (MTT), a main effect of time was observed for response latency ($P = 0.010$), correct responses ($P = 0.049$) and incorrect responses ($P = 0.036$), but no main effects of time, or interaction effects were observed across these parameters (all $P > 0.05$) (Table 5.2). Overall, there was no difference in cognitive performance between conditions in either the RTI, or MTT assessments (Table 5.2).

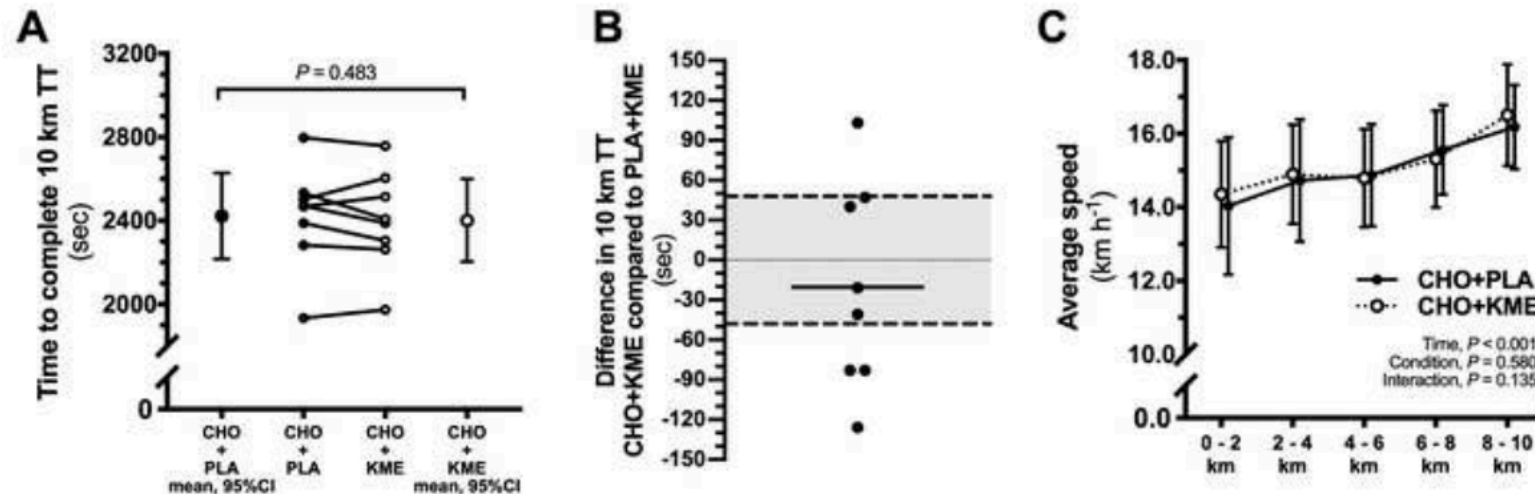


Figure 5.3 10-km time-trial performance (A), individual differences between CHO+KME compared to CHO+PLA (B), and running speeds for each 2 km split during the 10-km time-trials (C) in response to ketone monoester and placebo ingestion
 Data in (A) and (C) are presented as mean values, with error bars representing 95% confidence intervals. The shaded area in (B) represents the range for the smallest worthwhile difference in 10-km time-trial performance in this cohort. CHO+PLA, placebo; CHO+KME, ketone monoester

Table 5.2 Cognitive performance in response to ketone monoester and placebo ingestion

Reaction time test (RTI)									
	Release time ^{#,§} (msec)			Reaction time [†] (msec)			Errors		
	Pre	Post	Post-Pre	Pre	Post	Post-Pre	Pre	Post	Post-Pre
CHO+PLA	417 (373, 461)	401 (356, 446)	-16 (-35, 2)	223 (171, 276)	221 (165, 278)	-2 (-18, 14)	0.3 (-0.1, 0.6)	0.6 (0.0, 1.2)	0.4 (-0.4, 1.1)
CHO+KME	430 (383, 477)	409 (368, 450) [*]	-21 (-40, -2)	214 (176, 252)	232 (183, 282) [*]	18 (2, 34)	0.6 (-0.3, 1.5)	0.5 (0.0, 1.1)	-0.1 (-1.3, 1.1)

Multi-tasking test (MTT)									
	Response latency [#] (msec)			Correct responses [#]			Incorrect responses [#]		
	Pre	Post	Post-Pre	Pre	Post	Post-Pre	Pre	Post	Post-Pre
CHO+PLA	599 (500, 698)	561 (447, 674) ^{***}	-38 (-58, -18)	159 (157, 160)	157 (155, 159)	-2 (-4, 1)	1 (0, 3)	3 (1, 5)	2 (-1, 4)
CHO+KME	583 (513, 653)	541 (461, 622) ^{***}	-41 (-62, -21)	158 (157, 160)	157 (156, 159)	-1 (-4, 2)	2 (0, 3)	3 (1, 4)	1 (-2, 4)

Data are presented as mean (95%CI), $n=8$. Symbols are [#] $P < 0.05$ for main effect of Time; [§] $P < 0.05$ for main effect of Condition; [†] $P < 0.05$ for Condition x Time interaction effect; ^{*} $P < 0.05$ for Post vs. Pre; ^{***} $P < 0.001$ for Post vs. Pre.

Gastrointestinal symptoms

Out of 8 participants, 4 (50%) reported symptoms of GI distress during CHO+PLA and comprised 4 (50%), 3 (38%), 1 (13%), 1 (13%), and 1 (13%) incidences of belching, flatulence, reflux, urge to defecate, and diarrhoea, respectively. Out of 8 participants, 5 (63%) reported symptoms of GI distress during CHO+KME and comprised 3 (38%), 2 (25%), 1 (13%), 1 (13%), 1 (13%), and 1 (13%) incidences of belching, urge to defecate, cramps, reflux, nausea, and stitch, respectively.

Identification of CHO+KME and best performance trials

Out of 8 participants, 2 (25%) correctly identified the trial in which they received CHO+KME, identifying CHO+KME by taste and a perceived alteration of performance. Six (75%) participants declared that they could not differentiate between CHO+PLA and CHO+KME. Seven (88%) participants correctly identified the trial in which they performed their best 10-km TT.

Discussion

The present study investigated whether the acute ingestion of a commercially-available ketone monoester supplement altered metabolic responses, physical and cognitive performance in endurance-trained runners in response to 1 h of submaximal exercise immediately followed by a treadmill-based self-paced 10-km TT. Compared with placebo (CHO+PLA), ingestion of the ketone monoester (CHO+KME) elevated plasma β -hydroxybutyrate to ~ 1.0 mM at the onset of submaximal exercise, and reached ~ 1.3 mM at the end of the 10-km TT. However, CHO+KME did not alter the metabolic or

cardiorespiratory responses to exercise, or demonstrate benefit to physical or cognitive performance compared to CHO+PLA ingestion.

The present study adds to the growing body of literature investigating the effects on exercise performance of elevating ketone body concentrations by exogenous means. The term “exogenous ketone supplement” encompasses a range of different forms of supplements, with each having differential effects on the metabolic response to exercise, and exercise performance. These studies have included the acute ingestion of a (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester (KME) (Cox et al., 2016; Evans & Egan, 2018), and R,S-1,3-butanediol acetoacetate ketone diester (KDE) (Leckey et al., 2017), racemic ketone salts (KS) (Evans et al., 2017; Rodger et al., 2017; O'Malley et al., 2017; Waldman et al., 2018), and the ketogenic compound 1,3-butanediol (BD) (Scott et al., 2019; Shaw et al., 2019) prior to and/or during an exercise challenge. One of the key metabolic consequences of ingesting exogenous ketone supplements is the elevation in circulating β HB, but we speculate that exercise performance is unlikely to be affected unless β HB concentrations exceed 1.0 mM (Evans, Cogan and Egan, 2017). To date, the only supplement to consistently exceed this threshold prior to an exercise challenge is the KME supplement (Cox et al., 2016; Evans & Egan, 2018). KS and KDE elevate β HB concentrations into the 0.3 to 0.6 mM range (Evans et al., 2017; Rodger et al., 2017; Leckey et al., 2017), and ingestion of BD elevates β HB concentrations into the 0.6 to 0.8 mM range (Scott et al., 2019; Shaw et al., 2019).

Specifically focusing on KME ingestion and exercise studies, ingestion of 573 mg.kg⁻¹ body mass of KME in the fasted state elevated β HB concentrations to ~2.0 mM 20 min after ingestion where it remained throughout 1 h cycling exercise at 75%W_{max} and a subsequent 30 min TT (Cox et al., 2016). In the fed state, ingestion of 750 mg.kg⁻¹ body mass of KME elevated β HB concentrations to >1.5 mM after 15 min of exercise, and ~2.6 mM by the end of 75 min of intermittent running followed by a short duration shuttle run to exhaustion

(Evans & Egan, 2018). In contrast to this previous work, plasma β HB concentrations in the present study were elevated to ~ 1.3 mM during the exercise protocol, which is lower than previously observed at the same 573 mg.kg^{-1} body mass dose (Cox et al., 2016). This attenuated rise in plasma β HB concentrations is unsurprising given that ingestion of KME in the fasted states consistently elevates circulating β HB to >3.0 mM (Stubbs et al., 2017; Stubbs et al., 2018), whereas ingestion of KME in the postprandial state results in circulating β HB in the range from ~ 1.0 to 2.5 mM (Cox et al., 2016; Stubbs et al., 2017; Evans & Egan, 2018). For instance, ingestion of 395 mg.kg^{-1} body mass in the fasted state produces peak β HB concentrations of ~ 3.0 mM but only ~ 2.0 mM in the fed state, a 33% reduction in C_{\max} and coinciding with a 27% reduction in 4 h β HB AUC in resting participants (Stubbs et al., 2017). Given that our participants were fed a lower initial dose of KME of 287 mg.kg^{-1} body mass, that this ingestion occurred in a postprandial state, and that exercise commenced 30 min later, it is not surprising that we observed lower β HB concentrations prior to and during exercise compared to previous work.

Although the present protocol achieved acute nutritional ketosis, a benefit to endurance performance was not observed. This finding is consistent with a number of studies that have failed to find a performance benefit of exogenous ketone supplements in various exercise models (Rodger et al., 2017; O'Malley et al., 2017; Leckey et al., 2017; Waldman et al., 2017; Evans & Egan, 2018). The variety of exogenous ketones supplements used, the large range of changes in circulating β HB produced, and a lack of consistency in the nutrients co-ingested and type of exercise challenge performed, make it difficult to make broad conclusions on the efficacy of these supplements. However, only one study to date has demonstrated a performance benefit with the ingestion of KME, which when co-ingested with CHO increased the distance covered in a 30 min cycling TT by $\sim 2\%$ (411 ± 162 m), when preceded by 1 h pre-load exercise at $75\% W_{\max}$ (Cox et al., 2016). The proposed

mechanism for this improvement in performance was a shift in the contribution to energy provision from substrate utilization of carbohydrate to fat, as demonstrated by reduction in glycolytic flux resulting in a ‘sparing’ of muscle glycogen, and a concomitant increase in intramuscular triglyceride utilization during exercise (Cox et al., 2016).

The mechanistic basis whereby elevated ketones reduce carbohydrate utilization during exercise is likely an attenuation of glycolytic flux via an inhibition of pyruvate dehydrogenase and phosphofructokinase by increases in NADH:NAD⁺, acetyl-CoA:CoA, or citrate. A reduction in glycolytic flux has been proposed to explain the attenuated exercise-induced rise in plasma lactate observed in previous studies providing KME (Cox et al., 2016; Evans & Egan, 2018). This attenuation was ~50% during 60 min at 75%W_{max} and 30 min TT in trained cyclists (Cox et al., 2016), and ~10% to 30% during 75 min of intermittent running in team sport athletes (Evans & Egan, 2018). However, no differences in plasma lactate were observed between trials in the present study either during the pre-load or TT periods. The submaximal exercise intensity of ~65% $\dot{V}O_{2peak}$ employed was below lactate threshold for all participants, and therefore an intensity too low to observe an attenuation, if any, of the exercise-induced rise in plasma lactate. However, plasma β HB concentrations were elevated >1.0 mM before and at the cessation of the 10-km TT, yet no difference in plasma lactate was observed between trials.

Similarly, while a glucose-lowering effect of KME ingestion is well-documented whether ingested alone (Cox et al., 2016; Stubbs et al., 2017; Stubbs et al., 2018), or co-ingested with carbohydrate or protein (Cox et al., 2016; Stubbs et al., 2017; Myette-Côté et al., 2018; Vandoorne et al., 2017; Evans & Egan, 2018), we observed an attenuation in the rise in plasma glucose concentrations only at 30 min after ingestion of the first bolus of CHO+KME compared to CHO+PLA. This difference in plasma glucose between trials was absent during the submaximal exercise period and cessation of the 10-km TT. When effects

of KME ingestion on plasma glucose have been observed, the mechanism proposed has been an attenuation of hepatic gluconeogenesis and an increase in hepatic glucose uptake (Myette-Côté et al., 2018). Under certain conditions, elevated ketone body concentrations may have an insulintropic action (Balasse & Fery, 1989) but is not always observed (Nair et al., 1988; Mikkelsen et al., 2015). When co-ingested with carbohydrate and/or protein, the effect of exogenous ketones to attenuate postprandial glycemia occurs despite similar circulating insulin concentrations between conditions (Cox et al., 2016; Vandoorne et al., 2017; Myette-Côté et al., 2018).

We propose that the lack of differences between trials for plasma glucose and lactate in contrast to previous work suggests that the nature of the exercise challenge, or the degree of nutritional ketosis are key determinants of the metabolic effects of exogenous ketone supplements during exercise. While plasma β HB concentrations were elevated to ~ 1.3 mM at the cessation of the 10-km TT, concentrations were ~ 1.2 mM lower than observed in previous work demonstrating effects on plasma glucose and lactate during exercise (Evans and Egan, 2018; Cox et al., 2016). The lower plasma β HB concentrations are a consequence of the aforementioned particulars of the dosing and feeding strategy, and future research should be cognizant of these issues when designing study protocols.

The brain is the primary site of ketone utilization under conditions of low carbohydrate availability (Owen et al., 1967). Elevated β HB concentrations are associated with a neuroprotective role in non-exercise contexts (Ari et al., 2016; Kovacs et al., 2017; Svart et al., 2018), and short-term (5 d) feeding of a diet was supplemented with KME improved performance of rats in a radial maze task by 38%, and improved decision-making during the test (Murray et al., 2016). Moreover, in our previous work, acute ingestion of KME preserved cognitive performance, measured by the number of incorrect responses to a multi-tasking test (Evans & Egan, 2018). This test was performed at the cessation of a short

duration intermittent run to exhaustion proceeding the Loughborough Intermittent Shuttle Test (LIST), a variable intensity running protocol that mimics soccer match-play (Nicholas et al., 2000). In contrast to previous results, we observed no difference in cognitive performance with the addition of KME in the present study. The specifics of the exercise challenge may play a role in these divergent findings. The LIST is a mentally-demanding task that requires participants to be aware of current and subsequent running speeds for 75 min. Mental fatigue has a negative impact on aspects of cognitive performance, including altered attentional focus (Boksem et al., 2005) and slower and less accurate reaction times (Boksem et al., 2006), suggesting that the more mentally demanding the task the larger deficit should be evident in cognitive performance. In the present study, we observed no decline in cognitive performance in either condition. The absence of decline is important to note because in our previous work, it was a preservation of cognitive performance with KME, not an absolute improvement (Evans and Egan, 2018). These results suggest the exercise challenge presently employed was not sufficiently mentally-demanding to negatively impact reaction time or executive function, and therefore, potential benefits were unlikely to be observed.

Concerns have been raised about the practical use of exogenous ketone supplements by athletes due to the high rates of occurrence of GI distress in previous work using BD (Shaw et al., 2019), KS (Evans et al., 2017; Fischer et al., 2018), KDE (Leckey et al., 2017), and KME (Evans & Egan, 2018). However, in the present study, incidences of GI distress were similar between conditions, and this is consistent with previous work using KME (Cox et al., 2016). Typically, rates of occurrence of GI distress are higher with exogenous ketone ingestion occur at a higher rate with increasing doses (Clarke et al., 2012; Evans et al., 2017; Evans & Egan, 2018). Importantly, no participants nominated GI distress as a distraction or detriment to performance during CHO+KME trials.

In conclusion, the addition of a commercially-available ketone monoester supplement to a carbohydrate-based fuelling strategy prior to and during exercise did not improve performance in a self-paced, treadmill-based 10-km TT. Ingestion of the ketone monoester attenuated the rise in plasma glucose prior to exercise but concentrations were similar between trials thereafter, and no effect on the increase in plasma lactate concentrations during the 10-km TT was observed. Moreover, no differences between trials were observed for a range of physiological responses, and assessments of cognitive performance. Future research should evaluate different dosing strategies and exercise models to elucidate whether a threshold of plasma β HB concentration must be exceeded in order to exert performance benefits, and in which exercise contexts these benefits, if any, might be realized.

Chapter 6

Main research findings

When starting this body of work in winter of 2015, there was a dearth of published research on exogenous ketones and performance despite their use in professional sport being rumoured in that year (Abraham, 2015). Shortly thereafter, the first published report demonstrating pleiotropic effects on exercise metabolism and a meaningful ergogenic effect in elite time-trial performance emerged (Cox et al., 2016). Three years later, the area continues to grow in scope. The aim of this thesis was therefore to elucidate, if any, the metabolic effects of acute exogenous ketone ingestion prior to exercise and their ergogenic potential in other exercise contexts. The studies and their key findings are summarised as follows:

Study 1

Ingestion of β HB ketone salts (KetoCaNa, KetoSports, United States) in two boluses of 0.38 g.kg^{-1} body mass prior to exercise elevated plasma β HB concentrations to $\sim 0.4\text{-}0.5 \text{ mM}$. Ingestion of the ketone salts decreased blood glucose by $\sim 10\%$ but had no effect on blood lactate concentrations versus a water control. These results agree with other work demonstrating a decrease in blood glucose even when β HB concentrations are $< 1.0 \text{ mM}$ (O'Malley et al., 2017), but this does not always occur (Waldman et al., 2017; Rodger et al., 2017).

Study 2

Ingestion of 750 mg.kg^{-1} body mass of a β HB monoester (KME) (KE4, KetoneAid, United States) had no effect on 15 m sprint times during 75 min of soccer simulated intermittent activity. Intermittent running time to exhaustion may have been impaired with β HB monoester ingestion. A novel finding of this work was the preservation of cognitive function with β HB monoester ingestion, measured by incorrect decisions during a decision-making test but no difference was observed in reaction time or sustained attention. This work

contrasts with the seminal report on the ergogenic potential of exogenous ketones (Cox et al., 2016) where an ergogenic effect of a β HB monoester was observed in a 30-min maximum distance cycling time trial. However, the metabolic effects of elevated β HB concentrations in this study were a decrease in blood glucose and lactate concentrations, which are in agreement with other works (Cox et al., 2016; Stubbs et al., 2017; Leckey et al., 2017; Myette-Côte et al., 2018).

Study 3

Ingestion of 573 mg.kg⁻¹ body mass of the (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester (HVMN Ketone, HVMN, United States) had no effect on a 10-km self-paced time trial following a 60-min pre-load at 65% $\dot{V}O_{2peak}$. Cognitive performance improved to a similar extent in various measures in both conditions. These data contrast with previous results investigating the ergogenic potential of KME in endurance exercise (Cox et al., 2016) and in cognitive performance (Evans et al., 2018). We previously observed a preservation of cognitive performance with KME ingestion, but this is not to be confused with an improvement in cognitive performance. Both conditions in this study improved in several measures of cognitive performance in a similar manner.

Metabolic response to ketone salt ingestion during exercise

Acute ingestion of two servings ketone salts (KS) in the hour prior to exercise elevates plasma β HB concentrations to $\sim 0.44 \pm 0.27$ mM (Evans et al., 2018), while one serving KS elevates whole blood β HB concentrations between 0.5-0.8 mM (O'Malley et al., 2017; Rodger et al., 2017; Waldman et al., 2018), when measured by handheld point-of-care monitors. This modest elevation in β HB is accompanied by an $\sim 10\%$ decrease in plasma glucose during submaximal exercise (O'Malley et al., 2017; Evans et al., 2017). During submaximal exercise, plasma lactate is not affected by acute KS ingestion (Rodger et al.,

2017; O'Malley et al., 2017; Waldman et al., 2018; Evans et al., 2018). RER findings are divergent, both raising (Evans et al., 2018) and lowering (O'Malley et al., 2017) the RER, which may be accounted for by the difference in serving size and training status of participants. Doubling the recommended dosage of KS was accompanied by a high incidence of gastrointestinal distress during exercise, namely nausea, diarrhoea, vomiting and light headedness, ostensibly because of the hyperosmotic salt load delivered (Evans et al., 2018).

Higher HR values have been observed in response to ketone salt ingestion during exercise (Evans et al., 2018) and β HB infusions at rest (Gormsen et al., 2017; Svart et al., 2018), but this is not always observed during exercise (Waldman et al., 2018).

Metabolic response to ketone ester ingestion during exercise

In one of a series of experiments, ingestion of 573 mg.kg⁻¹ body mass (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester (KME) prior to 45 min cycling at 40% and 75% peak power output elevated β HB concentrations to ~4.0 mM and ~3.0 mM respectively and β HB contributed 18% and 16% towards total oxygen consumption. The exercise induced rise in plasma lactate was attenuated by ~50% during 1 h cycling at 75% W_{\max} compared to isocaloric carbohydrate ingestion (Cox et al., 2016). Similarly, in study 2 of this thesis, ingestion of KME resulted in elevated plasma β HB concentrations of ~1.0-3.0 mM, reduced plasma lactate and glucose concentrations in team sport athletes throughout a simulated soccer protocol (Evans & Egan, 2018). In study 3 of this thesis, we observed a reduction in plasma glucose 30 min after ingestion of KME, thereafter concentrations were similar between KME and placebo conditions throughout a 1 h pre-load at 65% $\dot{V}O_{2\text{peak}}$ and after a 10-km self-paced treadmill based time trial, but with no differences in plasma lactate concentrations.

The glucose-lowering effect of KME and KDE when ingested prior to exercise is well-demonstrated (Cox et al., 2016; Leckey et al., 2017; Vandoorne et al., 2017; Myette-Côté et al., 2018; Evans & Egan, 2018). This effect is most likely explained by reduced hepatic gluconeogenesis (Mikkelsen et al., 2015) and occurs independently of differences in circulating insulin between conditions where KME is present or absent with carbohydrate or carbohydrate-protein ingestion (Vandoorne et al., 2017; Myette-Côté et al., 2018; Vandoorne).

KME and KDE ingestion prior to exercise attenuates the exercise-induced increase in plasma lactate during endurance and intermittent activity (Cox et al., 2016, Leckey et al., 2017; Evans & Egan, 2018), but in study 3 on this thesis did not occur during a 60-min pre-load at 65% $\dot{V}O_{2peak}$ or after the 10 km time-trial in a population of trained endurance runners following KME ingestion. These differences are explained by the intensity of the pre-load, which was below each runner's lactate threshold and did not elevate plasma lactate in either condition. One of the main proposed benefits of exogenous ketones is a 'glycogen sparing' mechanism that will confer an advantage during periods of competition that are high intensity in nature and carbohydrate-dependent (Pinckaers et al., 2017; Evans et al., 2017). Carbohydrate utilisation is reduced during submaximal exercise with KME ingestion in the fasted state (Cox et al., 2016), and it remains to be seen whether this sparing of carbohydrate can be overcome later in exercise or whether it will impair performance if the exercise duration is long enough. When compared to the lack of effect of KS ingestion on plasma lactate, β HB concentrations between ~1.0 to 2.0 mM may be required to elicit an attenuation in plasma lactate concentrations.

Effect of acute ingestion of a β HB ketone monoester on physical performance

Ingestion of 573 mk.kg⁻¹ body mass KME improved 30-min max distance cycling time trial performance by ~2% (411 ± 162 m), following a 1 h pre-load at 75% W_{max} . The

drinks consumed in these trials were isocaloric, meaning performance improved despite receiving less carbohydrate during the KME trial (Cox et al., 2016). Since then, only two reports on the physical performance effects of acute ketone ester ingestion have been published (Leckey et al., 2017; Evans & Egan, 2018). Only the latter (study 2 of this thesis) utilised a KME and therefore is discussed in detail for contrast with the work of Cox et al. (2016). Study 3 of this thesis will add to literature in this area.

Ingestion of 750 mg.kg⁻¹ body mass KME ingested 20-min before exercise and at 30- and 60-min of soccer simulated intermittent activity divided in three aliquots (50:25:25) had no effect on 15 m sprint times during a 75 min simulated soccer protocol or subsequent run to fatigue time (KME, 229±72 s; PLA 267±96 s; p=0.126; ES=0.45 small) in team sport athletes (Evans & Egan, 2018).

The divergent findings in relation to performance between study 2 and the original KME work (Cox et al., 2016) may be explained by the training level of the participants. KME ingestion elevated plasma βHB to >1.5 mM at start of exercise and reached ~2.6 mM at the end of exercise, and reduced plasma lactate and glucose concentrations during exercise (Evans & Egan, 2018). The concentrations are similar to those achieved in endurance trained cyclists, however, KB are transported across the skeletal muscle membrane by monocarboxylate transports (MCT). MCT concentrations are highest in type 1 muscle fibres, and are increased in response to endurance training (Bonen, 2001; Thomas et al., 2012). This would suggest endurance athletes would have a higher capacity to extract KB from the blood and oxidise them as a substrate.

In trained runners, ingestion of 573 mg.kg⁻¹ body mass KME did not improve 10 km treadmill based time trial performance. KME was split into three aliquots (50:25:25) and ingested 30 min prior to and at 20 and 40 min of a 1 h pre load at 65% $\dot{V}O_{2peak}$. These findings contrast to the ergogenic effect observed in cyclists with a similar dosing and timing

strategy (Cox et al., 2016). This may be accounted for in the different endurance modalities employed in the studies (cycling vs. running). On the rationale of an ergogenic benefit due to a ‘glycogen sparing’ effect, it is possible that either the intensity of the pre-load was not sufficient to elicit an substantial depletion of muscle glycogen, or the duration of the performance test (35-45 min) was not long enough to be limited by muscle glycogen availability. However, the previous report utilised a 30 min maximal distance cycling time trial so this cannot account for the difference between studies.

Furthermore, the central tenet is that the combination of fuel sparing and improved energetic efficiency during acute nutritional ketosis confers performance benefits (Cox et al. 2016). Alterations in fuel selection during steady-state exercise have been demonstrated, which indicate reduced glycolytic flux, sparing of CHO and increased contribution of IMTG and β HB to energy provision (Cox et al., 2016). Whether this sparing of CHO, in fact, manifests as impaired CHO utilisation remains to be determined. The mechanistic basis for CHO sparing by exogenous ketones is presently proposed as inhibition of glycolytic flux via inhibition of PDH and PFK by increases in NADH:NAD^+ , acetyl-CoA:CoA ratio or citrate. In theory, this could be problematic for sports that rely heavily on contributions from glycolytic pathways, or a range of sports that are intermittent and/or require periods of high intensity ‘bursts’ on a moderate intensity background. However, the use of ‘sparing’ or ‘impairing’ of glycogen may be a misnomer, as both refer to the downregulation of carbohydrate metabolism via the same mechanism. In the current work, glycogen sparing refers to a downregulation of carbohydrate metabolism early in exercise, alongside an elevation in circulating β HB due to the ingestion of exogenous ketones, which can then be utilised later in the exercise challenge. If this downregulation of carbohydrate metabolism cannot be overcome later in exercise, resulting in an impairment of carbohydrate oxidation,

this would be termed glycogen impairing and likely lead to a performance decrement in sporting events that require a high rate of ATP turnover.

Cognitive performance after exercise after ingestion of a β HB ketone monoester

Cognitive performance after acute exogenous KS or KME ingestion is not well described in humans with only two recent reports in the published literature (Waldman et al., 2018; Evans & Egan, 2018) and study 3 will add to this area. Ingestion of 750 mg.kg⁻¹ body mass KME before and during exercise preserved executive function compared to a placebo drink, measured by the number of incorrect responses to a multi-tasking test. KME ingestion had no effect on reaction time or sustained attention, measured by latency of response and correct/incorrect responses. Cognitive tasks were performed immediately after a 75 min simulated soccer protocol and subsequent 4-6 min run to exhaustion (Evans & Egan, 2018). Following acute ingestion of one serving KS, participants completed a 5-min reaction test during their warm up and immediately following 4x15 s anaerobic Wingate sprints on a cycle ergometer. No effect on reaction time was observed after the anaerobic activity despite a higher fatigue index in the KS condition (KET: 32.3 \pm 13.9 W.s⁻¹; PLA: 29.2 \pm 12.6 W.s⁻¹), however a learning effect is noted by the authors for hits and misses on the same test (Waldman et al., 2018). These studies used different forms of exogenous ketones, dosing strategies and performance tests. These protocols may not have been long enough to cause the aforementioned reduction in whole body energy stores or dehydration to cause substantial decline in cognitive function (Tomprowski, 2003), and would seemingly agree that short exercise has little impact on cognitive function and exercise lasting <90 min exerts a selective influence on cognitive ability.

In study 3 of this thesis, in contrast to previous results in study 2, we observed no difference in cognitive performance with the addition of KME to carbohydrate compared to

placebo ingestion. The specifics of the exercise challenge may play a role in these divergent findings. The Loughborough Intermittent Shuttle Test used in study 2 is a mentally-demanding task that requires participants to be aware of current and subsequent running speeds for 75-min. Mental fatigue has a negative impact on aspects of cognitive performance, including altered attentional focus (Boksem et al., 2005) and slower and less accurate reaction times (Boksem et al., 2006), suggesting that the more mentally demanding the task the larger deficit should be evident in cognitive performance. In study 3, we observed no decline in cognitive performance in either condition. The absence of decline is important to note because in our previous work, it was a preservation of cognitive performance with KME, not an absolute improvement (Evans & Egan, 2018). These results suggest the exercise challenge employed and the degree of mental challenge are important factors when evaluating the effect on cognitive performance

Table 6. 1 Summary of the effects of ketogenic supplements on physical and cognitive performance.

Paper	Exercise challenge	Study design	Exogenous ketone supplement	Dose of ketone supplement	β HB concentrations	Effects on physical performance	Effects on cognitive performance
Cox et al., (2016)	1 h steady state cycling (75% W_{max}) 30 min maximal distance cycling time trial	β HB monoester plus carbohydrate vs isocaloric carbohydrate	(R)-3-hydroxybutyrate-(R)- 1,3-butanediol monoester	573 mg.kg ⁻¹ body mass (2:1:1)	2.0-2.5 mM	Time trial performance improved with addition of β HB monoester by approx. 2% (411 \pm 162 m)	N/A
O'Malley et al., (2017)	3x5 min cycling (30, 60, 90% VT) 150 kJ time trial (approx. 10km)	β HB salt vs flavour matched acaloric placebo	β HB salt	0.3 g.kg ⁻¹ body mass β HB salt	0.8 mM	Time trial performance impaired with β HB salt β HB salt: 711 \pm 137 s Placebo: 665 \pm 120 s	N/A
Rodger et al., (2017)	90 min cycling (80% VT ₂) 4 min maximal cycling	β HB salt vs flavour matched acaloric placebo	β HB salt	2 servings (11.7 β HB per serving)	0.6 mM	No difference in maximal or average wattage between	N/A

	performance task					conditions	
Waldman et al., (2017)	Four maximal 15 s cycling sprint trials (4 min active recovery)	β HB salt vs flavour matched acaloric placebo	β HB salt	1 serving (11.38 g β HB)	0.5 mM	Higher fatigue index during 15 s sprints in β HB salt condition. No effect of condition on mean power, peak power or total work performed.	No effect of β HB salt on average or slowest response time during a reaction time task
Leckey et al., (2017)	4x5 min warm up 31.2 km cycling time trial	AcAc diester vs taste matched placebo	R,S-1,3-butanediol acetoacetate diester.	500 mg.kg ⁻¹ body mass split in to two boluses	0.4 mM	Time to complete time trial approx. 2% longer with AcAc diester vs. placebo ingestion (58 s).	N/A
Evans and Egan (2018)	75 min intermittent running Intermittent run to exhaustion	β HB monoester plus carbohydrate vs isocaloric carbohydrate	(R)-3-hydroxybutyrate-(R)- 1,3-butanediol monoester	750 mg.kg ⁻¹ body mass	1.0 - 2.6 mM	No effect on 15 m sprint times during 75 min intermittent running My have impaired short duration intermittent	No effect of condition on tests of reaction time or sustained attention. Addition of β HB

						running to exhaustion	monoester may have preserved decision making
Evans et al., (2019)	1 h treadmill running (65% $\text{VO}_{2\text{peak}}$) Self paced treadmill based 10 km time trial	βHB monoester plus carbohydrate vs isocaloric carbohydrate	(R)-3-hydroxybutyrate-(R)- 1,3-butanediol monoester	573 mg.kg^{-1} body mass	1.6 mM	No effect on self paced 10 km time trial performance	No effect of condition on tests of reaction time or decision making

Emerging issues and future directions for research in exogenous ketone supplementation in athletic performance

Considering the recent emergence as exogenous ketone supplements as a method to induce acute nutritional ketosis and alter the metabolic response to exercise, more research on their effects on physical and cognitive performance needs to be undertaken before any statements can be made on the ergogenic potential for athletes and active adults. There remains only report of an ergogenic effect of exogenous ketones (Cox et al. 2016). Reports on various forms of exogenous ketones have reported either no effect (Rodger et al. 2017, Waldman et al. 2017, Evans and Egan. 2018, Study 3) or an impairment in physical performance (Leckey et al. 2017, O'Malley et al. 2017).

As with any ergogenic aid or nutrition strategy, optimising dosing strategies including quantity and timing will be important. Given the saturation kinetics of KB oxidation by skeletal muscle and curvilinear relationship between oxidation and plasma concentrations, it is likely that there is an optimal range for performance benefits. At present, we speculate that this exists between 1 and 3 mM β HB. As with many ergogenic acids, more is unlikely to be better and may even be deleterious given the potential for acidosis at higher KB concentrations, and aforementioned gastrointestinal distress and other side-effects sometimes observed with KS, KME and KDE, so careful consideration should be given to these issues.

KME, KDE and KS are proposed as performance-enhancing dietary supplements and are available for purchase commercially, with the exception of KDE, which is only available for research purposes. Performance-enhancing dietary supplements pose a greater challenge than other forms of dietary supplements due to the scarcity of quality research applicable to the elite athlete (Maughan et al., 2018). Future studies in exogenous ketone supplementation should be cognisant of several of the following important aspects of performance science

research to make the largest impact possible (Maughan et al., 2018; Burke & Peeling, 2018). Studies should be conducted in double-blinded design where participants are randomised to a control or experimental group or in a crossover design where participants receive both treatments. Researchers should aim for adequate sample sizes and appropriate participant characteristics to gain statistical power, should mimic conditions of real-life events, standardise pre-trial preparation across trials (exercise/diet/caffeine ingestion etc.) and environmental factors during each trial (temperature, encouragement), use a tested verified non-contaminated substance with an appropriate protocol (dosage/timing) and use a reliable and valid performance test (Maughan et al., 2018; Burke & Peeling, 2018). Studies 2 and 3 of this thesis have aimed to satisfy these recommendations.

Standardising of reporting methodologies specifically related to exogenous ketone supplementation will be important to elucidate that may benefit from supplementation and are outlined below.

βHB measurement: Verification that a potential performance-enhancing supplement was ingested and elicited a biological response should be undertaken in sports performance research (Maughan et al., 2018). Ingestion of KS, KME and KDE elevates blood/plasma βHB concentrations and should be used to measure responses in experimental and placebo conditions. Laboratory assays using reagent and colorimetric assays (Daytona; Sigma Aldrich) should be considered the gold standard for measurement of plasma βHB concentrations to standardise interpretation of results. Handheld monitors (Precision Neo, etc) measure whole blood βHB and overestimated βHB concentrations by ~1.0 mM during a 31.2 km time trial in cyclists compared to βHB measured in serum samples (Leckey et al., 2017). If using handheld monitors, overestimation should be clearly mentioned and should be reported as whole blood βHB concentrations. The use of handheld monitors raises an issue with the blinding of a performance study, as they allow immediate measurement of whole blood βHB

concentrations. In a performance study utilising crossover design comparing exogenous ketones to a carbohydrate fed condition, β HB concentrations will remain at baseline in the carbohydrate condition and will be elevated in the ketone condition, de-blinding the study. If using a handheld monitor, a researcher who is not involved in the performance testing (encouragement etc.) should analyse whole blood samples for β HB concentrations and ensure that the participant is not aware of the concentrations in order to ensure that the experiment remains blinded.

Recovery: KB have an anti-catabolic effect (Thomson & Wu, 1991), attenuating leucine oxidation, and increasing MPS by 10% when β HB is elevated to ~ 2.0 mM (Nair et al., 1988). Work utilising a practically-relevant glycogen-depleting protocol and optimal carbohydrate and protein-based recovery strategies will elucidate whether the addition of exogenous ketones confers any benefit to glycogen re-synthesis and MPS. Recovery from exercise, in terms of glycogen re-synthesis becomes more important when a short amount of time is available between bouts of exercise (i.e. multiple heats). Recently, infusion of ketone bodies produced a robust anticatabolic response under an acute inflammatory condition insult provided by lipopolysaccharide endotoxin, reducing phenylalanine efflux by 70% in fasting conditions over the course of 6 h compared to saline and free fatty acids (Thomsen et al., 2018). Maintenance of lean mass during calorie restriction and weight loss, which may be applicable to weight dependent sports or during periods of intensive training is of interest. To our knowledge, there is no work investigating the anticatabolic effects of ketone bodies in skeletal muscle in combination with amino acids. However, ingestion of 0.5 mg.kg^{-1} body mass KME provided alongside carbohydrate and protein during a 5 h recovery period from a glycogen-depleting protocol resulted in higher mTORC1 activation via downstream signalling compared to a placebo carbohydrate and protein drink (Vandoorne et al., 2017). If the addition of ketone bodies to amino acids produces a greater net muscle protein balance,

either by increasing MPS or reducing muscle protein breakdown, it may represent a novel recovery paradigm worth investigating in athletes.

Exogenous ketone dose: KME, KDE and KS dosages are reported using a variety of methods: mg.kg^{-1} body mass (Cox et al., 2016; Leckey et al., 2017; Evans et al., 2018), mM.kg^{-1} body mass (Stubbs et al., 2017), kcal.kg^{-1} (Stubbs et al., 2018) and mL.kg^{-1} (Holdsworth et al., 2017; Myette-Côté et al., 2018) and $\text{g.kg}^{-1}.\text{h}^{-1}$ (Vandoorne et al., 2017). Researchers and practitioners require a standardised reporting method to inform their own work and avoid misinterpretations. A standardised measurement of mg.kg^{-1} body mass total supplement is recommended. A salient issue with KS ingestion is the associated gastrointestinal distress experienced with higher dosages (Evans et al., 2017). Despite receiving double the manufacturers recommended dosage, βHB was elevated to $\sim 0.4\text{--}0.5$ mM during a graded exercise session, which is well below the ~ 1.0 mM threshold needed to have a meaningful contribution to fuel provision and the hypothesised level to enhance performance (Cox et al., 2016; Evans et al., 2017). Under fed conditions, KME dose >573 mg.kg^{-1} body mass may be required to elevate βHB to $1.0\text{--}3.0$ mM and elicit an ergogenic effect.

Substrate oxidation: Traditional equations used to calculate substrate oxidation assume negligible contributions from non-carbohydrate and fat sources, including KB (Frayn, 1983). KME ingestion increases the contribution of βHB to total oxygen consumptions to 16% to 18%, calculated using novel and possibly flawed methods of estimation (Cox et al., 2016). Therefore, use of traditional methods for calculating substrate oxidation are unsuitable during acute nutritional ketosis. Careful interpretation of RER during exercise is needed, as the stoichiometry of AcAc, the final step in KB oxidation, is 1.00, similar to that of carbohydrate (Frayn, 1983). A recent report on a shift from carbohydrate to fat oxidation during an incremental steady state protocol and subsequent time trial using traditional equations likely

do not reflect the true substrate oxidation proportions during the exercise challenge (O'Malley et al., 2017). Reporting of RER is likely to be all that is valid in reporting indirect calorimetry data unless the methods of Frayn (1983) are employed, but these too rely on assumptions that might not be valid in the context of acute nutritional ketosis.

Product racemity: Commercially-available KS provide a racemic mixture of β HB, i.e. containing both the D- and L- enantiomers of β HB (also designated R- and S-, respectively), whereas various β HB assays only determine D- β HB concentrations. Ingestion of a commercially available KS (KetoForce, KetoSports, United States) elevates L- β HB to ~2.0 mM (Stubbs et al., 2017). However, L- β HB is not a substrate for mitochondrial BDH a key ketolytic enzyme and is not metabolised to AcAc, and is therefore not a substrate for skeletal muscle metabolism (Schofield et al., 1982). If possible, purity analysis defined as the D- β HB content of the supplement, or L- β HB analysis should be undertaken to account for differences in product racemity.

Storage of samples: β HB is a very stable metabolite in plasma and whole blood samples, allowing for measurement up to 7 days after sample collection when stored at 4 °C, or for longer periods when stored at -80 °C (Custer et al., 2001) Degradation of AcAc in plasma samples was thought to be rapid, with guidelines suggesting measurement must take within 24 h of sample collection or stored for no longer than 3 days at -20 °C. This was based on data showing 80% and 30% of AcAc was lost from plasma samples after 3 d storage at room temperature and -20 °C, respectively (Price et al., 1977; Stubbs et al., 2018). However, degradation of AcAc in plasma is reduced when stored at -80 °C, with only 14% lost over 40 days of storage (Fritzsche et al., 2001). Moreover, AcAc is stable when de-proteinised with perchloric acid and stored at -80 °C for 60 days, and is more stable in plasma than whole blood samples. De-proteinisation ensures removal of 3-hydroxybutyrate dehydrogenase, meaning that AcAc is not being reduced to β HB (McNeil et al., 2014). A comparison

between non de-proteinised and de-proteinised samples was not undertaken in this latest study (McNeil et al., 2014), but no difference in AcAc concentrations were noted when samples were left proteinised and non-de-proteinised prior to analysis in a previous study (Galán et al., 2001). Analysis of AcAc concentrations should be undertaken in studies using ketogenic supplements as AcAc can be reduced to β HB in collected samples. Samples for AcAc analysis should be immediately stored and measurement validity may be improved by de-proteinisation with perchloric acid.

Concluding remarks

In conclusion, although data are preliminary and somewhat conflicting, acute nutritional ketosis achieved by ingestion of exogenous ketone supplements has certainly the potential to alter fuel selection and the metabolic response during exercise, but the evidence to confer performance benefits remains sparse. Benefits to performance, if any, are most likely to be observed in trained individuals who have a greater capacity to uptake and oxidise KB during exercise because of their aerobic training base and the consequent ketolytic adaptations in skeletal muscle. Additionally, a strong physiological basis, albeit presently circumstantial, exists that suggests potential benefits for supporting training adaptation and recovery. While much work remains to be performed, particularly in relation to sport-specific strategies, this promises to be an exciting topic for scientists, practitioners and athletes alike for the coming years.

References

- Abraham, R. (2015). Ketones: Controversial new energy drink could be next big thing in cycling. *Cycling Weekly*.
- Adams, J.H and Koeslag, J.H. (1988). Carbohydrate homeostasis and post-exercise ketosis in trained and untrained rats. *J Physiol* 407:453-461.
- Adams, J.H and Koeslag, J.H. (1989). Glycogen metabolism and post-exercise ketosis in carbohydrate-restricted trained and untrained rats. *Q J Exp Physiol* 74:27-34.
- Ari, C., Kovacs Z, Juhasz, G., Murdun, C., Goldhagen, C.R, Koutnik, A.P, Poff, A.M, Kesl, S.L and D'Agostino, D.P. (2016) Exogenous Ketone Supplements Reduce Anxiety-Related Behavior in Sprague-Dawley and Wistar Albino Glaxo/Rijswijk Rats. *Front Mol Neurosci*. 9:137.
- Askew, E.W., Dohm, G.L. and Huston RL. (1975). Fatty acid and ketone body metabolism in the rat: response to diet and exercise. *J Nutr* 105:1422-1432.
- Bach, A.C. and Babayan, V.K. (1982) Medium-chain triglycerides: an update. *Am J Clin Nutr*. 36(5):950-62.
- Balasse, E.O. (1979) Kinetics of ketone body metabolism in fasting humans. *Metabolism*. 28(1):41-50.
- Balasse, E.O. and Ooms, H.A. (1968) Changes in the concentrations of glucose, free fatty acids, insulin and ketone bodies in the blood during sodium beta-hydroxybutyrate infusions in man. *Diabetologia*. 4(3):133-5.
- Balasse, E.O. and Neef, M.A. (1975) Inhibition of ketogenesis by ketone bodies in fasting humans. *Metabolism* 24(9):999-1007.
- Balasse, E.O. and Fery, F. (1989). Ketone body production and disposal: effects of fasting, diabetes, and exercise. *Diabetes Metab Rev* 5:247-270.
- Balasse, E.O., Fery, F. and Neef MA. (1978). Changes induced by exercise in rates of turnover and oxidation of ketone bodies in fasting man. *J Appl Physiol Respir Environ Exerc Physiol* 44:5-11.
- Baldwin, K.M., Fitts, R.H., Booth, F.W., Winder, W.W. and Holloszy JO. (1975). Depletion of muscle and liver glycogen during exercise. Protective effect of training. *Pflugers Arch* 354:203-212.

- Bangsbo, J., Iaia, F.M. and Krstrup, P. The Yo-Yo intermittent recovery test : a useful tool for evaluation of physical performance in intermittent sports. *Sports Med.* 2008;38(1):37-51.
- Barnes, K.R. and Kilding, A.E. (2015) Running economy: measurement, norms, and determining factors. *Sports Med Open.* 1(1):8.
- Bartlett, J.D., Hawley, J.A. and Morton, J.P. (2015). Carbohydrate availability and exercise training adaptation: too much of a good thing? *Eur J Sport Sci* 15:3-12.
- Beattie, M.A. and Winder, W.W. (1984). Mechanism of training-induced attenuation of postexercise ketosis. *Am J Physiol* 247:R780-785.
- Beattie, M.A. and Winder, W.W. (1985). Attenuation of postexercise ketosis in fasted endurance-trained rats. *Am J Physiol* 248:R63-67.
- Bergström, J., Hermansen, L., Hultman, E. and Saltin, B. (1967) Diet, Muscle Glycogen and Physical Performance. *Acta Physiologica Scandinavica.* 71(2-3);150-160.
- Beylot, M., Beaufrere, B., Normand, S., Riou, J.P., Cohen, R. and Mornex, R. (1986) Determination of human ketone body kinetics using stable-isotope labelled tracers. *Diabetologia.* 29(2):90-6.
- Birkhahn, R.H., McMenamy, R.H. and Border, J.R. (1977) Intravenous feeding of the rat with short chain fatty acid esters. I. Glycerol monobutyrate. *Am J Clin Nutr.* 30(12): 2078-82.
- Birkhahn, R.H. (1983) Experience with alternative fuels, in Johnston, I.D.A. (ed.) *Advances in Clinical Nutrition.* USA: MTP Press: 325-337.
- Birkhahn, R.H., McCombs, C., Clemens, R. and Hubbs, J. (1997) Potential of the monoglyceride and triglyceride of dl-3-hydroxybutyrate for parenteral nutrition: Synthesis and preliminary biological testing in the rat, *Nutrition*, 13(3): 213-219.
- Boksem, M.A., Meijman, T.F. and Lorist, M.M. (2005) Effects of mental fatigue on attention: an ERP study. *Brain Res Cogn Brain Res.* 25(1):107-16.
- Boksem, M.A., Meijman, T.F. and Lorist, M.M. (2006) Mental fatigue, motivation and action monitoring. *Biol Psychol.* 72(2):123-32.

- Bonen, A. (2001) The expression of lactate transporters (MCT1 and MCT4) in heart and muscle. *Eur J Appl Physiol* 86:6-11.
- Bosch, A.N., Weltan, S.M., Dennis, S.C. and Noakes, T.D. (1996). Fuel substrate kinetics of carbohydrate loading differs from that of carbohydrate ingestion during prolonged exercise. *Metabolism* 45:415-423.
- Brunengraber, H. (1997) Potential of ketone body esters for parenteral and oral nutrition, *Nutrition*, 13(3): 233-235.
- Burke, L.M. (2015). Re-Examining High-Fat Diets for Sports Performance: Did We Call the 'Nail in the Coffin' Too Soon? *Sports Med* 45 Suppl 1: S33-49.
- Burke, L.M., Loucks, A.B. and Broad, N. (2006) Energy and carbohydrate for training and recovery. *J Sports Sci.* 24(7):675-85.
- Burke, L.M., Ross, M.L., Garvican-Lewis, L.A., Welvaert, M., Heikura, I.A., Forbes, S.G., Mirtschin, J.G., Cato, L.E., Strobel, N., Sharma, A.P. and Hawley, A.P. (2017) Low carbohydrate, high fat diet impairs exercise economy and negates the performance benefit from intensified training in elite race walkers. *J Physiol.* 595(9):2785-807.
- Burke, L.M. and Peeling, P. (2018) Methodologies for Investigating Performance Changes With Supplement Use, *Int J Sport Nutr Exerc Metab.* 28(2):159-169.
- Carlin, J.I., Olson, E.B.Jr., Peters, H.A. and Reddan, W.G. (1987). The effects of post-exercise glucose and alanine ingestion on plasma carnitine and ketosis in humans. *J Physiol* 390: 295-303.
- Cerioti, F., Kaczmarek, E., Guerra, E., Mastrantonio, F., Lucarelli, F., Valgimigli, F. and Mosca, A. (2014) Comparative Performance Assessment of Point-of-Care Testing Devices for Measuring Glucose and Ketones at the Patient Bedside. *J Diabetes Sci Technol.* 9(2):268-277.
- Cermak, N.M. and van Loon, L.J. (2013). The use of carbohydrates during exercise as an ergogenic aid. *Sports Med* 43:1139-1155.
- Chari, A. and Wertheimer, E. (1953) Effect of Acetoacetate on Glycogen Synthesis and Glucose Utilization in the Isolated Rat Diaphragm, *Nature.* 171:44.
- Cian, C., Barraud, P.A., Melin, B. and Raphel, C. (2001) Effects of fluid ingestion on cognitive function after heat stress or exercise-induced dehydration, *Int J Psychophysiol.* 42(3): 243-51.

- Cian, C., Koulmann, N., Barraud, P.A., Raphel, C., Jimenez, C. and Melin, B. (2000) Influence of Variations in Body Hydration on Cognitive Function. *Int J of Psychophysiol.* 14(1):29-36.
- Ciarlone, S.L., Grieco, J.C., D'Agostino, D.P. and Weeber, E.J. (2016) Ketone ester supplementation attenuates seizure activity, and improves behavior and hippocampal synaptic plasticity in an Angelman syndrome mouse model. *Neurobiol Dis.* 96:38-46.
- Clarke, K., Tchabanenko, K., Pawlosky, R., Carter, E., Todd King, M., Musa-Veloso, K., Ho, M., Roberts, A., Robertson, J., Vanitallie, T.B. and Veech R.L. (2012). Kinetics, safety and tolerability of (R)-3-hydroxybutyl (R)-3-hydroxybutyrate in healthy adult subjects. *Regul Toxicol Pharmacol* 63:401-408.
- Cox, P.J. and Clarke, K. (2014). Acute nutritional ketosis: implications for exercise performance and metabolism. *Extrem Physiol Med* 3:17.
- Cox, P.J., Kirk, T., Ashmore, T., Willerton, K., Evans, R., Smith, A., Murray, A.J., Stubbs, B., West, J., McLure, S.W., King, M.T., Dodd, M.S., Holloway, C., Neubauer, S., Drawer, S., Veech, R.L., Griffin, J.L. and Clarke K. (2016). Nutritional Ketosis Alters Fuel Preference and Thereby Endurance Performance in Athletes. *Cell Metab* 24:256-268.
- Coyle, E.F., Coggan, A.R., Hemmert, M.K. and Ivy, J.L. (1986) Muscle glycogen utilization during prolonged strenuous exercise when fed carbohydrate, *J Appl Physiol.* 61(1):165-172.
- Cummins, C., Orr, R., O'Connor, H. and West, C. (2013) Global positioning systems (GPS) and microtechnology sensors in team sports: a systematic review. *Sports Med.* 43(10):1025-42.
- Cunnane, S.C., Courchesne-Loyer, A., Vandenberghe, C., St-Pierre, V., Fortier, M., Hennebelle, M., Croteau, E., Bocti, C., Fulop, T. and Castellano, C.A. (2016) Can Ketones Help Rescue Brain Fuel Supply in Later Life? Implications for Cognitive Health during Aging and the Treatment of Alzheimer's Disease. *Front Mole Neurosci.* 9:53.
- Custer, E.F., Myers, J.L., Poffenbarger, P.L. and Schoen, I. (1983). The storage stability of 3-hydroxybutyrate in serum, plasma, and whole blood. *Am J Clin Pathol.* 80(3):375-80.
- Davison, R.C. R., and Wooles, A.L. (2007). Cycling. In E. M. Winter, A. M. Jones, R. C. R. Davison, P. D. Bromley & T. H. Mercer (Eds.), *Sport and Exercise Physiology Testing Guidelines: Volume I – Sport Testing* (pp. 160-164). Oxon, UK: Routledge.

- Decombaz, J., Arnaud, M.J., Milon, H., Moesch, H., Philippoussian, G., Thelin, A.L. and Howald, H. (1983) Energy metabolism of medium-chain triglycerides versus carbohydrates during exercise, *Eur J Appl Physiol Occup Physiol*. 52(1):9-14.
- Desrochers, S., Dubreuil, P., Brunet, J., Jetté, M., David, F., Landau, B.R., Brunengraber, H. (1995) Metabolism of (R,S)-1,3-butanediol acetoacetate esters, potential parenteral and enteral nutrients in conscious pigs. *Am J Physiol*. E660-667.
- Dhatariya, K. (2016) Blood Ketones: Measurement, Interpretation, Limitations, and Utility in the Management of Diabetic Ketoacidosis. *Rev Diabet Stud*, 13(4):217-225.
- Edwards, C., Canfield, J., Copes, N., Rehan, M., Lipps, D., & Bradshaw, P.C. (2014). D-beta-hydroxybutyrate extends lifespan in *C. elegans*. *Aging*. 6(8):621-644.
- Egan, B., Carson, B.P., Garcia-Roves, P.M., Chibalin, A.V., Sarsfield, F.M., Barron, N., McCaffrey, N., Moyna, N.M., Zierath, J.R. and O'Gorman, D.J. (2010). Exercise intensity-dependent regulation of peroxisome proliferator-activated receptor gamma coactivator-1 alpha mRNA abundance is associated with differential activation of upstream signalling kinases in human skeletal muscle. *J Physiol* 588:1779-1790.
- Egan, B. and Zierath, J.R. (2013). Exercise metabolism and the molecular regulation of skeletal muscle adaptation. *Cell Metab* 17:162-184.
- Egan, B., D'Agostino, D.P. (2016) Fueling Performance: Ketones Enter the Mix. *Cell Metab*. 24(3):373-5.
- El Midaoui, A., Chiasson, J.L., Tancrede, G. and Nadeau, A. (2006). Physical training reverses the increased activity of the hepatic ketone body synthesis pathway in chronically diabetic rats. *Am J Physiol Endocrinol Metab* 290:E207-212.
- Elia, M., Wood, S., Khan, K. and Pullicino, E. (1990). Ketone body metabolism in lean male adults during short-term starvation, with particular reference to forearm muscle metabolism. *Clin Sci*. 78:579-584.
- Evans, M., Cogan, K. E., and Egan, B. (2017). Metabolism of ketone bodies during exercise and training: physiological basis for exogenous supplementation. *J Physiol*. 595(9):2857-2871.
- Evans, M., Patchett, E., Nally, R., Kearns, R., Larney, M. and Egan B. (2018) Effect of acute ingestion of beta-hydroxybutyrate salts on the response to graded exercise in trained cyclists. *Eur J Sport Sci*. 18(3):376-86.
- Evans, M., Egan, B. (2018) Intermittent Running and Cognitive Performance after Ketone Ester Ingestion. *Med Sci Sports Exerc*. 50(11):2330-2338.

- Farquhar, W.B., Paul, E.E., Prettyman, A.V. and Stillabower, M.E. (2005). Blood pressure and hemodynamic responses to an acute sodium load in humans. *J Appl Physiol.* 99(4):1545-1551.
- Fischer, T., Och, U., Klawon, I., Och, T., Grüneberg, M., Fobker, M., Borderwick Dell, U. and Marquardt, T. (2018). Effect of a sodium and calcium DL- β -hydroxybutyrate salt in healthy adults. *J Nutr Metab.* 2018:8.
- Fery, F. and Balasse, E.O. (1983). Ketone body turnover during and after exercise in overnight-fasted and starved humans. *Am J Physiol* 245:E318-325.
- Fery, F. and Balasse, E.O. (1986). Response of ketone body metabolism to exercise during transition from postabsorptive to fasted state. *Am J Physiol* 250:E495-501.
- Fery, F. and Balasse, E.O. (1988). Effect of exercise on the disposal of infused ketone bodies in humans. *J Clin Endocrinol Metab* 67:245-250.
- Fournier, P.A., Brau, L., Ferreira, L.D., Fairchild, T., Raja, G., James, A. and Palmer, T.N. (2002). Glycogen resynthesis in the absence of food ingestion during recovery from moderate or high intensity physical activity: novel insights from rat and human studies. *Comp Biochem Physiol A Mol Integr Physiol* 133:755-763.
- Ford, K. and Glymour, C. (2014) The enhanced warfighter. *Bulletin of the Atomic Scientists.* 70(1): 43-53.
- Frayn, K.N. (1983). Calculation of substrate oxidation rates in vivo from gaseous exchange. *J Appl Physiol* 55: 628-634.
- Fritzsche, I., Buhrdel, P., Melcher, R. and Bohme, H. J. (2001) Stability of ketone bodies in serum in dependence on storage time and storage temperature. *Clin Lab.* 47(7-8):399-403.
- Gaesser, G.A., and Brooks, G.A. (1975). Muscular efficiency during steady-rate exercise: effects of speed and work rate. *J Appl Physiol.* 38(6):1132-1139.
- Gaitanos, G.C., Williams, C., Boobis, L.H. and Brooks S. (1993). Human muscle metabolism during intermittent maximal exercise. *J Appl Physiol.* 75(2):712-9.
- Galán, A., Hernández, J. and Jimenez, O. (2001) Measurement of blood acetoacetate and β -hydroxybutyrate in an automatic analyser. *J Autom Methods Manag Chemi.* 23(3):69-76.

- Gao, Z., Yin, J., Zhang, J., Ward, R.E., Martin, R.J., Lefevre, M., Cefalu, W.T. and Ye, J. (2009). Butyrate improves insulin sensitivity and increases energy expenditure in mice. *Diabetes* 58:1509-1517.
- Gormsen, L.C., Svart, M., Thomsen, H.H., Sondergaard, E., Vendelbo, M.H., Christensen, N., Tolbod, L.P., Harms, H.J., Nielsen, R., Wiggers, H., Jessen, N., Hansen, J., Bøtker, H.E and Møller, N. (2017). Ketone Body Infusion With 3-Hydroxybutyrate Reduces Myocardial Glucose Uptake and Increases Blood Flow in Humans: A Positron Emission Tomography Study. *J Am Heart Assoc.* 6(3) pii: e005066.
- Guimont, M.C., Desjobert, H., Fonfrede, M., Vitoux, D., Benoist, J. F., Launay, J. M., Peoc'h, K. and Lefevre, G. (2015). Multicentric evaluation of eight glucose and four ketone blood meters. *Clin Biochem*, 48(18):1310-1316.
- Hasselbalch, S.G., Madsen, P.L., Hageman, L.P., Olsen, K.S., Justesen, N., Holm, S. and Paulson, O. B. (1996) Changes in cerebral blood flow and carbohydrate metabolism during acute hyperketonemia. *Am J Physiol.* 270(5 Pt 1):E746-51.
- Hasselbalch, S.G., Knudsen, G.M., Jakobsen, J., Hageman, L.P., Holm, S., Paulson, O.B. (1994) Brain metabolism during short-term starvation. *J Cereb Blood Flow Metab* 14(1):125-131.
- Hagenfeldt, L. and Wahren, J. (1968). Human forearm muscle metabolism during exercise. 3. Uptake, release and oxidation of beta-hydroxybutyrate and observations on the beta-hydroxybutyrate/acetoacetate ratio. *Scand J Clin Lab Invest* 21:314-320.
- Hagenfeldt, L. and Wahren, J. (1971). Human forearm muscle metabolism during exercise. VI. Substrate utilization in prolonged fasting. *Scand J Clin Lab Invest* 27:299-306.
- Hashimoto, T., Hussien, R., Oommen, S., Gohil, K. and Brooks GA. (2007). Lactate sensitive transcription factor network in L6 cells: activation of MCT1 and mitochondrial biogenesis. *FASEB J* 21:2602-2612.
- Havemann, L., West, S.J., Goedecke, J.H., Macdonald, I.A., St Clair Gibson, A., Noakes, T.D. and Lambert, E.V. (2006). Fat adaptation followed by carbohydrate loading compromises high-intensity sprint performance. *J Appl Physiol* (1985). 100:194-202.
- Hawley, J.A. and Leckey, J.J. (2015) Carbohydrate Dependence During Prolonged, Intense Endurance Exercise. *Sports Med.* 45 Suppl 1(Suppl 1):S5-S12.
- Henry, R.R., Brechtel, G. and Lim, K.H. (1990) Effects of ketone bodies on carbohydrate metabolism in non-insulin-dependent (type II) diabetes mellitus, *Metabolism*, 39(8):853-8.

- Holdsworth, D., Cox, P.J., Kirk, T., Stradling, H., Impey, S.G. and Clarke K. (2017). A ketone ester drink increases postexercise muscle glycogen synthesis in humans. *Med Sci Sports Exerc.* 49(9):1789-1795,
- Hopkins, W.G., Marshall, S.W., Batterham, A.M., Hanin, J. (2009) Progressive statistics for studies in sports medicine and exercise science. *Med Sci Sports Exerc.* 41(1):3-13.
- Hsu, W.Y., Kuo, C.Y., Fukushima, T., Imai, K., Chen, C. M., Lin, P.Y., & Lee, J.A. (2011). Enantioselective determination of 3-hydroxybutyrate in the tissues of normal and streptozotocin-induced diabetic rats of different ages. *J Chromatogr B Analyt Technol Biomed Life Sci.* 879(29), 3331-3336.
- Impey, S.G., Hammond, K.M., Shepherd, S.O., Sharples, A.P., Stewart, C., Limb, M., Smith, K., Philp, A., Jeromson, S., Hamilton, D.L., Close, G.L. and Morton JP. (2016). Fuel for the work required: a practical approach to amalgamating train-low paradigms for endurance athletes. *Physiol Rep* 4: pii: e12803.
- Jacobs, I., Westlin, N., Karlsson, J., Rasmusson, M., Houghton, B. (1982) Muscle glycogen and diet in elite soccer players. *Eur J Appl Physiol Occup Physiol.* 48(3):297-302.
- Jeukendrup, A.E. (2004) Carbohydrate intake during exercise and performance. *Nutrition.* 20(7):669-677.
- Jeukendrup, A.E., Saris, W.H., Schrauwen, P., Brouns, F. and Wagenmakers, A.J. (1995) Metabolic availability of medium-chain triglycerides coingested with carbohydrates during prolonged exercise. *J Appl Physiol.* 79(3):756-762.
- Jeukendrup, A.E., Thielen, J.J., Wagenmakers, A.J., Brouns, F. and Saris, W.H. (1998) Effect of medium-chain triacylglycerol and carbohydrate ingestion during exercise on substrate utilization and subsequent cycling performance. *Am J Clin Nutr.* 67(3):397-404.
- Jeukendrup, A.E., and Wallis, G.A. (2005). Measurement of substrate oxidation during exercise by means of gas exchange measurements. *Int J Sports Med.* 26(Suppl 1):S28-S37
- Johnson, R.H. and Walton, J.L. (1972). The effect of exercise upon acetoacetate metabolism in athletes and non-athletes. *Q J Exp Physiol Cogn Med Sci.* 57:73-79.
- Johnson, R.H., Walton, J.L., Krebs, H.A. and Williamson, D.H. (1969). Metabolic fuels during and after severe exercise in athletes and non-athletes. *Lancet.* 2:452-455.

- Kahle, L.E., Kelly, P.V., Eliot, K.A., and Weiss, E. P. (2013). Acute sodium bicarbonate loading has negligible effects on resting and exercise blood pressure but causes gastrointestinal distress. *Nutr Res.* 33(6):479-486.
- Kamysheva, V.A. and Ostrovskaia, R.U. (1980). Effect of sodium hydroxybutyrate on the ammonia level in the rat muscles under physical exercise. *Biull Eksp Biol Med.* 89:25-27.
- Kashiwaya, Y., King, M.T. and Veech, R.L. (1997). Substrate signaling by insulin: a ketone bodies ratio mimics insulin action in heart. *Am J Cardiol* 80: 50a-64a.
- Kemppainen, J., Aalto, S., Fujimoto, T., Kalliokoski, K. K., Långsjö, J., Oikonen, V., Rinne, J., Nuutila, P. and Knuuti, J. (2005) High intensity exercise decreases global brain glucose uptake in humans. *J Physiol* 568(Pt 1):323-332.
- Kesl, S.L., Poff, A.M., Ward, N.P., Fiorelli, T.N., Ari, C., Van Putten, A.J., Sherwood, J.W., Arnold, P. and D'Agostino, D.P. (2016). Effects of exogenous ketone supplementation on blood ketone, glucose, triglyceride, and lipoprotein levels in Sprague-Dawley rats. *Nutr Metab (Lond)* 13:9.
- Kies, C., Tobin, R. B., Fox, H. M. and Mehلمان, M. A. (1973) Utilization of 1,3-butanediol and nonspecific nitrogen in human adults. *J Nutr.* 103(8):1155-63.
- Kiens, B. and Richter, E.A. (1998). Utilization of skeletal muscle triacylglycerol during postexercise recovery in humans. *Am J Physiol* 275:E332-E337.
- Koeslag, J.H. (1982). Post-exercise ketosis and the hormone response to exercise: a review. *Med Sci Sports Exerc* 14:327-334.
- Koeslag, J.H., Levinrad, L.I., Lochner, J.D. and Sive A.A. (1985). Post-exercise ketosis in post-prandial exercise: effect of glucose and alanine ingestion in humans. *J Physiol* 358: 395-403.
- Koeslag, J.H., Noakes, T.D. and Sloan, A.W. (1980). Post-exercise ketosis. *J Physiol* 301:79-90.
- Koeslag, J.H., Noakes, T.D. and Sloan, A.W. (1982). The effects of alanine, glucose and starch ingestion on the ketosis produced by exercise and by starvation. *J Physiol.* 325: 363-376.
- Kovacs, Z., D'Agostino, D.P., Dobolyi, A. and Ari, C. (2017). Adenosine A1 Receptor Antagonism Abolished the Anti-seizure Effects of Exogenous Ketone

- Supplementation in Wistar Albino Glaxo Rijswijk Rats. *Front Molec Neurosci.* 10:235.
- Krikorian, R., Shidler, M.D., Dangelo, K., Couch, S.C., Benoit, S.C. and Clegg, D.J. (2012) Dietary ketosis enhances memory in mild cognitive impairment. *Neurobiol Aging* 33(2):425.
- Laffel, L. (1999). Ketone bodies: a review of physiology, pathophysiology and application of monitoring to diabetes. *Diabetes Metab Res Rev* 15:412-426.
- Laughlin, M.R., Taylor, J., Chesnick, A.S. and Balaban, R.S. (1994). Nonglucose substrates increase glycogen synthesis in vivo in dog heart. *Am J Physiol* 267:H219-223.
- Lardy, H.A., Hansen, R.G. and Phillips, P.H. (1945) The metabolism of bovine epididymal spermatozoa. *Arch. Biochem.* 6:41-51.
- Lardy, H.A. and Phillips, P.H. (1945) Studies of fat and carbohydrate oxidation in mammalian spermatozoa. *Arch. Biochem.* 6:53-61.
- Leckey, J.J., Ross, M.L., Quod, M., Hawley, J.A. and Burke, L.M. (2017) Ketone Diester Ingestion Impairs Time-Trial Performance in Professional Cyclists. *Front Physiol.* 8:806.
- Lestan, B., Walden, K., Schmaltz, S., Spsychala, J. and Fox, I.H. (1994). beta-Hydroxybutyrate decreases adenosine triphosphate degradation products in human subjects. *J Lab Clin Med* 124:199-209.
- Maiz, A., Moldawer, L.L., Bistran, B.R., Birkhahn, R.H., Long, C.L. and Blackburn, G.L. (1985) Monoacetoacetin and protein metabolism during parenteral nutrition in burned rats. *Biochem J.* 226(1):43-50
- Maizels, E.Z., Ruderman, N.B., Goodman, M.N. and Lau, D. (1977). Effect of acetoacetate on glucose metabolism in the soleus and extensor digitorum longus muscles of the rat. *Biochem J* 162:557-568.
- Maughan, R.J., Burke, L.M., Dvorak, J., Larson-Meyer, D.E., Peeling, P., Phillips, S.M., Rawson, E.S., Walsh, N.P., Garthe, I., Geyer, H., Meeusen, R., van Loon, L.J.C., Shirreffs, S.M., Spriet, L.L., Stuart, M., Verne, A., Currell, K., Ali, V. M., Budgett, R. G.M., Ljungqvist, A., Mountjoy, M., Pitsiladis, Y.P., Soligard, T., Erdener, U. and Engebretsen, L. (2018) IOC consensus statement: dietary supplements and the high-performance athlete. *Br J Sports Med.* 52(7):439-455.
- Mazzeo, R.S., Brooks, G.A., Schoeller, D.A. and Budinger, T.F. (1986). Disposal of blood [1-13C]lactate in humans during rest and exercise. *J Appl Physiol* (1985) 60:232-241.

- McGee, S.L., Fairlie, E., Garnham, A.P. and Hargreaves M. (2009). Exercise-induced histone modifications in human skeletal muscle. *J Physiol* 587:5951-5958.
- McGee, S.L. and Hargreaves M. (2004). Exercise and myocyte enhancer factor 2 regulation in human skeletal muscle. *Diabetes* 53:1208-1214.
- McKinsey, T.A., Zhang, C.L. and Olson, E.N. (2001). Control of muscle development by dueling HATs and HDACs. *Curr Opin Genet Dev* 11:497-504.
- McKenzie, E., Holbrook, T., Williamson, K., Royer, C., Valberg, S., Hinchcliff, K., Jose-Cunilleras, E., Nelson, S., Willard, M. and Davis, M. (2005) Recovery of muscle glycogen concentrations in sled dogs during prolonged exercise. *Med Sci Sports Exerc.* 37(8):1307-12.
- McKenzie, E.C., Hinchcliff, K.W., Valberg, S.J., Williamson, K.K., Payton, M.E. and Davis, M.S. (2008) Assessment of alterations in triglyceride and glycogen concentrations in muscle tissue of Alaskan sled dogs during repetitive prolonged exercise. *A J Vet Res.* 69(8):1097-1103.
- McNeil, C.A., Pramfalk, C., Humphreys, S.M. and Hodson, L. (2014) The storage stability and concentration of acetoacetate differs between blood fractions. *Clin Chim Acta.* 433:78-83.
- McSwiney, F.T., Wardrop, B., Hyde, P.N., Lafountain, R.A., Volek, J.S. and Doyle, L. (2018) Keto-adaptation enhances exercise performance and body composition responses to training in endurance athletes. *Metabolism.* 81:25-34.
- Mikkelsen, K.H., Seifert, T., Secher, N.H., Grondal, T. and van Hall, G. (2015). Systemic, cerebral and skeletal muscle ketone body and energy metabolism during acute hyper-D-beta-hydroxybutyratemia in post-absorptive healthy males. *J Clin Endocrinol Metab* 100:636-643.
- Mohr, M., Krstrup, P. and Bangsbo, J. (2003) Match performance of high-standard soccer players with special reference to development of fatigue. *J Sports Sci.* 21(7):519-28.
- Morton, J.P., Croft, L., Bartlett, J.D., Maclaren, D.P., Reilly, T., Evans, L., McArdle, A. and Drust, B. (2009). Reduced carbohydrate availability does not modulate training-induced heat shock protein adaptations but does upregulate oxidative enzyme activity in human skeletal muscle. *J Appl Physiol* 106: 1513-1521.
- Murray, A.J., Knight, N.S., Cole, M.A., Cochlin, L.E., Carter, E., Tchabanenko, K., Pichulik, T., Gulston, M.K., Atherton, H.J., Schroeder, M.A., Deacon, R.M., Kashiwaya, Y.,

- King, M.T., Pawlosky, R., Rawlins, J.N, Tyler, D.J., Griffin, J.L., Robertson, J., Veech R.L. and Clarke, K. (2016) Novel ketone diet enhances physical and cognitive performance. *FASEB J*.30(12):4021-32.
- Myette-Côté, É., Neudorf, H., Rafiei, H., Clarke, K. and Little JP. (2018) Prior ingestion of exogenous ketone monoester attenuates the glycaemic response to an oral glucose tolerance test in healthy young individuals. *J Physiol*. 596(8):1385-95.
- Nair, K.S., Welle, S.L., Halliday, D. and Campbell, R.G. (1988). Effect of beta-hydroxybutyrate on whole-body leucine kinetics and fractional mixed skeletal muscle protein synthesis in humans. *J Clin Invest* 82:198-205.
- Nath, M.C. and Brahmachari, H.D. (1944) Experimental Hyperglycaemia by Injection of Intermediary Fat Metabolism Products in Rabbits. *Nature*. 154(3911):487-488.
- Nath, M.C. and Brahmachari, H.D. (1946) Inactivation of Insulin by Intermediary Fat Metabolism Products. *Nature*. 157:336.
- Nath, M.C. and Brahmachari, H.D. (1948) Relation of intermediary metabolites to the lowering of the potency of pancreatic insulin in the animal system. *Nature*. 161:18-19.
- Nicholas, C.W., Nuttall, F.E., Williams, C. (2000) The Loughborough Intermittent Shuttle Test: a field test that simulates the activity pattern of soccer. *J Sports Sci*. 18(2):97-104.
- Noakes, T.D. (2011). Time to move beyond a brainless exercise physiology: the evidence for complex regulation of human exercise performance. *Appl Physiol Nutr Metab* 36: 23-35.
- O'Malley, T., Myette-Cote, E., Durrer, C., and Little, J.P. (2017). Nutritional ketone salts increase fat oxidation but impair high-intensity exercise performance in healthy adult males. *Appl Physiol Nutr Metab*. 42(10): 1031-1035.
- Ohmori, H., Kawai, K. and Yamashita, K. (1990). Enhanced ketone body uptake by perfused skeletal muscle in trained rats. *Endocrinol Jpn* 37:421-429.
- Oosthuyse, T., and Bosch, A.N. (2010). The effect of the menstrual cycle on exercise metabolism: implications for exercise performance in eumenorrhoeic women. *Sports Med*. 40(3), 207-227.
- Owen, O.E. (1967) Brain metabolism during fasting. *J Clin Invest*. 46(10):1589-1595.

- Owen, O.E. and Reichard, G.A., Jr. (1971). Human forearm metabolism during progressive starvation. *J Clin Invest* 50:1536-1545.
- Quistorff, B., Secher, N.H. and Van Lieshout, J.J. (2008) Lactate fuels the human brain during exercise. *FASEB J.* 22(10):3443-9.
- Paoli, A., Rubini, A., Volek, J.S. and Grimaldi, K.A. (2013). Beyond weight loss: a review of the therapeutic uses of very-low-carbohydrate (ketogenic) diets. *Eur J Clin Nutr.* 67:789-796.
- Parolin, M.L., Chesley, A., Matsos, M.P., Spriet, L.L., Jones, N.L. and Heigenhauser, G.J. (1999). Regulation of skeletal muscle glycogen phosphorylase and PDH during maximal intermittent exercise. *Am J Physiol.* 277(5 Pt 1):E890-E900.
- Pawlosky, R.J., Kemper, M.F., Kashiwaya, Y., King, M.T., Mattson, M.P. and Veech, R.L. (2017) Effects of a dietary ketone ester on hippocampal glycolytic and tricarboxylic acid cycle intermediates and amino acids in a 3xTgAD mouse model of Alzheimer's disease. *J Neurochem.* 141(2):195-207.
- Phinney, S.D., Bistrian, B.R., Evans, W.J., Gervino, E. and Blackburn, G.L. (1983) The human metabolic response to chronic ketosis without caloric restriction: preservation of submaximal exercise capability with reduced carbohydrate oxidation. *Metabolism* 32(8):769-76.
- Phinney, S.D., Horton, E.S., Sims, E.A., Hanson, J.S., Danforth, E., Jr. and LaGrange, B.M. (1980) Capacity for moderate exercise in obese subjects after adaptation to a hypocaloric, ketogenic diet. *J Clin Invest.* 66(5):1152-61.
- Pinckaers, P.J., Churchward-Venne, T.A., Bailey, D. and van Loon, L.J. (2016). Ketone Bodies and Exercise Performance: The Next Magic Bullet or Merely Hype? *Sports Med.* 47(3): 383-391.
- Price, C.P., Llyod, B. and Alberti, G.M. (1977) A kinetic spectrophotometric assay for rapid determination of acetoacetate in blood. *Clin Chem.* 23(10):1893-7.
- Randle, P.J., Newsholme, E.A. and Garland, P.B. (1964). Regulation of glucose uptake by muscle. 8. Effects of fatty acids, ketone bodies and pyruvate, and of alloxan-diabetes and starvation, on the uptake and metabolic fate of glucose in rat heart and diaphragm muscles. *Biochem J.* 93:652-665.

- Rasmussen, P., Wyss, M.T. and Lundby, C. (2011) Cerebral glucose and lactate consumption during cerebral activation by physical activity in humans *FASEB J.* 25(9):2865-73.
- Reger, M.A., Henderson, S.T., Hale, C., Cholerton, B., Baker, L.D., Watson, G. S., Hyde, K., Chapman, D., and Craft, S. Effects of beta-hydroxybutyrate on cognition in memory-impaired adults. (2004). *Neurobiol Aging.* 25(3):311-4.
- Rennie, M.J., Jennett, S. and Johnson, R.H. (1974). The metabolic effects of strenuous exercise: a comparison between untrained subjects and racing cyclists. *Q J Exp Physiol Cogn Med Sci* 59:201-212.
- Rennie, M.J. and Johnson, R.H. (1974). Alteration of metabolic and hormonal responses to exercise by physical training. *Eur J Appl Physiol Occup Physiol* 33:215-226.
- Rennie, M.J. and Johnson, R.H. (1974). Effects of an exercise-diet program on metabolic changes with exercise in runners. *J Appl Physiol* 37:821-825.
- Rhea, M.R. (2004). Determining the magnitude of treatment effects in strength training research through the use of the effect size. *Journal of Strength and Conditioning Research*, 18(4), 918-920.
- Roberts, L.D., Bostrom, P., O'Sullivan, J.F., Schinzel, R.T., Lewis, G.D., Dejam, A., Lee, Y.K., Palma, M.J., Calhoun, S., Georgiadi, A., Chen, M.H., Ramachandran, V.S., Larson, M.G., Bouchard, C., Rankinen, T., Souza, A.L., Clish, C.B., Wang, T.J., Estall, J.L., Soukas, A.A., Cowan, C.A., Spiegelman, B.M. and Gerszten, R.E. (2014). beta-Aminoisobutyric acid induces browning of white fat and hepatic beta-oxidation and is inversely correlated with cardiometabolic risk factors. *Cell Metab* 19:96-108.
- Robinson, A.M. and Williamson, D.H. (1980). Physiological roles of ketone bodies as substrates and signals in mammalian tissues. *Physiol Rev* 60:143-187.
- Rodger, S., Plews, D., Laursen, P., & Driller, M.W. (2017). Oral β -hydroxybutyrate salt fails to improve 4-minute cycling performance following submaximal exercise. *Journal of Science and Cycling*, 6(1), 26-31.
- Russell, R.D., Redmann, S.M., Ravussin, E., Hunter, G.R., Larson-Meyer, D.E. (2004) Reproducibility of endurance performance on a treadmill using a preloaded time trial. *Med Sci Sports Exerc.* 36(4):717-24.
- Sato, K., Kashiwaya, Y., Keon, C.A., Tsuchiya, N., King, M.T., Radda, G.K., Chance, B., Clarke, K. and Veech, R.L. (1995). Insulin, ketone bodies, and mitochondrial energy transduction. *FASEB J* 9:651-658.

- Scofield, R.F., Brady, P.S., Schumann, W.C., Kumaran, K., Ohgaku, S., Margolis, J.M., and Landau, B. R. (1982). On the lack of formation of L-(+)-3-hydroxybutyrate by liver. *Arch Biochem Biophys*. 214(1), 268-272.
- Scott, B.E., Laursen, P.B., James, L.J., Boxer, B., Chandler, Z., Lam, E., Gascoyne, T., Messenger, J. And Mears, S.A. (2018). The effect of 1,3-butanediol and carbohydrate supplementation on running performance. *J Sci Med Sport*. S1440-2440(18)30553-X.
- Shaw, D.M., Merien, F., Braakhuis, A., Plews, D., Laursen, P. and Dulson DK. The Effect of 1,3-Butanediol on Cycling Time-Trial Performance. *Int J Sport Nutr Exerc Metab*. 2019:1-27.
- Sherwin, R.S., Hendler, R.G. and Felig, P. (1975). Effect of ketone infusions on amino acid and nitrogen metabolism in man. *J Clin Invest*. 55:1382-1390.
- Shimazu, T., Hirschey, M.D., Newman, J., He, W., Shirakawa, K., Le Moan, N., Grueter, C.A., Lim, H., Saunders, L.R., Stevens, R.D., Newgard, C.B., Farese, R.V., de Cabo, R., Ulrich, S., Akassoglou, K. and Verdin, E. (2013). Suppression of oxidative stress by beta-hydroxybutyrate, an endogenous histone deacetylase inhibitor. *Science* 339:211-214.
- Spencer, M., Bishop, D., Dawson, B. and Goodman, C. (2005) Physiological and metabolic responses of repeated-sprint activities : specific to field-based team sports. *Sports Med*. 35(12):1025-44.
- Spurway, N., and Jones, A. M. (2007). Lactate testing. In E. M. Winter, A. M. Jones, R. C. R. Davison, P. D. Bromley & T. H. Mercer (Eds.), *Sport and Exercise Physiology Testing Guidelines: Volume I – Sport Testing* (pp. 112-119). Oxon, UK: Routledge.
- Stellingwerff, T., Spriet, L.L., Watt, M.J., Kimber, N.E., Hargreaves, M., Hawley, J.A. and Burke, L.M. (2006). Decreased PDH activation and glycogenolysis during exercise following fat adaptation with carbohydrate restoration. *Am J Physiol Endocrinol Metab* 290:E380-E388.
- Stubbs, B.J., Cox, P.J., Evans, R.D., Santer, P., Miller, J.J., Faull, O.K., Magor-Elliott, S., Hiyama, S. and Clarke, K. (2017). On the metabolism of exogenous ketones in humans. *Front Physiol*. 8:848.
- Stubbs, B.J., Cox, P.J., Evans, R.D., Cyranka, M., Clarke, K. and de Wet, H. (2018) A Ketone Ester Drink Lowers Human Ghrelin and Appetite. *Obesity (Silver Spring)*. 26(2):269-73.
- Svart, M., Gormsen, L.C., Hansen, J., Zeidler, D., Gejl, M., Vang, K., Aanerud, J., Moeller, N. (2018). Regional cerebral effects of ketone body infusion with 3-hydroxybutyrate

- in humans: Reduced glucose uptake, unchanged oxygen consumption and increased blood flow by positron emission tomography. A randomized, controlled trial. *PLoS One*. 2018;13(2):e0190556.
- Svensson, K., Albert, V., Cardel, B., Salatino, S. and Handschin C. (2016). Skeletal muscle PGC-1 α modulates systemic ketone body homeostasis and ameliorates diabetic hyperketonemia in mice. *FASEB J*. 30:1976-1986.
- Tidwell, H.C. and Axelrod, H.E. (1948) Blood sugar after injection of acetoacetate. *J Biol Chem*. 172(1):179-184.
- Tidwell, H.C. and Nagler, M.E. (1952) Effect of acetoacetate upon utilization of carbohydrate. *J Biol Chem*. 201:727-733
- Thomas, C., Bishop, D.J., Lambert, K., Mercier, J. and Brooks, G.A. (2012). Effects of acute and chronic exercise on sarcolemmal MCT1 and MCT4 contents in human skeletal muscles: current status. *Am J Physiol Regul Integr Comp Physiol* 302:R1-14.
- Thomas, L.K., Ittmann, M. and Cooper, C. (1982). The role of leucine in ketogenesis in starved rats. *Biochem J* 204:399-403.
- Thomsen, H.H. Rittig, N., Johannsen, M., Møller, A.B., Jørgensen, J.O. and Møller, N. (2018) Effects of 3-hydroxybutyrate and free fatty acids on muscle protein kinetics and signaling during LPS-induced inflammation in humans: antitabolic impact of ketone bodies. *Am J Clin Nutr*. 108:857–867.
- Tomprowski, P. D. (2003) Effects of acute bouts of exercise on cognition. *Acta Psychol (Amst)*. 112(3):297-324.
- Torrens, S.L., Areta, J.L., Parr, E.B. and Hawley, J.A. (2016). Carbohydrate dependence during prolonged simulated cycling time trials. *Eur J Appl Physiol* 116:781-790.
- Tsai, Y.C., Chou, Y.C., Wu, A.B., Hu, C.M., Chen, C.Y., Chen, F.A., & Lee, J.A. (2006). Stereoselective effects of 3-hydroxybutyrate on glucose utilization of rat cardiomyocytes. *Life Sciences*. 78(12):1385-1391.
- Vandoorne, T., De Smet, S., Ramaekers, M., Van Thienen, R., De Bock, K., Clarke, K. and Hespel, P. (2017). Intake of a Ketone Ester Drink during Recovery from Exercise Promotes mTORC1 Signaling but Not Glycogen Resynthesis in Human Muscle. *Front Physiol*. 8:310.

- van Loon, L.J., Greenhaff, P.L., Constantin-Teodosiu, D., Saris, W.H. and Wagenmakers, A.J. (2001). The effects of increasing exercise intensity on muscle fuel utilisation in humans. *J Physiol* 536:295-304.
- van Loon, L.J., Thomason-Hughes, M., Constantin-Teodosiu, D., Koopman, R., Greenhaff, P.L., Hardie, D.G., Keizer, H.A., Saris, W.H. and Wagenmakers, A.J. (2005). Inhibition of adipose tissue lipolysis increases intramuscular lipid and glycogen use in vivo in humans. *Am J Physiol Endocrinol Metab* 289:E482-E493.
- Veech, R.L. (2004). The therapeutic implications of ketone bodies: the effects of ketone bodies in pathological conditions: ketosis, ketogenic diet, redox states, insulin resistance, and mitochondrial metabolism. *Prostaglandins Leukot Essent Fatty Acids* 70:309-319.
- Volek, J.S., Freidenreich, D.J., Saenz, C., Kunces, L.J., Creighton, B.C., Bartley, J.M., Davitt, P.M., Munoz, C.X., Anderson, J.M., Maresh, C.M., Lee, E.C., Schuenke, M.D., Aerni, G., Kraemer, W.J. and Phinney SD. (2016). Metabolic characteristics of keto-adapted ultra-endurance runners. *Metabolism* 65:100-110.
- Volek, J.S., Noakes, T., Phinney, S.D. (2015) Rethinking fat as a fuel for endurance exercise. *Eur J Sport Sci.* 15(1):13-20.
- Wagenmakers, A.J., Beckers, E.J., Brouns, F., Kuipers, H., Soeters, P.B., van der Vusse, G.J. and Saris, W.H. (1991). Carbohydrate supplementation, glycogen depletion, and amino acid metabolism during exercise. *Am J Physiol* 260:E883-E890.
- Wahren, J., Sato, Y., Ostman, J., Hagenfeldt, L. and Felig P. (1984). Turnover and splanchnic metabolism of free fatty acids and ketones in insulin-dependent diabetics at rest and in response to exercise. *J Clin Invest* 73:1367-1376.
- Waldman, H.S., Basham, S.A., Price, F.G., Smith, J.W., Chander, H., Knight, A.C., Krings, B.M. and McAllister (2018). Exogenous ketone salts do not improve cognitive responses after a high-intensity exercise protocol in healthy college-aged males. *Appl Physiol Nutr Metab.* 43(7) 711-717.
- Webber, R.J., and Edmond, J. (1977). Utilization of L(+)-3-hydroxybutyrate, D(-)-3-hydroxybutyrate, acetoacetate, and glucose for respiration and lipid synthesis in the 18-day-old rat. *Journal Biol Chem.* 252(15):5222-5226.
- Weeber, E., Ciarlone, S. and D'Agostino, D. (2016) Ketone esters for treatment of angelman syndrome. [Online]. *USF Patents*. Patent Number: 9364456.

- Winder, W.W., Baldwin, K.M. and Holloszy, J.O. (1973). Exercise-induced adaptive increase in rate of oxidation of beta-hydroxybutyrate by skeletal muscle. *Proc Soc Exp Biol Med* 143:753-755.
- Winder, W.W., Baldwin, K.M. and Holloszy, J.O. (1974). Enzymes involved in ketone utilization in different types of muscle: adaptation to exercise. *Eur J Biochem.* 47:461-467.
- Winder, W.W., Baldwin, K.M. and Holloszy, J.O. (1975). Exercise-induced increase in the capacity of rat skeletal muscle to oxidize ketones. *Can J Physiol Pharmacol.* 53:86-91.
- Yamada, T., Zhang, S.J., Westerblad, H., and Katz, A. (2010). {beta}-Hydroxybutyrate inhibits insulin-mediated glucose transport in mouse oxidative muscle. *Am J Physiol Endocrinol Metab.*299(3):E364-373.
- Youm, Y.H., Nguyen, K.Y., Grant, R.W., Goldberg, E.L., Bodogai, M., Kim, D., D'Agostino, D.P., Planavsky, N., Lupfer, C., Kanneganti, T.D., Kang, S., Horvath, T.L., Fahmy, T.M., Crawford, P.A., Biragyn, A., Alnemri, E. and Dixit, V.D. (2015). The ketone metabolite beta-hydroxybutyrate blocks NLRP3 inflammasome-mediated inflammatory disease. *Nat Med* 21:263-269.
- Zou, X., Meng, J., Li, L., Han, W., Li, C., Zhong, R., Miao, X., Cai, J., Zhang, Y. and Zhu, D. (2016). Acetoacetate Accelerates Muscle Regeneration and Ameliorates Muscular Dystrophy in Mice. *J Biol Chem* 291: 2181-2195.

Appendices


Appendix A: Published Journal of Physiology article

J Physiol 000.00 (2016) pp 1–15

1

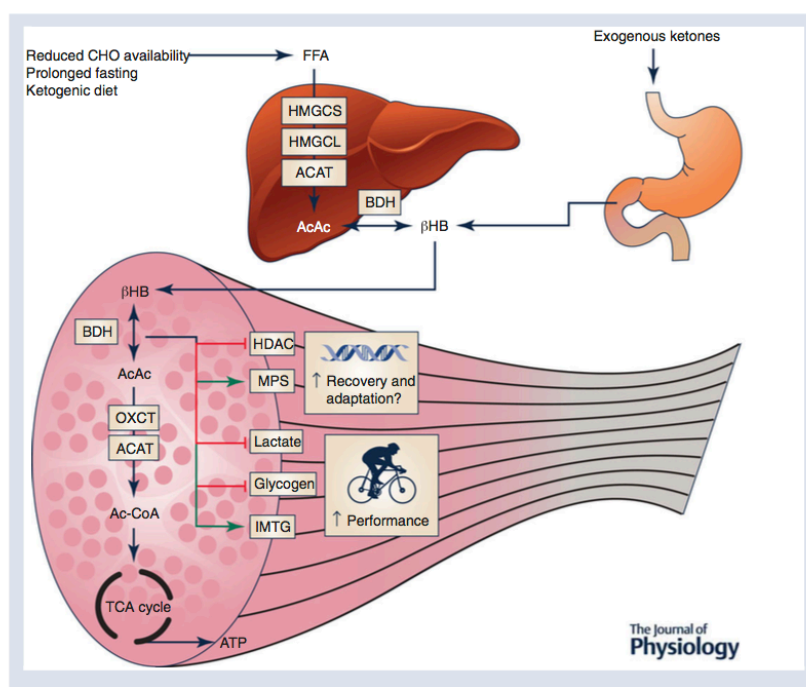
TOPICAL REVIEW

Metabolism of ketone bodies during exercise and training: physiological basis for exogenous supplementation

Mark Evans¹, Karl E. Cogan¹ and Brendan Egan^{1,2} 

¹Institute for Sport and Health, School of Public Health, Physiotherapy and Sports Science, University College Dublin, Belfield, Dublin, 4, Ireland

²School of Health and Human Performance, Dublin City University, Glasnevin, Dublin, 9, Ireland



Mark Evans and Karl Cogan are graduate students at the Institute for Sport and Health, University College Dublin, Ireland. Mark received his MSc in Sport Nutrition from Liverpool John Moores University in 2015. Karl received his MSc in Biotechnology from University College Dublin in 2013. Their research explores optimising nutrition strategies for performance and recovery in athletes with specific interest in ketone bodies and protein hydrolysates, respectively. Brendan Egan PhD is Senior Lecturer in Sport and Exercise Physiology at Dublin City University's School of Health and Human Performance, and Visiting Associate Professor at University College Dublin. His research group investigates the molecular regulation of skeletal muscle function, adaptation and performance across the life course with special interest in the synergy between nutrition and exercise interventions ranging from athletes to older adults. All three authors are accomplished sportsmen in their own right, and currently involved in the provision of sports science support to team sport athletes.



Abstract Optimising training and performance through nutrition strategies is central to supporting elite sportspeople, much of which has focused on manipulating the relative intake of carbohydrate and fat and their contributions as fuels for energy provision. The ketone bodies, namely acetoacetate, acetone and β -hydroxybutyrate (β HB), are produced in the liver during conditions of reduced carbohydrate availability and serve as an alternative fuel source for peripheral tissues including brain, heart and skeletal muscle. Ketone bodies are oxidised as a fuel source during exercise, are markedly elevated during the post-exercise recovery period, and the ability to utilise ketone bodies is higher in exercise-trained skeletal muscle. The metabolic actions of ketone bodies can alter fuel selection through attenuating glucose utilisation in peripheral tissues, anti-lipolytic effects on adipose tissue, and attenuation of proteolysis in skeletal muscle. Moreover, ketone bodies can act as signalling metabolites, with β HB acting as an inhibitor of histone deacetylases, an important regulator of the adaptive response to exercise in skeletal muscle. Recent development of ketone esters facilitates acute ingestion of β HB that results in nutritional ketosis without necessitating restrictive dietary practices. Initial reports suggest this strategy alters the metabolic response to exercise and improves exercise performance, while other lines of evidence suggest roles in recovery from exercise. The present review focuses on the physiology of ketone bodies during and after exercise and in response to training, with specific interest in exploring the physiological basis for exogenous ketone supplementation and potential benefits for performance and recovery in athletes.

(Received 4 August 2016; accepted after revision 24 October 2016; first published online 10 November 2016)

Corresponding author B. Egan: School of Health and Human Performance, Dublin City University, Glasnevin, Dublin 9, Ireland. Email: brendan.egan@dcu.ie

Abstract figure legend Acetoacetate (AcAc) and β -hydroxybutyrate (β HB) are ketone bodies produced in hepatic mitochondria during conditions of reduced carbohydrate availability and serve as an alternative fuel source for peripheral tissues including skeletal muscle. Elevations in β HB can result from endogenous production i.e. ketogenesis, but also by ingestion of exogenous ketone supplements such as ketone salts or ketone esters. Ketogenesis from free fatty acids (FFA) involves sequential reactions of Ac-CoA acetyltransferase (ACAT), hydroxymethylglutaryl CoA synthase (HMGCS), and hydroxymethylglutaryl-CoA lyase (HMGCL). The end product of ketogenesis is AcAc, the majority of which is reduced to β HB by 3-hydroxybutyrate dehydrogenase (BDH) before entering the circulation. Upon uptake into peripheral tissues, β HB is oxidised to AcAc. Reactions of succinyl-CoA:3-oxoacid CoA transferase (OXCT) and ACAT ultimately produce acetyl CoA (Ac-CoA), which enters the TCA cycle for ATP synthesis. The metabolic actions of β HB include altered fuel selection during exercise through attenuating glycogen utilisation, lowering lactate production and increasing reliance on intramuscular triglyceride (IMTG). Additionally, β HB may regulate adaptive processes in skeletal muscle by acting as a signalling metabolite inhibiting histone deacetylases (HDAC), or through positive effects on muscle protein synthesis (MPS). Ketone ester supplements facilitate acute ingestion of β HB resulting in nutritional ketosis, which, through these mechanisms, may alter exercise metabolism, improve exercise performance, and influence recovery and the adaptive response to exercise.

Abbreviations AcAc, acetoacetate; AcAc-CoA, acetoacetyl CoA; ACAT, acetyl-CoA acetyltransferase; β HB, β -hydroxybutyrate; BDH, 3-hydroxybutyrate dehydrogenase; CHO, carbohydrate; CPT1, carnitine palmitoyltransferase; FFA, free-fatty acid; HDAC, histone deacetylase; HMG-CoA, hydroxymethylglutaryl-CoA; HMGCL, HMG-CoA lyase; HMGCS, HMG CoA synthase; KB, ketone body; KE, (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester; MCT, monocarboxylate transporter; OXCT, succinyl-CoA:3-oxoacid CoA transferase; PDH, pyruvate dehydrogenase; PEK, post-exercise ketosis; PFK, phosphofructokinase; PGC-1, peroxisome proliferator-activated receptor gamma coactivator 1; SLC, solute ligand carrier; TCA, tricarboxylic acid.

Introduction

Over the past century, exercise physiologists have appreciated the role of carbohydrate (CHO) and fat in energy provision to exercising skeletal muscle. Much of the work examining the metabolic response to exercise and the impact of exercise on metabolic regulation

and adaptive responses to training has focused on the relative contribution of these fuels (Egan & Zierath, 2013). Optimising training and nutrition strategies by manipulating the relative intakes of these macronutrients is central to supporting elite sports performance (Cermak & van Loon, 2013; Bartlett *et al.* 2015; Burke, 2015).

An alternative fuel source to CHO and fat are ketone bodies (KBs), namely acetoacetate (AcAc), acetone, and β -hydroxybutyrate (β HB), which are produced in the liver during physiological states and nutritional manipulations that result in reduced CHO availability, most commonly during prolonged fasting, starvation, and ketogenic (very low CHO (~5%), low protein (~15%), high fat (~80%)) diets (Robinson & Williamson, 1980; Laffel, 1999). This relative glucose deprivation and concomitant elevation in circulating free-fatty acids (FFAs) results in the production of KBs to replace glucose as the primary fuel for peripheral tissues such as the brain, heart and skeletal muscle in these states.

Aside from a role as an alternative fuel source, KBs exert a range of metabolic effects including attenuating glucose utilisation in peripheral tissues, anti-lipolytic effects on adipose tissue, and potential attenuation of proteolysis in skeletal muscle (Robinson & Williamson, 1980). KBs are utilised by working muscle during exercise (Fery & Balasse, 1986, 1988), and the capacity to take up and oxidise KBs during exercise is higher in exercise-trained skeletal muscle (Winder *et al.* 1975). Despite these observations, in addition to a glucose sparing action (Maizels *et al.* 1977) and potential to lower the exercise-induced rise in plasma [lactate] (Fery & Balasse, 1988), the potential performance benefits of KBs when provided as an exogenous fuel source has received little attention, but has been postulated (Cox & Clarke, 2014; Pinckaers *et al.* 2016). Apart from a role as an alternative fuel source, KBs may act as signalling molecules to regulate gene expression and adaptive responses (Shimazu *et al.* 2013; Zou *et al.* 2016). Moreover, therapeutic roles for KBs have long been proposed in a variety of disease states including aberrant glucose metabolism, genetic myopathies, hypoxic states and neurodegenerative pathologies (Veech, 2004). For therapeutic effects, exogenous ketones are ingested in the form of β HB salts or ketone esters to produce acute (~0.5 to 6 h) nutritional ketosis (Clarke *et al.* 2012; Kesl *et al.* 2016), but a surge in interest in KBs as a performance aid for athletes arose when ketone ester supplementation was confirmed in professional cycling (Abraham, 2015; Pinckaers *et al.* 2016). Moreover, a recent report provides the first evidence for acute nutritional ketosis achieved by ketone ester ingestion to alter the metabolic response to exercise and enhance exercise performance (Cox *et al.* 2016). Aspects of ketogenic diets, ketogenesis and ketone body metabolism have been reviewed elsewhere (Robinson & Williamson, 1980; Laffel, 1999; Paoli *et al.* 2013), so the present review will focus on the physiology of ketone bodies during and after exercise and in response to training, with specific interest in exploring the physiological basis for exogenous supplementation and potential benefits for performance and recovery in athletes.

Overview of ketone body metabolism

Ketone bodies in circulation. Plasma [KB] reflects the balance between hepatic production ('ketogenesis') and peripheral breakdown and utilisation ('ketolysis') in extra-hepatic tissues, both of which are under various levels of control as detailed in previous reviews (Robinson & Williamson, 1980; Laffel, 1999). Ketogenesis is an evolutionarily conserved adaptive response playing a critical role in survival during an energy crisis by providing a substrate for brain, which cannot utilise FFAs as a fuel source. AcAc, acetone, and β HB comprise the KBs, although β HB is not technically a ketone because the ketone moiety has been reduced to a hydroxyl group. AcAc and β HB are short-chain, four carbon organic acids that act as FFA-derived circulating substrates to provide energy to extra-hepatic tissues, whereas the contribution of acetone, readily generated by the spontaneous decarboxylation of AcAc, to energy provision is negligible. Plasma [KB] is <0.1 mM in the post-prandial state, whereas hyperketonaemia is accepted as [KB] exceeding 0.2 mM (Robinson & Williamson, 1980). Various states of CHO restriction, depletion and dysregulation produce hyperketonaemia to different degrees (Fig. 1).

Ketogenesis. The primary substrate for ketogenesis is FFAs liberated from adipose tissue. Ketogenic amino acids, namely leucine, lysine, phenylalanine, isoleucine, tryptophan, and tyrosine also serve ketogenesis, but are likely contribute to less than 5% of circulating KBs (Thomas *et al.* 1982). The rise in FFAs is consequent to the stimulation of lipolysis as a result of declines in plasma glucose and insulin that are characteristic of reduced CHO availability. Factors stimulating ketogenesis include an elevated glucagon-to-insulin ratio and decline in hepatic glycogen concentration, while reduced blood flow to the liver or elevations in [KBs] suppress ketogenesis (Robinson & Williamson, 1980; Laffel, 1999). Ketogenesis involves a series of sequential reactions beginning with acetyl CoA (Ac-CoA) and acetoacetyl CoA (AcAc-CoA), and ending with the liberation of AcAc (Fig. 2). Some AcAc is exported, but the majority is reduced to β HB in an NAD^+ -NADH-coupled near equilibrium reaction catalysed by 3-hydroxybutyrate dehydrogenase (BDH), in which the equilibrium constant favours β HB formation. These KBs are transported into the circulation via the solute carrier (SLC) protein 16A (SLC16A) family of monocarboxylate transporters (MCTs) in mitochondrial and sarcolemmal membranes.

Ketolysis in extra-hepatic tissues. In peripheral tissues, KBs, primarily in the form of β HB, enter the mitochondrial matrix again via MCT1-mediated

transport. β HB is re-oxidised to AcAc via BDH after which sequential reactions result in the generation of two molecules of Ac-CoA (Fig. 2). These are incorporated into the TCA cycle via citrate synthase for terminal oxidation and production of ATP, which in skeletal muscle contributes to fuelling muscular work (Fery & Balasse, 1986, 1988). Succinyl-CoA:3-oxoacid CoA transferase (OXCT) is essential for ketolysis in extra-hepatic tissues, with very low abundance in hepatocytes explaining the lack of ketolytic activity in these cells (Robinson & Williamson, 1980).

Activity of OXCT is highest in heart and kidney, followed by skeletal muscle and the brain (Robinson & Williamson, 1980), but because skeletal muscle accounts for ~40% of body mass in adult humans, this organ accounts for the highest fraction of total KB metabolism at rest (Balasse & Fery, 1989; Laffel, 1999). Beginning almost 50 years ago, models using various durations of fasting, and combined with primed constant infusion of radio-labelled either AcAc or β HB tracers and arteriovenous difference measures to quantify KB turnover, established that skeletal muscle is a major site of ketolysis at rest (Hagenfeldt & Wahren, 1968; Owen & Reichard, 1971; Wahren *et al.* 1984; Elia *et al.* 1990; Mikkelsen *et al.* 2015). Skeletal muscle has a high affinity to KBs, but because of low circulating concentrations under normal conditions, the contribution to energy provision in muscle is less than 5%, and FFAs are the main source of energy provision in the post-absorptive state. The relationship between ketone oxidation and [KB] is curvilinear such that contribution to energy provision in skeletal muscle rises to ~10% after an overnight fast (Hagenfeldt & Wahren, 1968; Owen & Reichard, 1971), 20% to 50% after 72 h of fasting (Owen & Reichard, 1971; Elia *et al.* 1990), but declines to ~15% after 24 days of starvation (Owen & Reichard, 1971). Thus, skeletal muscle demonstrates saturation kinetics for the

KB concentration–oxidation relationship, with saturation likely between 1 and 2 mM as demonstrated by fasting of various durations (compiled in Balasse & Fery, 1989) or step-wise β HB infusion (Mikkelsen *et al.* 2015).

Effect of aerobic exercise training on enzymes of ketogenesis and ketolysis

Adaptations to exercise training reduce perturbations to homeostasis during subsequent bouts of exercise, and thereby enhance resistance to fatigue. Central to these effects are enhanced respiratory capacity and contractile parameters, and importantly adaptations that contribute towards maximising delivery and utilisation of circulating substrates (reviewed in Egan & Zierath, 2013). Therefore, if KBs make a meaningful contribution to energy provision during exercise, it is pertinent to explore analogous regulation in skeletal muscle. Training-induced changes in expression and activities of enzymes of ketolysis in skeletal muscle have not been described in humans, but differences in KB metabolism during and after exercise between trained and untrained individuals have been reported (Johnson *et al.* 1969; Johnson & Walton, 1972; Rennie *et al.* 1974; Rennie & Johnson, 1974a). The general pattern is for attenuation in trained individuals of the post-exercise rise in [KB], but this is influenced by nutritional manipulation and relative exercise intensity, the latter of which has often been poorly controlled (see later sections).

Nevertheless, circulating concentrations reflect the balance between ketogenesis and ketolysis, these differences may be explained by the factors influencing one or both. For ketogenesis, data are limited but suggest that in exercise-trained rodents enzymatic activity of BDH or ACAT (Winder *et al.* 1974), or HMGCS (Askew *et al.* 1975) is unaltered in liver, and, in fact, the overall activity of the ketogenic pathway may be lower (El

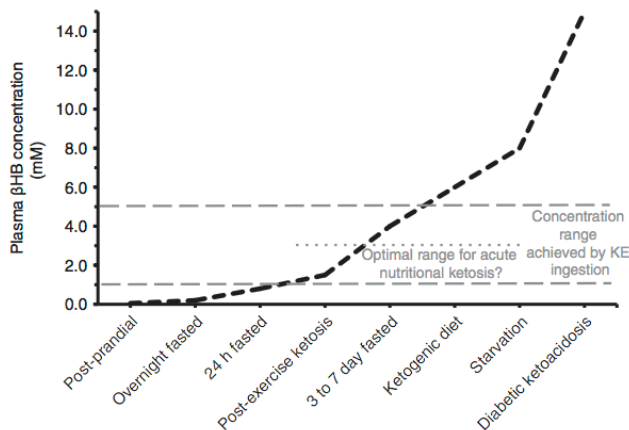


Figure 1. Changes in [β HB] under various physiological states

Plasma [KB] is <0.1 mM in the postprandial state when consuming high CHO or high protein meals, and rises upward after an overnight fast and with ketogenic dieting, prolonged fasting, starvation, and pathological states of ketoacidosis. After prolonged aerobic exercise, post-exercise ketosis (0.3 to 2.0 mM) may ensue depending on intensity and duration of exercise, aerobic fitness and nutrition status. The circulating KB ratio of β HB:AcAc is generally ~1:1 to 3:1, but during the aforementioned nutritional states can rise six- to tenfold, such that [KB] primarily reflects changes in [β HB]. An optimal concentration range for β HB to improve performance after exogenous ketone ingestion is proposed as ~1 to 3 mM, with concentrations ranging from ~1 to 5 mM reported after ketone ester (KE) ingestion. See text for further details.

Midaoui *et al.* 2006) compared to untrained rodents. In these rodent models of intense aerobic exercise training, the activities of the ketolytic enzymes BDH, OXCT and ACAT are higher in trained skeletal muscle (Winder *et al.* 1974, 1975; Askew *et al.* 1975; Beattie & Winder, 1984). This coincides with two- to threefold higher *ex vivo* rates of β HB and AcAc oxidation in gastrocnemius muscle homogenates presented with concentrations of both β HB and AcAc at 0.1 and 0.5 mM (Winder *et al.* 1973, 1975).

In terms of muscle fibre type, enzymatic activities of BDH, OXCT and ACAT are all highest in type I fibres, intermediate in type IIA fibres, and lowest in type IIB fibres of rats (Winder *et al.* 1974). BDH is essentially undetectable in type IIB muscle fibres, and across the fibre types

BDH activity is much lower than activities of OXCT and ACAT (Winder *et al.* 1974). Although OXCT is essential for ketolysis, BDH activity is, therefore, potentially rate limiting in skeletal muscle. When rats performed 12 weeks of treadmill running, compared to sedentary rats BDH activity was almost threefold higher in type I fibres, but sixfold higher in type IIA fibres of trained skeletal muscle, resulting in levels comparable to the type I fibres (Winder *et al.* 1974). OXCT activity was 26% higher in type I, and approximately twofold higher in type IIA and IIB fibres, whereas ACAT activity was 40% to 45% higher in all three fibre types in trained skeletal muscle (Winder *et al.* 1974). Similarly, in skeletal muscle from mice with 8 weeks of access to running wheels, the difference compared to sedentary mice was greater for BDH mRNA expression

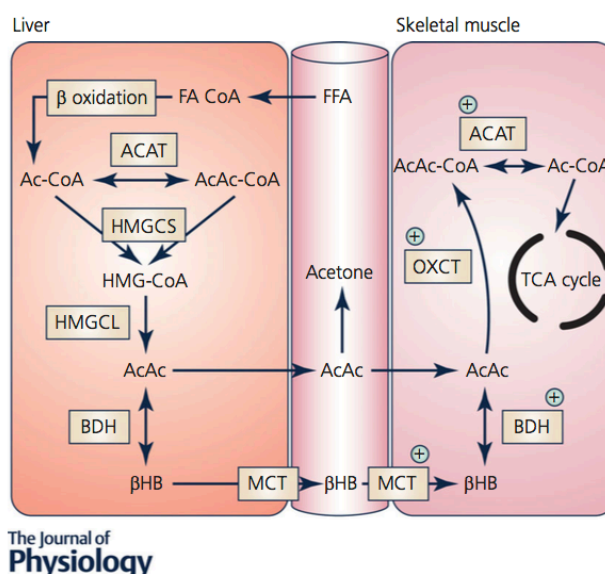


Figure 2. Metabolic pathways of ketone body metabolism in liver and skeletal muscle

Ketogenesis: FFAs are converted to fatty acyl CoA (FA-CoA), enter hepatic mitochondria via CPT1-mediated transport and undergo β -oxidation to acetyl CoA. Sequential reactions of condensation of Ac-CoA molecules to acetoacetyl CoA (AcAc-CoA) by mitochondrial thiolase activity of Ac-CoA acetyltransferase (ACAT), generation of hydroxymethylglutaryl-CoA (HMG-CoA) by hydroxymethylglutaryl CoA synthase (HMGCS), and decomposition of HMG-CoA, liberating AcAc and Ac-CoA, in a reaction catalysed by HMG-CoA lyase (HMGCL). AcAc is the central KB, and some will be exported to the circulation but the majority is reduced to β HB in an NAD^+ – NADH -coupled near equilibrium reaction catalysed by BDH, in which the equilibrium constant favours β HB formation. Ketolysis: The only metabolic fate of β HB is inter-conversion with AcAc, and upon entry into peripheral tissues it is re-oxidised to AcAc. Covalent activation of AcAc by CoA is catalysed by succinyl-CoA:3-oxoacid CoA transferase (OXCT) resulting in generation of AcAc-CoA. This near equilibrium reaction exchanges CoA between succinate and AcAc, with succinyl-CoA acting as a CoA donor. Because the free energy released by hydrolysis of AcAc-CoA is greater than that of succinyl-CoA, the equilibrium of this reaction thermodynamically favours the formation of AcAc. Two molecules of Ac-CoA are liberated by thiolytic cleavage of AcAc-CoA by ACAT, after which Ac-CoA is incorporated into the TCA cycle. Protein content and enzyme activity that are higher in exercise-trained skeletal muscle are indicated by the green cross (+).

(~twofold higher than sedentary) compared to differences in OXCT and ACAT mRNA expression (~30% to 50% higher) (Svensson *et al.* 2016). These changes in ketolytic enzymes are localised to the working muscle given the absence of change after training in the heart (Askew *et al.* 1975), kidney and brain (Winder *et al.* 1974).

In terms of KB transport into skeletal muscle, similarly to the ketolytic enzymes, MCT1 protein expression is highest in type I fibres, poorly expressed in type II fibres, and correlates well with muscle oxidative capacity (Bonen, 2001). Elevated MCT1 protein expression after exercise training is well-established for human skeletal muscle, and increases occur in an intensity-dependent manner (Thomas *et al.* 2012). Using a rodent perfused hindlimb model, the capacity for uptake of KBs in skeletal muscle at 1 mM each of β Hb and AcAc was higher in an aerobically trained group of rats, with uptake of total KB, AcAc and β Hb 33%, 27% and 53% higher, respectively, compared to untrained rats (Ohmori *et al.* 1990). Similarly, β Hb clearance during a β Hb tolerance test is higher in mice given 8 weeks of running wheel access, or with enhanced oxidative capacity consequent to skeletal muscle over-expression of PGC-1 α , a transcriptional co-activator and master regulator of mitochondrial biogenesis in adaptive responses such as exercise training (Svensson *et al.* 2016). In both conditions, this coincides with elevated expression of MCT1 and the ketolytic enzymes in skeletal muscle. Therefore, the uptake and utilisation of KBs in skeletal muscle is likely to be greatest in those individuals that are highly trained with a high proportion of type I muscle fibres and a high oxidative capacity in skeletal muscle.

Ketone body metabolism during exercise

The existing literature on fuel selection during exercise has focused almost exclusively on utilisation of CHO and fat, but skeletal muscle has the ability to resynthesize ATP from other substrates including protein, lactate and KBs (Fery & Balasse, 1986, 1988; Mazzeo *et al.* 1986; Wagenmakers *et al.* 1991). With increasing exercise intensity, the contribution of substrates to energy provisions shifts from blood-borne FFAs and glucose to increased reliance on intramuscular fuel stores, namely intramuscular triglyceride (IMTG) and muscle glycogen, such that at moderate to high intensities (>75% $\dot{V}_{O_{2max}}$) of exercise, muscle glycogen is the main source of energy provision (van Loon *et al.* 2001). This pattern is readily altered by nutritional manipulation such as CHO loading and acute CHO ingestion resulting in increased CHO utilisation (Bosch *et al.* 1996), glycogen depletion resulting in increased contribution of protein to energy provision (Wagenmakers *et al.* 1991), and habitual high fat consumption resulting in increased contribution of fat to energy provision (Volek *et al.* 2016). Clearly, skeletal muscle is a major site of ketolysis under fasting conditions, but central to the rationale for exogenous

ketone supplementation must be the observations that ketolysis increases during exercise, makes a meaningful contribution to energy provision, and can alter patterns of substrate utilisation.

The pioneering work of Hagenfeldt, Wahren and colleagues (Hagenfeldt & Wahren, 1968, 1971; Wahren *et al.* 1984) and Fery, Balasse and colleagues (Balasse *et al.* 1978; Fery & Balasse, 1983, 1986, 1988) established that KB disposal into human skeletal muscle is elevated as much as fivefold during exercise. This is generally reflected by a drop in [KB] soon after the onset of exercise, primarily β Hb, concomitant with increases in KB oxidation in skeletal muscle and elevated metabolic clearance rate (MCR). MCR is a measure of the ability of tissues to remove ketones from the blood, analogous to arteriovenous difference per unit time, but when measured during exercise is taken to represent an index of the ability of exercise to stimulate the capacity of working muscles to extract and utilise ketones (Fery & Balasse, 1983; Balasse & Fery, 1989). Because the stoichiometry of KB oxidation yields respiratory quotients of 1.00 and 0.89 for AcAc and β Hb, respectively (Frayn, 1983), calculation of oxidation rates for KBs from whole-body gas exchange data has not been routinely performed using methods that determine the relative contribution of CHO and fat oxidation. However, a recent attempt has been made (Cox *et al.* 2016) based on methods and assumptions described for KB utilisation during ketogenesis (Frayn, 1983). Previous to this, oxidation rates for KBs have historically been derived from arteriovenous differences of radiolabelled KBs across working muscles with rates calculated as a fraction of O_2 consumption or CO_2 production (Hagenfeldt & Wahren, 1968; Balasse *et al.* 1978).

Like CHO and fat utilisation, KB metabolism during exercise is influenced by a variety of factors including metabolic status (Wahren *et al.* 1984; Fery & Balasse, 1986), training status (Johnson & Walton, 1972; Rennie *et al.* 1974; Beattie & Winder, 1985), and the intensity of exercise (Cox *et al.* 2016). Given the aforementioned fibre type-specific differences for activities of ketolytic enzymes, the muscle fibre type profile of the working muscle is also likely to be an important determinant of ketolysis during exercise. However, the most important determinant of KB metabolism during exercise is the degree of ketonaemia, and the method by which this is achieved, i.e. of endogenous or exogenous origin.

Ketone body metabolism during exercise under conditions of endogenous ketosis. Like KB metabolism in resting skeletal muscle, the relationship between concentration and oxidation or MCR is curvilinear (reviewed in Balasse & Fery, 1989). At low ketonaemia (<1.0 mM) such as that produced by an overnight fast, resting MCR is as much as fourfold greater than

during prolonged fasting (Fery & Balasse, 1983). During prolonged exercise of low-to-moderate intensity after an overnight fast, MCR increases by 50% to 75% (Fery & Balasse, 1983, 1986), which indicates that working muscle has an increased capacity to extract ketones from blood compared to rest. However, when ketonaemia exceeds 2.5 mM such as that achieved by greater than 72 h of fasting, the exercise-induced rise in MCR is abolished (Fery & Balasse, 1986). Therefore, when ketosis is achieved by prolonged (>72 h) fasting there is a negligible contribution of KB oxidation to energy provision (Hagenfeldt & Wahren, 1971; Fery & Balasse, 1986), but after an overnight fast, the contribution ranges from 2 to 10% (Balasse *et al.* 1978; Fery & Balasse, 1983; Wahren *et al.* 1984). Under these conditions, the majority of energy provision in working muscle is from CHO and fat as classically described (van Loon *et al.* 2001). Moreover, unlike CHO and fat, there is progressive attenuation of the oxidation of KBs with rising ketonaemia, and thus the mobilisation of KBs is not the factor limiting oxidation in skeletal muscle. This attenuation of exercise-stimulated MCR suggests either that above a threshold concentration the capacity for skeletal muscle to oxidise KBs becomes saturated, and/or that hyperketonaemia itself is a self-inhibitory factor (Balasse & Fery, 1989). Mechanistically, this is likely to be mediated either through the inhibition of OXCT by elevated AcAc, and/or via FFA-mediated inhibition of ketolysis (Robinson & Williamson, 1980). This regulation is critical in the starvation response because the capacity of the liver to produce KBs closely matches the requirements of the brain to utilise KBs as an energy source (Robinson & Williamson, 1980). Therefore, excessive oxidation by working muscle would threaten survival, whereas its inhibition spares circulating substrate for the brain (Hagenfeldt & Wahren, 1971; Fery & Balasse, 1983).

Methods of exogenous ketone supplementation producing acute nutritional ketosis

Investigating effects of ketosis on skeletal muscle metabolism has been typically achieved by endogenous ketosis using fasting of various durations (Balasse & Fery, 1989), or by exogenous ketosis produced by either ketone salt ingestion (Johnson & Walton, 1972), or infusion of AcAc or β HB (Fery & Balasse, 1988; Mikkelsen *et al.* 2015). Endogenous ketosis may also be achieved by CHO restriction, particularly by ketogenic diets (Paoli *et al.* 2013). The practical relevance for athletes seeking performance gains of metabolic responses generated from prolonged fasting is negligible, whereas benefits of ketogenic dieting for performance with a high intensity component are equivocal (Burke, 2015). This has led to the exploration of exogenous ketone ingestion as a

means to achieve acute nutritional ketosis. Importantly, because endogenous ketosis results in concomitant elevations in FFAs and alterations in glucose, insulin and counter-regulatory hormones, isolating the metabolic effects specific to KBs has proved challenging. Therefore, exogenous ketone supplementation is a means to address these questions and explore potential for performance and therapeutic benefits.

Oral administration of KBs in their free acid form is expensive and ineffective at producing ketosis, so buffering the free acid form with sodium/potassium/calcium salts has been explored and these compounds are commercially available. These too are relatively ineffective at increasing [β HB], but may be improved by co-ingestion with medium chain triglycerides (C:8, C:10), at least in rats (Kesi *et al.* 2016). However, ingestion of large quantities of KB salts is impractical due to resulting gastrointestinal distress, and potentially undesirable consequences of cation overload or acidosis (Veech, 2004).

The development of ketone esters provides an alternative method to increase [β HB], which is well-tolerated in rodents and humans (Clarke *et al.* 2012; Cox *et al.* 2016; Kesi *et al.* 2016). Two prominent ketone esters in the published literature are the *R,S*-1,3-butanediol acetoacetate diester (Kesi *et al.* 2016) and the (*R*)-3-hydroxybutyl (*R*)-3-hydroxybutyrate ketone monoester (Clarke *et al.* 2012; Cox *et al.* 2016). Acute ingestion of either ester can result in short-term (~0.5 to 6 h) nutritional ketosis indicated by [β HB] >1 mM (Clarke *et al.* 2012; Kesi *et al.* 2016). For the ketone monoester, ingestion at a dose of 573 mg (kg body mass (BM))⁻¹ resulted in [β HB] of ~3 mM after 10 min and rising to ~6 mM 30 min after ingestion (Cox *et al.* 2016). Nutritional ketosis is therefore achieved without the impracticality of prolonged fasting or ketogenic dieting.

Ketone body metabolism during exercise under conditions of exogenous ketosis. The aforementioned self-inhibitory effect of rising ketonaemia underscores a key methodological issue when considering KB metabolism in skeletal muscle, namely the method of achieving ketosis. While fasting of various durations is a widely used model of ketosis, acute nutritional ketosis relevant to sports performance would be achieved with replete glycogen stores, and in the absence of prolonged elevations in FFAs and [KB] that would be likely to impair KB oxidation rates through these mechanisms. To our knowledge, only two studies have addressed this convincingly by examining effects of exercise on KB metabolism without interference from the various hormonal and metabolic perturbations associated with prolonged fasting or diabetes (Fery & Balasse, 1988; Cox *et al.* 2016).

In the former study (Fery & Balasse, 1988), infusion of sodium AcAc after an overnight fast achieved [KB]

of ~ 6 mM (β HB ~ 3.5 mM, AcAc ~ 2.5 mM) at the onset of 2 h of exercise at $\sim 52\%$ $\dot{V}_{O_{2\max}}$. Notably, AcAc did not change during exercise whereas β HB declined throughout exercise, to be reduced by ~ 2 mM at the end of exercise. This coincided with a progressive rise in MCR throughout exercise, peaking at $\sim 75\%$ higher than rest at the end of exercise. In contrast, this effect was abolished with similar ketonaemia in 3–5 day fasted participants. Importantly, although the inhibition of KB oxidation by hyperketonaemia is present during exogenous ketosis, an 'auto-amplification' was noted that is not present in fasting ketosis, i.e. the initial rise in MCR induced by exercise causes a reduction in concentration which, in turn, provokes a further rise in MCR and so on. Additionally, the threshold concentration at which hyperketonaemia inhibits MCR was higher in exogenous ketosis than in fasting ketosis. However, in terms of contribution to energy provision, this ultimately only resulted in a 2% contribution over the 2 h exercise bout. Nevertheless, plasma [lactate] did not rise during exercise after AcAc infusion compared to a ~ 1 mM rise in the fasted participants, which suggests that despite a modest contribution to energy provision, exogenous ketosis can impact on metabolic processes during exercise.

Despite this promise, these data remained largely isolated for almost 30 years with the exception of a couple of obscure reports that admittedly did recapitulate the effects of β HB to alter the metabolic response to very intense exercise in rats (Kamysheva & Ostrovskaya, 1980), and ischaemic exercise in humans (Lestan *et al.* 1994). The latter report, in fact, supported the ability of a modest elevation in β HB (~ 0.5 mM) via infusion of sodium β HB to reduce the plasma lactate response to exercise in an ischaemic forearm model. However, with the development of the (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester (KE), a comprehensive investigation of substrate metabolism in highly trained athletes in the presence of acute nutritional ketosis has recently been published (Cox *et al.* 2016).

In one of a series of experiments, ingestion of KE resulted in acute nutritional ketosis indicated by [β HB] of ~ 3 mM after 10 min and rising to ~ 6 mM 30 min after ingestion. During exercise lasting 45 min at either 40% or 75% \dot{W}_{\max} , [β HB] was ~ 2 and 3 mM, respectively, lower than ketosis produced after ingestion at rest. This provided the first evidence of intensity-dependent disposal of β HB during exercise. Moreover, based on expired air analysis adjusted for oxidation of KBs, β HB oxidation contributed 18% and 16% of oxygen consumption to energy provision at the respective intensities. The larger than previously reported contribution of β HB oxidation probably reflects the fact that the participants in these experiments were highly trained cyclists, therefore with a greater capacity of skeletal muscle to uptake and oxidise KBs. Moreover, this model is markedly different to the fasting-induced

ketonaemia and the associated self-inhibitory regulation so making direct comparisons are difficult. In a separate exercise bout lasting 60 min at 75% \dot{W}_{\max} and with similar [β HB] after KE ingestion, the rise in plasma [lactate] was blunted by ~ 2 to 3 mM ($\sim 50\%$ reduction) compared to ingestion of an isocaloric CHO drink. Subsequent experiments with ingestion of the KE demonstrated inhibition of glycolytic metabolism, sparing of muscle glycogen, reduced deamination of branched-chain amino acids, and increased reliance on IMTG during exercise (Cox *et al.* 2016). Lastly, after a 60 min pre-load at 75% \dot{W}_{\max} , cycling performance in a 30 min time-trial was improved by 2% (411 ± 162 m; mean \pm SEM, $n = 8$) with KE + CHO compared to isocaloric CHO ingestion. The KE + CHO fuelling strategy combined KE (40%; 573 mg (kg BM) $^{-1}$) with CHO (60%) and elevated [β HB] to between ~ 1.5 and 3 mM throughout. Importantly, the KE + CHO condition provided CHO at a minimum rate of 1.2 g min $^{-1}$, consistent with an optimal CHO-based fuelling strategy (Burke, 2015). Taken together, these data suggest that acute nutritional ketosis by consumption of exogenous ketones has dramatic effects on skeletal muscle metabolism during exercise, and can confer a performance benefit to elite athletes (Fig. 3). The positive findings notwithstanding, potential adverse effects should be considered for any performance aid prior to adoption. Side-effects of KE ingestion have been reported in humans (Clarke *et al.* 2012). Specifically, in a repeated dose design over 5 days, adverse effects such as flatulence, nausea, diarrhoea and dizziness were reported in five out of twenty-four participants at doses ranging from 420 to 1071 mg (kg BM) $^{-1}$. Such issues were prevalent in almost all participants when the dose was increased to 2142 mg (kg BM) $^{-1}$ per day, indicating a possible upper limit of tolerability in adults (Clarke *et al.* 2012). Therefore, these data combined with the dosing strategy associated with exercise performance benefits should be used to guide future investigations on ergogenic potential.

Ketone body metabolism after exercise: post-exercise ketosis

Despite the aforementioned decline in [KB] at the onset of exercise, this pertains to situations where exercise has begun during hyperketonaemia (Balasse *et al.* 1978; Fery & Balasse, 1983, 1988; Cox *et al.* 2016). In the post-absorptive state, the pattern generally observed is for [KB] to rise gradually during prolonged exercise up to 0.2 to 0.4 mM, after which time post-exercise ketosis (PEK) of 0.3 to 2.0 mM is observed for several hours into recovery (Koeslag, 1982). Explained in terms of plasma kinetics, at cessation of exercise, the rate of appearance of KBs increases coincident with a decrease in MCR relative to rates present during exercise. MCR remains above resting

values for several hours after exercise, but ketogenesis exceeds ketolysis during this period.

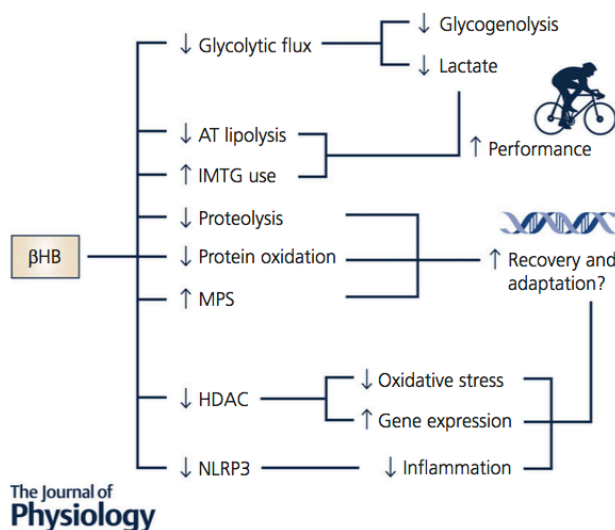
On a mechanistic level, regulation probably resides at several sites including malonyl CoA-mediated regulation of fat transport into hepatocytes via CPT-1 in addition to availability of Ac-CoA for ketogenesis, and oxaloacetate for the TCA cycle as classically described for ketogenic regulation. Because oxaloacetate is a product of pyruvate formed during glycolysis, reductions in glycolytic flux with low glycogen content after intense exercise result in oxaloacetate moving to cytoplasm for preferential use in gluconeogenesis, which allows diversion of Ac-CoA towards ketogenesis during the post-exercise period rather than to citrate synthesis for the TCA cycle. Additionally, the actions of insulin and glucagon exert a strong influence through activation and inhibition, respectively, of Ac-CoA carboxylase (ACC), which catalyses the synthesis of malonyl CoA from Ac-CoA. When liver glycogen becomes depleted and glucagon:insulin ratio is elevated, the synthesis of malonyl CoA is reduced, thereby relieving the inhibition of fat transport into hepatocytes, and resulting in elevated levels of Ac-CoA. These regulatory mechanisms are acutely sensitive to nutrient manipulations before and after exercise and to aerobic exercise training, given their respective influences on substrate availability and utilisation during exercise.

Modulation of post-exercise ketosis by aerobic exercise training and nutrition intervention. An attenuation of, or abolished, post-exercise ketosis has been consistently

observed in rodents and humans in response to aerobic exercise in trained *versus* untrained individuals (Johnson *et al.* 1969; Johnson & Walton, 1972; Rennie *et al.* 1974), or after a period of exercise training (Rennie & Johnson, 1974a; Beattie & Winder, 1984, 1985; Adams & Koeslag, 1988, 1989; Ohmori *et al.* 1990). The aforementioned enhanced ketolytic capacity and down-regulation of ketogenic capacity by training may play a role in these observations, but the majority of this work has been performed in comparisons, with the absolute exercise intensity and duration being the same for comparisons (reviewed in Koeslag, 1982). This is problematic because the relative exercise intensity is the key determinant of the metabolic and hormonal response to acute exercise, e.g. catecholamine responses, FFA mobilisation and glycogen utilisation among others. When trained and untrained participants have performed exercise at a similar relative intensity, PEK is blunted but not abolished in trained individuals (Rennie *et al.* 1974). Moreover, in rodents when exercise is completed to exhaustion, i.e. the trained rats exercise for longer than untrained, [β HB] is ~twofold higher at the exercise cessation in the trained group (Askew *et al.* 1975). These divergent findings are likely to be due to the degree of liver glycogen depletion that occurs (Adams & Koeslag, 1988), inasmuch as higher levels of resting liver glycogen and attenuated rates of depletion are a consequence of training (Baldwin *et al.* 1975).

Therefore, PEK is strongly influenced by nutrition manipulation. High CHO feeding prior to exercise attenuates PEK regardless of training status (Rennie &

Figure 3. β HB as a metabolic regulator and signalling metabolite
Effects of elevating β HB through acute nutritional ketosis may be mediated by acute regulation of substrate utilisation that may enhance performance, and/or possibly through regulation of recovery and adaptive processes related to inflammation, oxidative stress and changes in gene expression. See text for further discussion. AT, adipose tissue; HDAC, histone deacetylase; IMTG, intramuscular triglyceride; MPS, muscle protein synthesis.



Johnson, 1974b; Askew *et al.* 1975; Koeslag *et al.* 1980), and CHO restriction increases PEK (Impey *et al.* 2016). Glucose ingestion at 2 h into recovery (Koeslag *et al.* 1982; Carlin *et al.* 1987) and alanine during recovery (Koeslag *et al.* 1980, 1985; Carlin *et al.* 1987) attenuate PEK, but the glucose effect is not seen when glucose is ingested immediately after exercise. Alanine ingestion increases mitochondrial [oxaloacetate] in liver, thereby allowing condensation with Ac-CoA and diversion away from ketogenesis. This suggests that the early PEK response is determined by the extent of liver glycogen depletion and reduced glycolytic flux, whereas several hours into recovery it is under regulation by insulin and [FFA] related to nutrition intake.

Metabolic consequences of post-exercise ketosis during recovery: a role for exogenous ketones as a recovery aid? The physiological role for PEK is likely to favour the replenishment of muscle glycogen, consistent with classically described metabolic actions of ketosis in the sparing of protein and CHO stores during times of low CHO availability. During the post-exercise recovery period, in contrast to the reliance on CHO metabolism during exercise, muscle glycogen resynthesis has a high metabolic priority and is facilitated by an increase in fat oxidation and sparing of CHO sources for energy provision (Kiens & Richter, 1998). A priority for muscle glycogen resynthesis over liver glycogen resynthesis is suggested to occur because in ancestral terms, a depleted liver is less of a hindrance to intense exertion than depleted muscle (Adams & Koeslag, 1988). To this end, the priority for muscle glycogen resynthesis is observed even during CHO restriction (Adams & Koeslag, 1989), and is achieved through non-CHO sources such as lactate and alanine being used for hepatic gluconeogenesis and redistribution to skeletal muscle (Fournier *et al.* 2002). The contribution of PEK may be via the ability of KBs to inhibit glycolysis and increase the conversion of glucose to glycogen as demonstrated in rat skeletal muscle *in vitro* (Maizels *et al.* 1977), and a perfused heart model in dogs (Laughlin *et al.* 1994). This effect is likely to be mediated by inhibition of PDH and phosphofructokinase (PFK) by elevations in Ac-CoA and citrate formation, respectively, as a consequence of metabolism of AcAc in mitochondria (Randle *et al.* 1964; Maizels *et al.* 1977; Laughlin *et al.* 1994; Kashiwaya *et al.* 1997).

This raises the possibility that an optimal post-exercise recovery milieu exists that includes both CHO and ketones to enhance recovery of muscle glycogen. This is not possible by conventional nutrition strategies because elevations in glucose, lactate and alanine ultimately limit ketogenesis and PEK. The suggestion is that the co-ingestion of exogenous ketones and CHO in a recovery protocol can confer a metabolic advantage. This hypothesis remains to be tested rigorously, but a

preliminary report describes a 33% increase in glucose disposal and 50% increase in muscle glycogen content after 2 h of recovery when nutritional ketosis (~5 mM β HB) is superimposed on a hyperglycaemic (10 mM glucose) clamp in well-trained military servicemen (Holdsworth *et al.* 2016).

Repletion of muscle glycogen is only one component of post-exercise recovery, and nutrition strategies for recovery include protein ingestion, with the aim to limit muscle protein breakdown and enhance muscle protein synthesis (MPS). KBs have protein sparing effects in skeletal muscle as indicated by reduced alanine release during starvation (Sherwin *et al.* 1975), and reduced leucine oxidation (Nair *et al.* 1988). In the latter study, this coincided with a 10% increase in MPS measured by fractional synthesis rate and occurred with [β HB] of ~2 mM achieved via sodium β HB infusion. This raises the possibility that acute nutritional ketosis can complement current strategies for optimising MPS in the post-exercise period. Additionally, because low CHO stores during exercise lead to elevated rates of protein oxidation (Wagenmakers *et al.* 1991), exogenous ketone supplementation may provide both a fuel source and contribute to protein sparing and recovery during training in CHO-restricted states commonly practiced by athletes (reviewed in Bartlett *et al.* 2015). Together with the preliminary data for muscle glycogen resynthesis, this suggests that post-exercise recovery is another application where elite athletes may benefit from exogenous ketone supplementation, and where future research is warranted.

Effects beyond fuelling: β HB as a HDAC inhibitor

As investigative techniques in molecular biology evolve, so too does our appreciation of how complex integrative signalling networks regulate skeletal muscle adaptation in response to stimuli such as nutrient manipulation and exercise training (Egan & Zierath, 2013). Previously considered relatively inert outside their primary metabolic function, numerous substrates and metabolites are emerging as important regulators of intracellular signalling and tissue adaptation (Hashimoto *et al.* 2007; Gao *et al.* 2009; Morton *et al.* 2009; Roberts *et al.* 2014). Noteworthy for the present review is the recent identification of AcAc as a regulator of skeletal muscle satellite cell proliferation and muscle regeneration (Zou *et al.* 2016), and β HB as an inhibitor of HDACs (Shimazu *et al.* 2013) and the NLRP3 inflammasome (Youn *et al.* 2015). The latter observations are a consequence of β HB, in essence, acting as a signalling metabolite to regulate gene expression and metabolic processes (Fig. 3).

Histone acetyltransferases (HATs) and HDACs are enzymes that facilitate the addition or removal, respectively, of acetyl moieties from specific lysine residues on histones and target proteins (McKinsey

et al. 2001). In general, hyperacetylation of histone tails induces transcriptional activation while hypoacetylation is associated with transcriptional repression. Class IIa HDACs (HDAC4, -5, -7 and -9) are highly expressed in skeletal muscle (McKinsey *et al.* 2001) and their function is responsive to both aerobic endurance exercise in humans (McGee *et al.* 2009; Egan *et al.* 2010) and nutritional intervention in rodents (Gao *et al.* 2009; Shimazu *et al.* 2013). An acute bout of aerobic exercise increases class IIa HDAC phosphorylation and subsequent nuclear exclusion, thus inhibiting HDAC-mediated repression of specific exercise-responsive genes such as GLUT4 and PGC-1 α (McGee & Hargreaves, 2004; McGee *et al.* 2009; Egan *et al.* 2010). This suggests that compounds that inhibit or disrupt HDAC inhibition could be used to mimic or enhance adaptations to exercise.

Regulation of HDAC activity by nutrients including butyrate and β HB has also been established (Gao *et al.* 2009; Shimazu *et al.* 2013). Butyrate, a short chain fatty acid formed via the fermentation of indigestible dietary fibres by microbial species in the gut, is a potent inhibitor of HDAC activity (Gao *et al.* 2009). Mice supplemented with sodium butyrate are resistant to diet-induced obesity, and have elevations in markers of skeletal muscle mitochondrial biogenesis analogous to exercise effects (Gao *et al.* 2009). β HB is structurally similar to butyrate, and although not as potent as butyrate, also inhibits HDAC class I and II activity in a dose-dependent manner and suppressed oxidative stress responses (Shimazu *et al.* 2013). Importantly, HDAC inhibition by β HB both *in vitro* and *in vivo* is evident at physiologically relevant concentrations of β HB, i.e. 1 to 4 mM, which is similar to those attained during fasting, PEK and exogenous ketone ingestion (Fig. 1; Clarke *et al.* 2012; Kesl *et al.* 2016). However, although the inhibitory effects were observed in multiple tissues, they remain to be confirmed in skeletal muscle. If confirmed, it will be intriguing to explore whether, apart from the aforementioned ergogenic effects, exogenous ketone supplementation complements exercise-mediated adaptive changes associated with modulating HDAC function (Fig. 3).

Exogenous ketone supplementation for athletes: cautionary notes and future directions

Despite a strong physiological basis for a variety of benefits for performance and recovery, the relatively recent availability of exogenous ketones and thus far only one peer-reviewed paper examining exercise metabolism, performance and nutritional ketosis, means that much more research remains to be performed (Pinckaers *et al.* 2016). The central tenet is that the combination of fuel sparing and improved energetic efficiency during acute nutritional ketosis confers performance benefits (Fig. 3). Alterations in fuel selection during steady-state

exercise have been demonstrated, which indicate reduced glycolytic flux, sparing of CHO and increased contribution of IMTG and β HB to energy provision (Cox *et al.* 2016). Whether this sparing of CHO, in fact, manifests as *impaired* CHO utilisation remains to be determined. The mechanistic basis for CHO sparing by exogenous ketones is presently proposed as inhibition of glycolytic flux via inhibition of PDH and PFK by increases in NADH:NAD⁺, acetyl-CoA:CoA ratio or citrate. In theory, this could be problematic for sports that rely heavily on contributions from glycolytic pathways, or a range of sports that are intermittent and/or require periods of high intensity 'bursts' on a moderate intensity background. This is analogous to the lack of performance benefits for most athletes undertaking low CHO, high fat diets (Burke, 2015). In fact, impaired performance during high intensity efforts has been observed under such conditions (Havemann *et al.* 2006), and may be explained by sustained attenuation of PDH activity (Stellingwerff *et al.* 2006). Whether the same effects are observed with acute nutritional ketosis given that this is a very different metabolic milieu, especially in the context of exercise, remains to be explored.

The metabolic consequences of inhibition of adipose tissue lipolysis by KBs also warrants further exploration, given that this process is an important contributor to circulating FFAs, and therefore to the contribution of fat oxidation to energy provision during long duration, sub-maximal exercise. Nutritional ketosis achieved by either AcAc infusion (Fery & Balasse, 1988) or KE ingestion (Cox *et al.* 2016) inhibits the lipolytic effect of exercise, i.e. the amount of lipid-derived substrates available for working muscle is reduced. In the latter study, this did not manifest as increased glycogenolysis and/or glucose utilization, despite these usually being accelerated by the inhibition of FFA availability (van Loon *et al.* 2005). In fact, glycogenolysis was attenuated and IMTG utilisation was increased in the KE experiments (Cox *et al.* 2016), suggesting differential regulation to that achieved by nicotinic acid administration (van Loon *et al.* 2005). However, in each of the experimental conditions with KE, the duration of exercise was between 45 and 120 min at moderate intensity (Cox *et al.* 2016). Recently, the inhibition of lipolysis via nicotinic acid impaired cycling time-trial performance in long (120 min), but not shorter (60 and 90 min) duration efforts (Torrens *et al.* 2016). Thus, even in events with high CHO dependence (~80 to 95% of energy provision), inhibition of lipolysis may impair endurance performance, particularly in long duration activities analogous to professional cycling or triathlon. Clearly, the many nodes of metabolic regulation influencing skeletal muscle fuel selection that are altered by nutritional ketosis need to be fully elucidated before sports-specific ergogenic strategies can be advised.

Improved energetic efficiency is an often-cited potential benefit of acute nutritional ketosis (Veech, 2004; Cox & Clarke, 2014). In this model, exogenous ketones may provide thermodynamic advantages over CHO and fat, because the available free energy to perform work, the free energy of ATP hydrolysis (ΔG_{ATP}), is greater with KBs, and require less oxygen per mole of carbon to oxidise. Support for this hypothesis comes from a perfused working rat heart model where adding KBs to the perfusate suppressed glycolytic flux, and increased hydraulic efficiency (expressed as work in J (mol O₂ consumed)⁻¹) by 28% (Sato *et al.* 1995; Kashiwaya *et al.* 1997). In practical terms, if the same effect occurs in skeletal muscle, this would translate as a higher power output for the same oxygen consumption (i.e. improved muscular efficiency) during exercise with nutritional ketosis, but this remains unexplored at present.

Because KBs serve as a substrate for the brain, and therapeutic uses for KBs have been proposed for cognitive enhancement and neurodegenerative pathologies (Veech, 2004), the central nervous system (CNS) may be another target for performance-enhancing effects of nutritional ketosis. Although speculative at present, effects related to motor recruitment, perceived exertion, pacing strategies, skill execution, reaction time, and decision-making will be interesting for future research, in addition to the proposed role for the CNS in regulating performance beyond effects related to skeletal muscle metabolism (Noakes, 2011).

As with any ergogenic aid or nutrition strategy, optimising dosing strategies including quantity and timing will be important. Given the saturation kinetics of KB oxidation by skeletal muscle and curvilinear relationship between oxidation and plasma concentrations, it is likely that there is an optimal range for performance benefits. At present, we speculate that this exists between 1 and 3 mM β HB. As with many ergogenic acids, more is unlikely to be better and may even be deleterious given the potential for acidosis at higher [KB], and aforementioned gastrointestinal distress and other side-effects sometimes observed with KE, so careful consideration should be given to these issues.

In conclusion, although data are preliminary, acute nutritional ketosis achieved by exogenous ketone supplementation has the potential to alter fuel selection during exercise and confer performance benefits. This is most likely to be the case in trained individuals who have a greater capacity to take up and oxidise KBs during exercise as a result of training. Additionally, a strong physiological basis exists that suggests potential benefits for supporting training and recovery. While much work remains to be performed, particularly in relation to sport-specific strategies, this promises to be an exciting topic for scientists, practitioners and athletes alike for the coming years.

References

- Abraham R (2015). Ketones: Controversial new energy drink could be next big thing in cycling. *Cycling Weekly*.
- Adams JH & Koeslag JH (1988). Carbohydrate homeostasis and post-exercise ketosis in trained and untrained rats. *J Physiol* **407**, 453–461.
- Adams JH & Koeslag JH (1989). Glycogen metabolism and post-exercise ketosis in carbohydrate-restricted trained and untrained rats. *Q J Exp Physiol* **74**, 27–34.
- Askew EW, Dohm GL & Huston RL (1975). Fatty acid and ketone body metabolism in the rat: response to diet and exercise. *J Nutr* **105**, 1422–1432.
- Balasse EO & Fery F (1989). Ketone body production and disposal: effects of fasting, diabetes, and exercise. *Diabetes Metab Rev* **5**, 247–270.
- Balasse EO, Fery F & Neef MA (1978). Changes induced by exercise in rates of turnover and oxidation of ketone bodies in fasting man. *J Appl Physiol Respir Environ Exerc Physiol* **44**, 5–11.
- Baldwin KM, Fitts RH, Booth FW, Winder WW & Holloszy JO (1975). Depletion of muscle and liver glycogen during exercise. Protective effect of training. *Pflugers Arch* **354**, 203–212.
- Bartlett JD, Hawley JA & Morton JP (2015). Carbohydrate availability and exercise training adaptation: too much of a good thing? *Eur J Sport Sci* **15**, 3–12.
- Beattie MA & Winder WW (1984). Mechanism of training-induced attenuation of postexercise ketosis. *Am J Physiol Regul Integr Comp Physiol* **247**, R780–R785.
- Beattie MA & Winder WW (1985). Attenuation of postexercise ketosis in fasted endurance-trained rats. *Am J Physiol Regul Integr Comp Physiol* **248**, R63–R67.
- Bonen A (2001). The expression of lactate transporters (MCT1 and MCT4) in heart and muscle. *Eur J Appl Physiol* **86**, 6–11.
- Bosch AN, Weltan SM, Dennis SC & Noakes TD (1996). Fuel substrate kinetics of carbohydrate loading differs from that of carbohydrate ingestion during prolonged exercise. *Metabolism* **45**, 415–423.
- Burke LM (2015). Re-examining high-fat diets for sports performance: did we call the 'nail in the coffin' too soon? *Sports Med* **45** (Suppl. 1), S33–49.
- Carlini JJ, Olson EB Jr, Peters HA & Reddan WG (1987). The effects of post-exercise glucose and alanine ingestion on plasma carnitine and ketosis in humans. *J Physiol* **390**, 295–303.
- Cermak NM & van Loon LJ (2013). The use of carbohydrates during exercise as an ergogenic aid. *Sports Med* **43**, 1139–1155.
- Clarke K, Tchabanenko K, Pawlosky R, Carter E, Todd King M, Musa-Veloso K, Ho M, Roberts A, Robertson J, Vanitallie TB & Veech RL (2012). Kinetics, safety and tolerability of (R)-3-hydroxybutyl (R)-3-hydroxybutyrate in healthy adult subjects. *Regul Toxicol Pharmacol* **63**, 401–408.
- Cox PJ & Clarke K (2014). Acute nutritional ketosis: implications for exercise performance and metabolism. *Extrem Physiol Med* **3**, 17.

- Cox PJ, Kirk T, Ashmore T, Willerton K, Evans R, Smith A, Murray AJ, Stubbs B, West J, McLure SW, King MT, Dodd MS, Holloway C, Neubauer S, Drawer S, Veech RL, Griffin JL & Clarke K (2016). Nutritional ketosis alters fuel preference and thereby endurance performance in athletes. *Cell Metab* **24**, 256–268.
- Egan B, Carson BP, Garcia-Roves PM, Chibalin AV, Sarsfield FM, Barron N, McCaffrey N, Moyna NM, Zierath JR & O’Gorman DJ (2010). Exercise intensity-dependent regulation of peroxisome proliferator-activated receptor γ coactivator-1 α mRNA abundance is associated with differential activation of upstream signalling kinases in human skeletal muscle. *J Physiol* **588**, 1779–1790.
- Egan B & Zierath JR (2013). Exercise metabolism and the molecular regulation of skeletal muscle adaptation. *Cell Metab* **17**, 162–184.
- El Midaoui A, Chiasson JL, Tancrede G & Nadeau A (2006). Physical training reverses the increased activity of the hepatic ketone body synthesis pathway in chronically diabetic rats. *Am J Physiol Endocrinol Metab* **290**, E207–E212.
- Elia M, Wood S, Khan K & Pulicino E (1990). Ketone body metabolism in lean male adults during short-term starvation, with particular reference to forearm muscle metabolism. *Clin Sci (Lond)* **78**, 579–584.
- Fery F & Balasse EO (1983). Ketone body turnover during and after exercise in overnight-fasted and starved humans. *Am J Physiol Endocrinol Metab* **245**, E318–E325.
- Fery F & Balasse EO (1986). Response of ketone body metabolism to exercise during transition from postabsorptive to fasted state. *Am J Physiol Endocrinol Metab* **250**, E495–E501.
- Fery F & Balasse EO (1988). Effect of exercise on the disposal of infused ketone bodies in humans. *J Clin Endocrinol Metab* **67**, 245–250.
- Fournier PA, Brau L, Ferreira LD, Fairchild T, Raja G, James A & Palmer TN (2002). Glycogen resynthesis in the absence of food ingestion during recovery from moderate or high intensity physical activity: novel insights from rat and human studies. *Comp Biochem Physiol A Mol Integr Physiol* **133**, 755–763.
- Frayn KN (1983). Calculation of substrate oxidation rates *in vivo* from gaseous exchange. *J Appl Physiol* **55**, 628–634.
- Gao Z, Yin J, Zhang J, Ward RE, Martin RJ, Lefevre M, Cefalu WT & Ye J (2009). Butyrate improves insulin sensitivity and increases energy expenditure in mice. *Diabetes* **58**, 1509–1517.
- Hagenfeldt L & Wahren J (1968). Human forearm muscle metabolism during exercise. 3. Uptake, release and oxidation of beta-hydroxybutyrate and observations on the beta-hydroxybutyrate/acetate ratio. *Scand J Clin Lab Invest* **21**, 314–320.
- Hagenfeldt L & Wahren J (1971). Human forearm muscle metabolism during exercise. VI. Substrate utilization in prolonged fasting. *Scand J Clin Lab Invest* **27**, 299–306.
- Hashimoto T, Hussien R, Oommen S, Gohil K & Brooks GA (2007). Lactate sensitive transcription factor network in L6 cells: activation of MCT1 and mitochondrial biogenesis. *FASEB J* **21**, 2602–2612.
- Havemann L, West SJ, Goedecke JH, Macdonald IA, St Clair Gibson A, Noakes TD & Lambert EV (2006). Fat adaptation followed by carbohydrate loading compromises high-intensity sprint performance. *J Appl Physiol* (1985) **100**, 194–202.
- Holdsworth D, Cox PJ & Clarke K (2016). Oral ketone body supplementation accelerates and enhances glycogen synthesis in human skeletal muscle following exhaustive exercise [Abstract]. In *Proceedings of the Physiological Society*. London, UK.
- Impey SG, Hammond KM, Shepherd SO, Sharples AP, Stewart C, Limb M, Smith K, Philp A, Jeromson S, Hamilton DL, Close GL & Morton JP (2016). Fuel for the work required: a practical approach to amalgamating train-low paradigms for endurance athletes. *Physiol Rep* **4**, e12803.
- Johnson RH & Walton JL (1972). The effect of exercise upon acetoacetate metabolism in athletes and non-athletes. *Q J Exp Physiol Cogn Med Sci* **57**, 73–79.
- Johnson RH, Walton JL, Krebs HA & Williamson DH (1969). Metabolic fuels during and after severe exercise in athletes and non-athletes. *Lancet* **2**, 452–455.
- Kamysheva VA & Ostrovskaia RU (1980). [Effect of sodium hydroxybutyrate on the ammonia level in the rat muscles under physical exercise]. *Biull Eksp Biol Med* **89**, 25–27.
- Kashiwaya Y, King MT & Veech RL (1997). Substrate signaling by insulin: a ketone bodies ratio mimics insulin action in heart. *Am J Cardiol* **80**, 50a–64a.
- Kesl SL, Poff AM, Ward NP, Fiorelli TN, Ari C, Van Putten AJ, Sherwood JW, Arnold P & D’Agostino DP (2016). Effects of exogenous ketone supplementation on blood ketone, glucose, triglyceride, and lipoprotein levels in Sprague-Dawley rats. *Nutr Metab (Lond)* **13**, 9.
- Kiens B & Richter EA (1998). Utilization of skeletal muscle triacylglycerol during postexercise recovery in humans. *Am J Physiol Endocrinol Metab* **275**, E332–E337.
- Koeslag JH (1982). Post-exercise ketosis and the hormone response to exercise: a review. *Med Sci Sports Exerc* **14**, 327–334.
- Koeslag JH, Levinrad LI, Lochner JD & Sive AA (1985). Post-exercise ketosis in post-prandial exercise: effect of glucose and alanine ingestion in humans. *J Physiol* **358**, 395–403.
- Koeslag JH, Noakes TD & Sloan AW (1980). Post-exercise ketosis. *J Physiol* **301**, 79–90.
- Koeslag JH, Noakes TD & Sloan AW (1982). The effects of alanine, glucose and starch ingestion on the ketosis produced by exercise and by starvation. *J Physiol* **325**, 363–376.
- Laffel L (1999). Ketone bodies: a review of physiology, pathophysiology and application of monitoring to diabetes. *Diabetes Metab Res Rev* **15**, 412–426.
- Laughlin MR, Taylor J, Chesnick AS & Balaban RS (1994). Nonglucose substrates increase glycogen synthesis *in vivo* in dog heart. *Am J Physiol Heart Circ Physiol* **267**, H219–H223.
- Lestan B, Walden K, Schmaltz S, Spyckala J & Fox IH (1994). β -Hydroxybutyrate decreases adenosine triphosphate degradation products in human subjects. *J Lab Clin Med* **124**, 199–209.

- McGee SL, Fairlie E, Garnham AP & Hargreaves M (2009). Exercise-induced histone modifications in human skeletal muscle. *J Physiol* **587**, 5951–5958.
- McGee SL & Hargreaves M (2004). Exercise and myocyte enhancer factor 2 regulation in human skeletal muscle. *Diabetes* **53**, 1208–1214.
- McKinsey TA, Zhang CL & Olson EN (2001). Control of muscle development by dueling HATs and HDACs. *Curr Opin Genet Dev* **11**, 497–504.
- Maizels EZ, Ruderman NB, Goodman MN & Lau D (1977). Effect of acetoacetate on glucose metabolism in the soleus and extensor digitorum longus muscles of the rat. *Biochem J* **162**, 557–568.
- Mazzeo RS, Brooks GA, Schoeller DA & Budinger TF (1986). Disposal of blood [^{1-13}C]lactate in humans during rest and exercise. *J Appl Physiol* (1985) **60**, 232–241.
- Mikkelsen KH, Seifert T, Secher NH, Grondal T & van Hall G (2015). Systemic, cerebral and skeletal muscle ketone body and energy metabolism during acute hyper-D-beta-hydroxybutyrate in post-absorptive healthy males. *J Clin Endocrinol Metab* **100**, 636–643.
- Morton JP, Croft L, Bartlett JD, Maclaren DP, Reilly T, Evans L, McArdle A & Drust B (2009). Reduced carbohydrate availability does not modulate training-induced heat shock protein adaptations but does upregulate oxidative enzyme activity in human skeletal muscle. *J Appl Physiol* **106**, 1513–1521.
- Nair KS, Welle SL, Halliday D & Campbell RG (1988). Effect of beta-hydroxybutyrate on whole-body leucine kinetics and fractional mixed skeletal muscle protein synthesis in humans. *J Clin Invest* **82**, 198–205.
- Noakes TD (2011). Time to move beyond a brainless exercise physiology: the evidence for complex regulation of human exercise performance. *Appl Physiol Nutr Metab* **36**, 23–35.
- Ohmori H, Kawai K & Yamashita K (1990). Enhanced ketone body uptake by perfused skeletal muscle in trained rats. *Endocrinol Jpn* **37**, 421–429.
- Owen OE & Reichard GA Jr (1971). Human forearm metabolism during progressive starvation. *J Clin Invest* **50**, 1536–1545.
- Paoli A, Rubini A, Volek JS & Grimaldi KA (2013). Beyond weight loss: a review of the therapeutic uses of very-low-carbohydrate (ketogenic) diets. *Eur J Clin Nutr* **67**, 789–796.
- Pinckaers PJ, Churchward-Venne TA, Bailey D & van Loon LJ (2016). Ketone bodies and exercise performance: the next magic bullet or merely hype? *Sports Med.* (in press; DOI: 10.1007/s40279-016-0577-y).
- Randle PJ, Newsholme EA & Garland PB (1964). Regulation of glucose uptake by muscle. 8. Effects of fatty acids, ketone bodies and pyruvate, and of alloxan-diabetes and starvation, on the uptake and metabolic fate of glucose in rat heart and diaphragm muscles. *Biochem J* **93**, 652–665.
- Rennie MJ, Jennett S & Johnson RH (1974). The metabolic effects of strenuous exercise: a comparison between untrained subjects and racing cyclists. *Q J Exp Physiol Cogn Med Sci* **59**, 201–212.
- Rennie MJ & Johnson RH (1974a). Alteration of metabolic and hormonal responses to exercise by physical training. *Eur J Appl Physiol Occup Physiol* **33**, 215–226.
- Rennie MJ & Johnson RH (1974b). Effects of an exercise-diet program on metabolic changes with exercise in runners. *J Appl Physiol* **37**, 821–825.
- Roberts LD, Bostrom P, O'Sullivan JF, Schinzel RT, Lewis GD, Dejam A, Lee YK, Palma MJ, Calhoun S, Georgiadi A, Chen MH, Ramachandran VS, Larson MG, Bouchard C, Rankinen T, Souza AL, Clish CB, Wang TJ, Estall JL, Soukas AA, Cowan CA, Spiegelman BM & Gerszten RE (2014). β -Aminoisobutyric acid induces browning of white fat and hepatic β -oxidation and is inversely correlated with cardiometabolic risk factors. *Cell Metab* **19**, 96–108.
- Robinson AM & Williamson DH (1980). Physiological roles of ketone bodies as substrates and signals in mammalian tissues. *Physiol Rev* **60**, 143–187.
- Sato K, Kashiwaya Y, Keon CA, Tsuchiya N, King MT, Radda GK, Chance B, Clarke K & Veech RL (1995). Insulin, ketone bodies, and mitochondrial energy transduction. *FASEB J* **9**, 651–658.
- Sherwin RS, Hendler RG & Felig P (1975). Effect of ketone infusions on amino acid and nitrogen metabolism in man. *J Clin Invest* **55**, 1382–1390.
- Shimazu T, Hirschey MD, Newman J, He W, Shirakawa K, Le Moan N, Grueter CA, Lim H, Saunders LR, Stevens RD, Newgard CB, Farese RV Jr, de Cabo R, Ulrich S, Akassoglou K & Verdin E (2013). Suppression of oxidative stress by β -hydroxybutyrate, an endogenous histone deacetylase inhibitor. *Science* **339**, 211–214.
- Stellingwerff T, Spriet LL, Watt MJ, Kimber NE, Hargreaves M, Hawley JA & Burke LM (2006). Decreased PDH activation and glycogenolysis during exercise following fat adaptation with carbohydrate restoration. *Am J Physiol Endocrinol Metab* **290**, E380–E388.
- Svensson K, Albert V, Cardel B, Salatino S & Handschin C (2016). Skeletal muscle PGC-1 α modulates systemic ketone body homeostasis and ameliorates diabetic hyperketonemia in mice. *FASEB J* **30**, 1976–1986.
- Thomas C, Bishop DJ, Lambert K, Mercier J & Brooks GA (2012). Effects of acute and chronic exercise on sarcolemmal MCT1 and MCT4 contents in human skeletal muscles: current status. *Am J Physiol Regul Integr Comp Physiol* **302**, R1–R14.
- Thomas LK, Ittmann M & Cooper C (1982). The role of leucine in ketogenesis in starved rats. *Biochem J* **204**, 399–403.
- Torrens SL, Areta JL, Parr EB & Hawley JA (2016). Carbohydrate dependence during prolonged simulated cycling time trials. *Eur J Appl Physiol* **116**, 781–790.
- van Loon LJ, Greenhaff PL, Constantin-Teodosiu D, Saris WH & Wagenmakers AJ (2001). The effects of increasing exercise intensity on muscle fuel utilisation in humans. *J Physiol* **536**, 295–304.
- van Loon LJ, Thomason-Hughes M, Constantin-Teodosiu D, Koopman R, Greenhaff PL, Hardie DG, Keizer HA, Saris WH & Wagenmakers AJ (2005). Inhibition of adipose tissue lipolysis increases intramuscular lipid and glycogen use *in vivo* in humans. *Am J Physiol Endocrinol Metab* **289**, E482–E493.

- Veech RL (2004). The therapeutic implications of ketone bodies: the effects of ketone bodies in pathological conditions: ketosis, ketogenic diet, redox states, insulin resistance, and mitochondrial metabolism. *Prostaglandins Leukot Essent Fatty Acids* **70**, 309–319.
- Volek JS, Freidenreich DJ, Saenz C, Kunces LJ, Creighton BC, Bartley JM, Davitt PM, Munoz CX, Anderson JM, Maresh CM, Lee EC, Schuenke MD, Aerni G, Kraemer WJ & Phinney SD (2016). Metabolic characteristics of keto-adapted ultra-endurance runners. *Metabolism* **65**, 100–110.
- Wagenmakers AJ, Beckers EJ, Brouns F, Kuipers H, Soeters PB, van der Vusse GJ & Saris WH (1991). Carbohydrate supplementation, glycogen depletion, and amino acid metabolism during exercise. *Am J Physiol Endocrinol Metab* **260**, E883–E890.
- Wahren J, Sato Y, Ostman J, Hagenfeldt L & Felig P (1984). Turnover and splanchnic metabolism of free fatty acids and ketones in insulin-dependent diabetics at rest and in response to exercise. *J Clin Invest* **73**, 1367–1376.
- Winder WW, Baldwin KM & Holloszy JO (1973). Exercise-induced adaptive increase in rate of oxidation of β -hydroxybutyrate by skeletal muscle. *Proc Soc Exp Biol Med* **143**, 753–755.
- Winder WW, Baldwin KM & Holloszy JO (1974). Enzymes involved in ketone utilization in different types of muscle: adaptation to exercise. *Eur J Biochem* **47**, 461–467.
- Winder WW, Baldwin KM & Holloszy JO (1975). Exercise-induced increase in the capacity of rat skeletal muscle to oxidize ketones. *Can J Physiol Pharmacol* **53**, 86–91.
- Youm YH, Nguyen KY, Grant RW, Goldberg EL, Bodogai M, Kim D, D'Agostino D, Planavsky N, Lupfer C, Kanneganti TD, Kang S, Horvath TL, Fahmy TM, Crawford PA, Biragyn A, Alnemri E & Dixit VD (2015). The ketone metabolite β -hydroxybutyrate blocks NLRP3 inflammasome-mediated inflammatory disease. *Nat Med* **21**, 263–269.
- Zou X, Meng J, Li L, Han W, Li C, Zhong R, Miao X, Cai J, Zhang Y & Zhu D (2016). Acetoacetate accelerates muscle regeneration and ameliorates muscular dystrophy in mice. *J Biol Chem* **291**, 2181–2195.

Additional information

Competing interests

The authors declare no conflict of interest.

Author contributions

B.E. conceived the review and drafted the outline. M.E., K.E.C. and B.E. drafted the initial manuscript, revised and finalised the content. All authors have approved the final version of the manuscript and agree to be accountable for all aspects of the work. All persons designated as authors qualify for authorship, and all those who qualify for authorship are listed.

Funding

M.E. is supported by funding through a UCD Institute for Sport and Health Student Bursary. K.E.C. is supported by funding from Food for Health Ireland (FHI).

Appendix B: Published European Journal of Sport Science article



European Journal of Sport Science



ISSN: 1746-1391 (Print) 1536-7290 (Online) Journal homepage: <http://www.tandfonline.com/loi/tejs20>

Effect of acute ingestion of β -hydroxybutyrate salts on the response to graded exercise in trained cyclists

Mark Evans, Ella Patchett, Rickard Nally, Rachel Kearns, Matthew Larney & Brendan Egan

To cite this article: Mark Evans, Ella Patchett, Rickard Nally, Rachel Kearns, Matthew Larney & Brendan Egan (2018): Effect of acute ingestion of β -hydroxybutyrate salts on the response to graded exercise in trained cyclists, European Journal of Sport Science, DOI: [10.1080/17461391.2017.1421711](https://doi.org/10.1080/17461391.2017.1421711)

To link to this article: <https://doi.org/10.1080/17461391.2017.1421711>



Published online: 16 Jan 2018.



Submit your article to this journal [↗](#)



View related articles [↗](#)



View Crossmark data [↗](#)

Full Terms & Conditions of access and use can be found at
<http://www.tandfonline.com/action/journalInformation?journalCode=tejs20>

during periods of low glucose availability such as during fasting, starvation, and ketogenic diets (Balasse & Féry, 1989; Laffel, 1999; Robinson & Williamson, 1980). Although principally acting as an alternative fuel source for the brain when glucose concentrations are diminished, ketone bodies are also used by skeletal muscle to provide up to 10% of energy during exercise in the fasted state (Balasse, Féry, & Neef, 1978; Féry & Balasse, 1983; Féry, Franken, Neef, & Balasse, 1974; Wahren, Sato, Ostman, Hagenfeldt, & Felig, 1984). However, the direct contribution to energy provision may be secondary to the potential metabolic action of supplemental ketones. For instance, ketone bodies have wide-ranging metabolic effects on peripheral tissues such as glucose sparing, anti-lipolytic effects, and stimulation of muscle protein synthesis (Maizels, Ruderman, Goodman, & Lau, 1977; Mikkelsen, Seifert, Secher, Grøndal, & van Hall, 2015; Nair, Welle, Halliday, & Campbell, 1988). During moderate intensity exercise, infusion of sodium AcAc after an overnight fast attenuates the rise in plasma lactate (Féry & Balasse, 1988), whereas sodium β HB infusion similarly alters the metabolic response to very intense exercise in rats (Kamysheva & Ostrovskaia, 1980) and ischemic forearm exercise in humans (Lestan, Walden, Schmaltz, Sychala, & Fox, 1994).

Despite these observations, the potential performance benefits of ketone bodies have been unexplored until the recent emergence of exogenous ketone supplements in the form of ketone esters and ketone salts. For instance, acute ingestion of the (*R*)-3-hydroxybutyl (*R*)-3-hydroxybutyrate ketone monoester produced plasma β HB concentrations of ~ 3 mM during exercise and improved 30 min time-trial performance by 2% in elite cyclists (Cox et al., 2016). Ketone esters are not commercially available to date, but ketone salts represent a cheaper, more readily-available exogenous ketone supplement. These salts comprise of the

free acid form of β HB buffered with sodium, potassium, and/or calcium salts but are less effective at elevating plasma β HB concentrations compared to the ketone monoester (Stubbs et al., 2017). The effect of acute ketone salt ingestion on short-duration, high-intensity exercise performance in humans has been the subject of two recent reports (O'Malley, Myette-Cote, Durrer, & Little, 2017; Rodger, Plews, Laursen, & Driller, 2017), both of which did not observe the performance benefits associated with the ketone monoester. Given that this is an emerging field of research and to better understand the impact of ketone salt ingestion on responses across a range of exercise intensities, the present study investigated the effect of acute ingestion of a commercially available β HB salt formulation on the metabolic and physiological responses to a graded submaximal exercise session in young, trained male and female cyclists.

Methods

Participants

Nineteen trained cyclists (12 male, 7 female; Table I) gave written informed consent to participate after written and verbal explanation of the procedures. Ethical approval (permit number: LS-15-82-Evans-Egan) was obtained from the University College Dublin Research Ethics Committee. All participants were active in regular cycling training (≥ 6 sessions per week) and competition in road, time-trial, and/or triathlon disciplines and had been competing in their respective discipline for at least one calendar year.

Experimental design

Participants visited the laboratory for exercise tests on three separate occasions. All tests were performed on the same electronically braked stationary cycle

Table I. Participant anthropometrics and fitness profile.

	Whole cohort (<i>n</i> = 19)	Males (<i>n</i> = 12)	Females (<i>n</i> = 7)	Males vs. females, <i>p</i> value
Age (y)	26.8 \pm 7.6	25.6 \pm 6.4	30.6 \pm 8.6	.148
Height (m)	174.3 \pm 8.9	178.7 \pm 7.4	166.7 \pm 5.7	.002
Body mass (kg)	69.0 \pm 9.7	73.8 \pm 6.8	60.9 \pm 8.5	<.001
Body fat (%)	17.6 \pm 6.8	13.7 \pm 3.9	24.3 \pm 5.0	<.001
FFM (kg)	57.7 \pm 10.5	64.3 \pm 5.9	46.4 \pm 5.3	<.001
W _{max} (W)	325 \pm 67	368 \pm 40	251 \pm 23	<.001
LT (W)	245 \pm 59	278 \pm 41	187 \pm 36	<.001
VO _{2peak} (L min ⁻¹)	4.27 \pm 0.85	4.83 \pm 0.43	3.33 \pm 0.43	<.001
VO _{2peak} (mL kg ⁻¹ min ⁻¹)	61.6 \pm 7.1	65.5 \pm 5.6	54.9 \pm 3.6	<.001
VO _{2peak} (mL kg FFM ⁻¹ min ⁻¹)	74.0 \pm 5.1	75.2 \pm 5.1	71.8 \pm 4.7	.171

Note: Data are presented as mean \pm SD. LT, power output at 4 mM lactate threshold; FFM, fat-free mass.

ergometer (Lode Excalibur Sport, Netherlands). Saddle height and handlebar position were adjusted to each participant's preference, but kept consistent for the three visits. Participants performed the exercise tests in their own cycling shoes with appropriate pedals provided by the laboratory. Body mass and height were measured using digital scales (SECA, Germany) and a wall-mounted stadiometer (Holtain, UK), respectively. Body composition was measured using dual-energy X-ray absorptiometry (Lunar iDXA, GE Healthcare, UK).

During their first visit to the lab, participants performed a submaximal incremental exercise test to establish their lactate threshold, after which they performed an incremental exercise test to volitional exhaustion to establish their peak oxygen uptake ($\text{VO}_{2\text{peak}}$). Two experimental trials, each comprised of a graded exercise test of six stages (at power outputs corresponding to approximately 30%, 40%, 50%, 60%, 70%, and 80% $\text{VO}_{2\text{peak}}$), with each stage lasting 8 min (Figure 1), were performed during subsequent visits in a randomized cross-over design. Each experimental trial was identical with the exception of a drink consumed in the hour prior to each exercise test, namely plain water (CON), or β HB salts (KET).

Incremental exercise tests

Assessment of lactate threshold and $\text{VO}_{2\text{peak}}$ was performed in accordance with guidelines from the British Association of Sport and Exercise Sciences (BASES) (Davison & Wooles, 2007; Spurway & Jones, 2007). Briefly, for the determination of lactate threshold, participants completed 4 min stages (3 min of cycling and 1 min of rest), starting at 50 W. The power output was increased by 50 W for the next two stages, and 30 W thereafter until a blood lactate concentration (Lactate Pro 2, Japan) of 4 mM was exceeded. After a 15 min rest, $\text{VO}_{2\text{peak}}$ was determined via an incremental test to exhaustion. Participants began cycling at a pre-determined power output based on body mass as per the BASES guidelines, and power output was progressively increased by 20 W min^{-1} for males and 15 W min^{-1} for females thereafter until volitional exhaustion.

Pre-trial preparation

All experimental trials were performed between 07:00 and 10:00, but on an individual basis, participants performed their second trial at the same time ± 1 h as their first trial. Pre-trial preparation was the same for each experimental trial. Participants were asked to abstain from alcohol for 48 h and caffeine

for at least 12 h and refrain from strenuous exercise training for the day prior to each trial. Each trial took place after a standardized 10 h overnight fast. Participants were asked to keep a one-day portion estimate food diary for the day corresponding to two days prior to the first trial. They were instructed to repeat this pattern of intake before their second trial. On the day immediately prior to both experimental trials, participants were provided with a standardized diet (Gourmet Fuel, Ireland), which provided 40 kcal kg body mass^{-1} at a macronutrient ratio of 40% carbohydrate, 30% protein, and 30% fat. Male participants performed the two experimental trials separated by 7 or 14 days. Because the phase of the menstrual cycle influences fuel utilization during exercise (Oosthuysen & Bosch, 2010), female participants performed the two experimental trials separated by 7 days, but within the early to mid-luteal phase of their menstrual cycle.

Experimental trials

Experimental trials were performed in a randomized cross-over open-label design and were identical with the exception of the drink consumed in the hour prior to each exercise test. The open-label design was chosen because of the difficulty in masking the pungent taste of the β HB salts and considered acceptable because there was no performance element to the experimental design. Therefore, neither the study participants, nor research personnel were blinded to allocation of condition, with the exception of the laboratory technician who did undertake analysis of the blood samples in a blinded manner.

During each trial, a bolus of a given drink was ingested at both 60 min prior to and 15 min prior to the commencement of exercise (Figure 1). Each bolus consisted of either (i) plain water provided at 3.8 mL kg body mass^{-1} (CON), or (ii) β HB salts (KetoCaNa, Prototype Nutrition, IL USA) provided at 0.38 g kg body mass^{-1} dissolved in 3.8 mL kg body mass^{-1} plain water (KET). Each bolus serving of KET provided ~ 18.5 g β HB, 2.1 g sodium and 1.8 g calcium, which is approximately 60% more β HB than the manufacturer's guidelines of 11.7 g β HB per serving. This timing and dosing strategy was based on our own pilot experiments (unpublished data) showing that plasma β HB concentration peaked at 60 min after ingestion of a single bolus, and that a greater elevation in plasma β HB concentration could be achieved with two smaller doses of β HB salts compared with a single larger dose equivalent to the same total amount of β HB salts.

Upon arrival at the laboratory, an indwelling catheter was introduced into an antecubital vein for

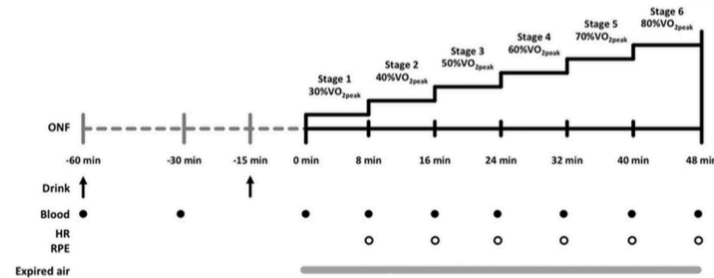


Figure 1. Experimental design schematic. After an overnight fast (ONF), test drinks [water (CON) and β HB salts (KET)] were consumed in two boluses at 60 and 15 min prior to exercise. The graded exercise test consisted of six stages of 8 min in duration performed on a stationary electronically-braked cycle ergometer. HR, RPE, and venous blood were sampled in the last 30 s of each stage. Expired air was collected continuously throughout.

serial blood sampling at rest (-60 , -30 , and 0 min) and during exercise (last 30 s of each 8 min stage) (Figure 1). The catheter was maintained patent between samples with saline (0.9% sodium chloride). The exercise test was graded and consisted of six stages at power outputs corresponding to approximately 30%, 40%, 50%, 60%, 70%, and 80% $\text{VO}_{2\text{peak}}$ with each stage lasting for 8 min (Figure 1). Expired air was collected continuously throughout each exercise test on a breath-by-breath basis (COSMED Quark b2, Italy). During the last 30 s of each 8 min stage, heart rate (HR; Polar, Finland) and rating of perceived exertion (RPE; Borg scale) were recorded, and a blood sample was collected for measurement of plasma β HB, lactate, and glucose concentrations.

Blood analysis

Blood samples (4 mL) were collected in plastic tubes containing sodium fluoride/potassium oxalate (Vacuette Glucose tubes, Greiner-Bio-One, Germany) for subsequent analysis. Samples were stored on ice before centrifugation at $3000g$ for 10 min at 4°C , after which three aliquots of plasma were separated for storage at -80°C until later analysis of plasma β HB, lactate, and glucose (RX Daytona, Randox Laboratories, UK; assay codes RB1007, LC2389, and GL364, respectively).

Data analysis

Cardiopulmonary and metabolic parameters. Minute ventilation (\dot{V}_E), VO_2 , carbon dioxide production (VCO_2), and RER were calculated from an average of breath-by-breath measurements during the last 30 s of each stage in the incremental exercise tests and during the last 2 min of each stage in main

experimental trials. Oxygen pulse (O_2 pulse), defined as oxygen uptake per heartbeat and expressed in mL beat^{-1} , was calculated by dividing VO_2 (L min^{-1}) by HR (beats min^{-1}) during the last 30 s of each stage.

Substrate utilization. The rate of energy expenditure (kcal min^{-1}) during each stage was calculated from average VO_2 and VCO_2 values during the last 30 s of each stage using equations applied on an intensity-dependent basis (Jeukendrup & Wallis, 2005). Rates of carbohydrate and fat oxidation are not reported because of the likely error introduced into these calculations by the oxidation of β HB and AcAc, which yield respiratory quotient values of 0.89 and 1.00, respectively (Frayn, 1983). Therefore, reporting oxidation rates based on RER is inaccurate during periods of nutritional ketosis unless appropriate correction factors for CO_2 displacement and excretion of ketone bodies in urine and expired air are employed (Frayn, 1983), which were beyond the scope of the current work.

Mechanical efficiency. Gross efficiency (GE) was calculated as the ratio of the work performed per minute (\dot{W} converted to kJ min^{-1}) to the energy expended per minute (kJ min^{-1}) at each stage, expressed as a percentage. Delta efficiency (DE) was calculated as the ratio of the change in work performed per minute to the change in energy expended per minute between each stage, expressed as a percentage (Gaesser & Brooks, 1975).

Statistical analysis

Data were evaluated using GraphPad Prism 6 (GraphPad Software, Inc., CA, USA), and are presented as mean \pm SD, with the exception of Figure 2 where

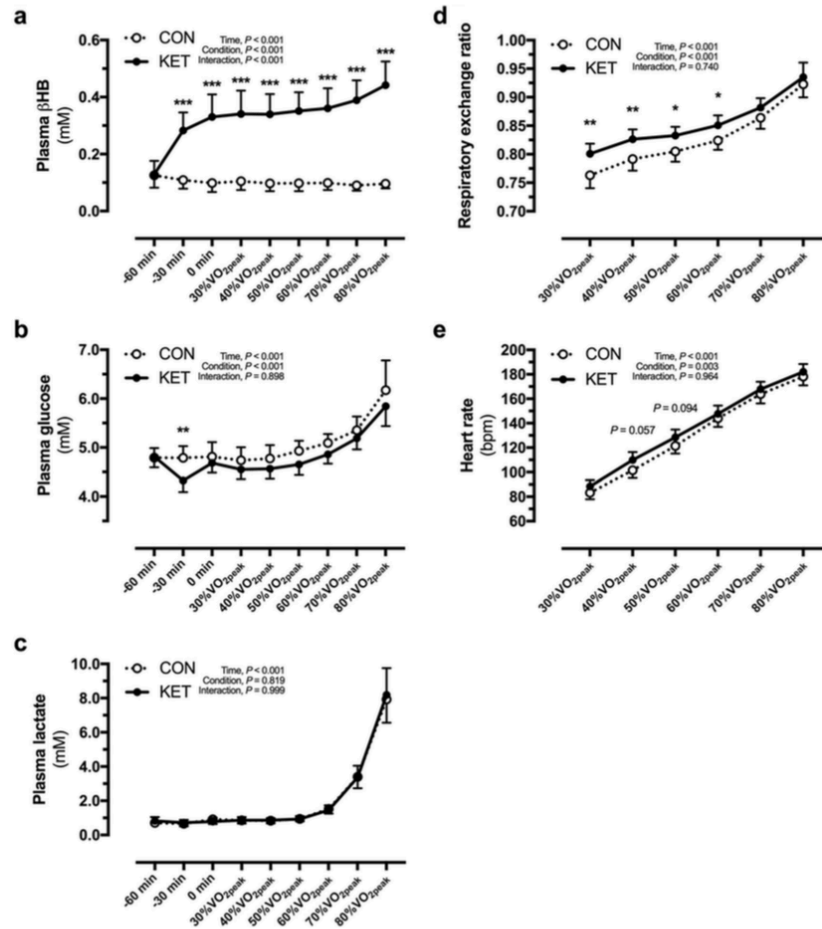


Figure 2. Plasma β HB (a), glucose (b) and lactate (c) concentrations, respiratory exchange ratio (d), and HR (e) during graded exercise on a cycle ergometer in a fasted state (CON) or after ingestion of β HB salts (KET). Data are presented as mean values, with error bars representing 95% confidence intervals. * $p < .05$ KET vs. CON; ** $p < .01$ KET vs. CON; *** $p < .001$ KET vs. CON.

error bars represent 95% confidence intervals. The experiment was powered based on change in RER as the primary outcome, which was chosen as a measure of an altered metabolic response. Based on the aforementioned pilot data where a 0.034 ± 0.015 difference in RER between KET and CON was observed, $n = 13$ participants would have been required given $\alpha = 0.05$ and $1 - \beta = 0.8$ (GPower v3.1). Independent samples t -tests were used to determine differences between male and female participants for baseline characteristics. Two-way (condition \times intensity) repeated measures analysis of

variance was used to determine differences between the two experimental trials for variables with serial measurements. When a main effect of condition, or an interaction effect between condition and intensity, was indicated, post-hoc testing was performed using Holm-Sidak's multiple comparisons test with multiplicity-adjusted p values to compare KET to CON at respective time points. The data were tested for normality and sphericity prior to proceeding with the tests described. In addition, standardized differences in the mean were used to assess magnitudes of effects between conditions at respective time points.

These were calculated using Cohen's *d* effect size (ES) and interpreted using thresholds of <0.25, >0.25, >0.5, and >1.0 for trivial, small, moderate, and large, respectively (Rhea, 2004). Pearson's product-moment correlation coefficient (*r*) was used to explore correlations between variables. No differences were observed between male and female participants for the effect of KET on the metabolic response to exercise compared to water, so male and female data are presented as combined (*n* = 19) data unless otherwise stated. The significance level was set at $\alpha = 0.05$ for all statistical tests.

Results

Plasma β HB, glucose and lactate

Fasting plasma β HB (KET, 0.13 ± 0.10 mM; CON, 0.12 ± 0.09 mM; ES = .05) and glucose (KET, 4.82 ± 0.46 mM; CON, 4.79 ± 0.40 mM; ES = .06) concentrations did not differ between trials (Figure 2). Ingestion of KET resulted in a rise in plasma β HB concentration to 0.28 ± 0.13 mM ($p < .001$) 30 min after ingestion and remained elevated throughout exercise ($p < .001$) (Figure 2(a)). The highest plasma β HB concentration during KET was observed in the final stage of exercise at 0.44 ± 0.15 mM ($p < .001$). Plasma glucose concentration averaged 0.44 ± 0.27 mM lower 30 min after ingestion of KET compared to CON ($p = .008$; ES = .96). An inverse correlation ($r = -0.647$, $p = .004$) was observed for the change in plasma β HB and glucose concentrations at this time point. Plasma glucose concentrations remained lower at all stages throughout exercise, with ES indicating small to moderate effects, i.e. 30%VO_{2peak} -0.19 ± 0.36 mM, ES = .39; 40%VO_{2peak} -0.21 ± 0.43 mM, ES = .44; 50%VO_{2peak} -0.27 ± 0.40 mM, ES = .66; 60%VO_{2peak} -0.21 ± 0.39 mM, ES = .62; 70%VO_{2peak} -0.17 ± 0.54 mM, ES = .33; and 80%VO_{2peak} -0.39 ± 1.24 mM, ES = .34 (Figure 2(b)). Plasma lactate concentrations were elevated above resting values during the final two stages of exercise, but no differences between KET and CON were observed for plasma lactate concentrations at any time point (Figure 2(c)).

Cardiorespiratory responses to graded exercise and KET ingestion

All cardiorespiratory parameters exhibited main effects for exercise intensity (all $p < .001$). No differences in %VO_{2peak}, VO₂, VCO₂, or V_E were observed between conditions (Table II). RER was elevated by KET ($p < .001$ for condition), and was ~ 0.03 higher for intensities up to 60%VO_{2peak} (all $p < .05$), with

ES indicating moderate effects at these intensities, i.e. 30%VO_{2peak} 0.038 ± 0.030 , $p = 0.003$, ES = .90; 40%VO_{2peak} 0.035 ± 0.036 , $p = 0.007$, ES = .92; 50%VO_{2peak} 0.028 ± 0.031 , $p = 0.025$, ES = .81; and 60%VO_{2peak} 0.027 ± 0.037 , $p = 0.031$, ES = .78 (Figure 2(d)). The effect of KET on RER was small at 70%VO_{2peak} (0.018 ± 0.030 , $p = 0.16$, ES = .50) and 80%VO_{2peak} (0.012 ± 0.045 , $p = 0.37$, ES = .28). HR was also elevated by KET ($P = .003$ for condition), wherein HR averaged ~ 4 to 8 bpm higher during KET and ES indicated small to moderate effects, i.e. 30%VO_{2peak} 5.6 ± 4.5 bpm, ES = .48; 40%VO_{2peak} 8.5 ± 7.1 bpm, ES = .66; 50%VO_{2peak} 7.8 ± 7.1 bpm, ES = .55; 60%VO_{2peak} 3.9 ± 8.6 , ES = .26; 70%VO_{2peak} 4.9 ± 8.2 bpm, ES = .29; and 80%VO_{2peak} 4.4 ± 7.0 bpm, ES = .34 (Figure 2(e)). No differences in oxygen pulse, RPE, GE, or DE were observed between conditions (Table II).

Gastrointestinal responses

Thirteen out of 19 (68%) participants reported symptoms of gastrointestinal distress in response to KET ingestion. These comprised of seven (37%), three (16%), two (11%), and one (5%) of the participants reporting nausea, diarrhoea, vomiting, and light-headedness, respectively. These symptoms manifested in the latter stages of and immediately after the cessation of exercise. No symptoms were reported during CON.

Discussion

The aim of the present study was to investigate the effect, if any, of acute ingestion of β HB salts on metabolic and physiological responses to a graded exercise session in trained cyclists. Ingestion of commercially available β HB salts resulted in elevated plasma β HB concentrations (>0.3 mM) at rest and during exercise. This coincided with elevated RER (moderate effects) and HR (small to moderate effects) during submaximal exercise intensities, and a lowering of plasma glucose concentrations (small to moderate effects), compared with the ingestion of water. However, a range of other parameters including plasma lactate, rate of perceived exertion, GE, and DE were unaffected by the acute ingestion of β HB salts.

Exogenous ketone supplements, such as β HB salts, represent a novel method to increase the concentration of circulating ketone bodies without implementing restrictive dietary practices such as fasting or low carbohydrate, ketogenic diets (Cox & Clarke, 2014; Evans et al., 2017). Despite the increasing

Table II. Cardiorespiratory responses during graded exercise in CON or KET.

	Stage 1 30% VO _{2peak}	Stage 2 40% VO _{2peak}	Stage 3 50% VO _{2peak}	Stage 4 60% VO _{2peak}	Stage 5 70% VO _{2peak}	Stage 6 80% VO _{2peak}	Intensity, <i>p</i> value	Condition, <i>p</i> value	Interaction, <i>p</i> value
%W _{max}									
KET	14 ± 6	26 ± 6	38 ± 5	50 ± 5	62 ± 4	74 ± 5	<.001	>.999	>.999
CON	14 ± 6	26 ± 6	38 ± 5	50 ± 5	62 ± 5	74 ± 5			
%VO _{2peak}									
KET	27 ± 4	38 ± 4	49 ± 4	61 ± 4	73 ± 5	85 ± 5	<.001	>.999	>.999
CON	27 ± 4	37 ± 4	49 ± 3	61 ± 3	73 ± 5	85 ± 4			
V _E (L·min ⁻¹)									
KET	26.6 ± 7.6	36.0 ± 8.9	46.4 ± 10.4	59.2 ± 12.7	79.3 ± 18.8	109.5 ± 30.4	<.001	.676	>.999
CON	25.3 ± 7.5	35.0 ± 9.1	45.9 ± 10.1	58.4 ± 12.5	78.4 ± 18.0	108.6 ± 30.8			
ES	.18	.12	.05	.06	.05	.03			
VO ₂ (L·min ⁻¹)									
KET	1.21 ± 0.35	1.65 ± 0.40	2.12 ± 0.45	2.59 ± 0.54	3.15 ± 0.63	3.63 ± 0.72	<.001	.954	>.999
CON	1.18 ± 0.35	1.63 ± 0.42	2.12 ± 0.45	2.61 ± 0.56	3.17 ± 0.67	3.66 ± 0.78			
ES	.08	.04	.01	.03	.03	.03			
VCO ₂ (L·min ⁻¹)									
KET	0.97 ± 0.30	1.37 ± 0.35	1.77 ± 0.41	2.21 ± 0.49	2.79 ± 0.61	3.41 ± 0.78	<.001	.399	>.999
CON	0.90 ± 0.29	1.29 ± 0.35	1.71 ± 0.38	2.15 ± 0.46	2.74 ± 0.61	3.38 ± 0.74			
ES	.23	.20	.16	.14	.08	.04			
RPE									
KET	6.4 ± 0.5	8.1 ± 1.2	10.1 ± 1.8	12.4 ± 1.9	14.4 ± 2.1	16.5 ± 2.3	<.001	.969	>.999
CON	6.4 ± 1.0	8.4 ± 1.7	10.9 ± 1.9	13.1 ± 1.3	14.9 ± 1.1	17.3 ± 1.5			
ES	.07	.23	.45	.43	.30	.38			
O ₂ pulse (mL·beat ⁻¹)									
KET	13.3 ± 3.5	14.7 ± 3.4	16.3 ± 3.1	17.3 ± 3.4	18.6 ± 3.5	19.4 ± 3.9	<.001	.075	.992
CON	14.0 ± 3.6	16.0 ± 3.8	17.5 ± 3.7	18.2 ± 4.1	19.3 ± 4.5	20.6 ± 4.7			
ES	.20	.36	.36	.23	.17	.26			
GE (%)									
KET	10.4 ± 3.6	15.0 ± 2.3	16.9 ± 1.9	18.2 ± 1.7	18.8 ± 1.6	19.1 ± 1.9	<.001	.789	.992
CON	10.7 ± 3.6	15.2 ± 2.0	16.9 ± 1.7	18.2 ± 1.1	18.8 ± 1.0	19.2 ± 1.0			
ES	.07	.10	.01	.04	.03	.12			
DE (%)									
KET	N/A	27.0 ± 4.7	23.1 ± 3.4	24.7 ± 5.7	21.6 ± 3.0	19.9 ± 6.8	<.001	.737	.830
CON	N/A	28.2 ± 9.6	22.7 ± 3.9	24.2 ± 5.2	20.9 ± 5.9	21.8 ± 7.0			
ES		.16	.12	.09	.16	.28			

Notes: Data are presented as mean ± SD. ES was calculated as Cohen's *d* and interpreted using thresholds of <0.25, >0.25, >0.5, and >1.0 for trivial, small, moderate, and large, respectively. % W_{max}, percentage of maximum power output; %VO_{2peak}, percentage of peak oxygen uptake; DE, delta efficiency; GE, gross efficiency; V_E, minute ventilation; VO₂, rate of oxygen uptake; VCO₂, rate of carbon dioxide production; RPE, rating of perceived exertion.

commercial availability of β HB salts, to date, there is a paucity of data from humans on the metabolic response to ingestion at rest or during exercise. The sodium/potassium β HB mineral salt ingested in the present study resulted in a modest elevation (~ 0.3 to 0.4 mM) in plasma β HB concentrations. These values are similar to those observed after a 24 h fast (Balasse & Féry, 1989; Laffel, 1999) and can be considered to have produced nutritional ketosis (i.e. >0.2 mM (Robinson & Williamson, 1980)). The dosing strategy employed involved a bolus ingested both at 60 min and 15 min prior to exercise, but plasma β HB concentrations peaked during the last stage of exercise at 0.44 ± 0.15 mM. This suggests that the supplement was still being released into circulation approximately one hour after the ingestion of the second bolus, a time course consistent with several recent reports describing β HB salt ingestion at rest (Stubbs et al., 2017) and prior to exercise (O'Malley et al., 2017; Rodger et al., 2017). However, these studies reported somewhat higher blood β HB concentrations (~ 0.6 to 1.0 mM) after ingestion of doses providing of 2×11.7 g of β HB (Rodger et al., 2017), ~ 21 to 27 g of β HB (O'Malley et al., 2017), and ~ 12 g or ~ 25 g of β HB (Stubbs et al., 2017), compared to the $2 \times \sim 18.5$ g in the present study. However, unlike the present study, these studies measured β HB concentrations in whole blood from finger-prick sampling using handheld devices, which are known to overestimate blood β HB concentration ranging from 50% to three-fold relative to lab-based measures performed on serum (Guimont et al., 2015; Leckey, Ross, Quod, Hawley, & Burke, 2017).

The aim of ingestion of exogenous ketone supplements is to achieve acute nutritional ketosis (Cox & Clarke, 2014), and this is readily-achieved by the (*R*)-3-hydroxybutyl (*R*)-3-hydroxybutyrate ketone monoester (Cox et al., 2016). Ingestion of 573 mg kg body mass⁻¹ of that supplement raises plasma β HB concentrations to ~ 3 mM 10 min after ingestion, which rise further to ~ 6 mM within the next 60 min at rest (Cox et al., 2016). Clearly the β HB salts ingested in the present study produce plasma β HB concentrations that are ~ 10 -fold less than this. Despite the modest change in plasma β HB concentrations, the acute ingestion of β HB salts does exert some metabolic action at rest and during exercise. For instance, a $\sim 10\%$ decline in plasma glucose was observed 30 min after the ingestion of β HB salts, with an inverse correlation observed between the respective changes in plasma β HB and glucose concentrations at this time. This is consistent with the acute infusion of ketone bodies producing β HB concentrations of ~ 0.5 to 1 mM resulting in a decline in plasma glucose of

$\sim 10\%$ (Mikkelsen et al., 2015; Sherwin, Hendler, & Felig, 1975), and similar results associated with β HB salt ingestion (Stubbs et al., 2017). Moreover, a slightly lower plasma glucose concentration (~ 0.2 mM; small to moderate effects) was evident throughout exercise in the present study, which confirms other recent reports (Leckey et al., 2017; O'Malley et al., 2017; Rodger et al., 2017).

Other effects observed during exercise in the present study include elevations in RER (moderate effects) and HR (small to moderate effects) during the low-to-moderate intensities of exercise. The elevation in RER may be indicative of oxidation of ketone bodies during exercise based on the stoichiometry of oxidation of AcAc. Before being oxidized as a fuel source in skeletal muscle, β HB is re-oxidized to AcAc through the action of 3-hydroxybutyrate dehydrogenase (BDH). The respiratory quotient for oxidation of AcAc (1.0) is identical to that glucose (Frayn, 1983). Therefore, a contribution of ketone oxidation to energy provision likely explains the elevation in RER during exercise after ingestion of β HB salts in the present study. The elevated RER is consistent with a recent report of prolonged submaximal exercise in trained male cyclists (Rodger et al., 2017), but the opposite of what was reported during graded exercise in recreationally active men (O'Malley et al., 2017). Like the former study, we studied trained cyclists, so whether training status is the only explanation for the divergent findings remains to be confirmed. However, this would be consistent with our previous suggestion that oxidation of ketone bodies during exercise is likely to be greatest in trained participants with a high proportion of type I muscle fibres and/or a high oxidative capacity in skeletal muscle (Evans et al., 2017).

Calculations of arteriovenous differences of radio-labelled ketone bodies across working muscles estimate the contribution of ketone bodies to energy provision of 2–10% during exercise in the fasted state (Balasse et al., 1978; Féry & Balasse, 1983; Wahren et al., 1984). This contribution is unlikely to be $>10\%$ unless plasma β HB concentrations are elevated above 1 mM and exercise is being performed by trained participants (Evans et al., 2017). In well-trained participants consuming exogenous ketones as a ketone monoester, the contribution of ketone bodies to energy provision is greater, i.e. 16–18% of total oxygen consumption (Cox et al., 2016). Therefore, although the elevation in plasma β HB concentration in the present study was modest, it is likely that this did result in an increased contribution of ketone bodies to energy provision during exercise.

Apart from a contribution to energy provision, the principal efficacy of supplemental ketones as a performance aid is likely to be secondary effects on

metabolism and alterations in fuel selection (Evans et al., 2017). For instance, acute infusion of sodium AcAc (Féry & Balasse, 1988) or sodium β HB (Lestan et al., 1994) attenuates the exercise-induced rise in plasma lactate, an effect also observed after ingestion of the aforementioned ketone monoester (Cox et al., 2016). In the latter work, reduced glycolytic flux, glycogen sparing, and increased contribution of intramuscular triglyceride to energy provision were observed during 2 h of cycling at $\sim 70\%$ $\text{VO}_{2\text{max}}$. However, an attenuation of the rise in plasma lactate was not observed in the present study or in other recent studies examining acute ingestion of β HB salts (O'Malley et al., 2017; Rodger et al., 2017). Again, this might be explained by the relatively lower increase in plasma β HB concentration produced by the β HB salts compared to the ketone monoester.

An important methodological note is that the β HB salts used in the present study provide a racemic mixture of β HB, i.e. containing both the D- and L-enantiomers of β HB (also designated R- and S-, respectively), whereas the β HB assay employed determines the concentration of D- β HB. D- β HB is the circulating and primary form of β HB (Tsai et al., 2006), but intracellular concentrations of L- β HB are sensitive to factors such as aging and metabolic health (Hsu et al., 2011). The D- and L- enantiomers of β HB exert divergent physiological effects on glucose metabolism in the heart (Tsai et al., 2006) and skeletal muscle (Yamada, Zhang, Westerblad, & Katz, 2010), and on longevity (Edwards et al., 2014). Recent work has demonstrated that racemic β HB ingested as β HB salts results in elevations in L- β HB concentrations of ~ 2 mM (Stubbs et al., 2017). However, it is doubtful that a change in circulating L- β HB concentration, if provided by an exogenous ketone supplement, would have any direct effect on substrate metabolism in skeletal muscle. For instance, L- β HB is not a substrate for mitochondrial BDH and thus is not metabolized to AcAc (Scofield et al., 1982), and its physiological role is most likely in the synthesis of sterols and fatty acids in non-muscle tissues (Webber & Edmond, 1977).

The small to moderate effects observed for an elevated HR of 4–8 bpm after ingestion of β HB salts compared to water may warrant future investigation. HR during the exercise was not reported in previous work with β HB infusion, or ketone monoester or β HB salt ingestion, but was elevated by 25% under resting conditions after sodium β HB infusion compared to saline infusion (Gormsen et al., 2017). This indicates an effect of β HB itself rather than sodium load, but occurred at a plasma β HB concentration of ~ 4 mM in contrast to ~ 0.4 mM in the present study. Alternatively, the sodium load

delivered by the β HB salts may exert some hemodynamic effects. Acute sodium ingestion can transiently elevate blood pressure (Farquhar, Paul, Prettyman, & Stillabower, 2005), and sodium bicarbonate ingestion providing a similar dose of sodium to the present study results in an elevation in HR of ~ 10 bpm during moderate intensity exercise (Kahle, Kelly, Eliot, & Weiss, 2013).

Also notable in the present study was that 13 out of 19 (68%) participants reported symptoms of gastrointestinal distress after exercise in the β HB salt condition. The hypertonic nature of the β HB salts ingested likely caused an intraluminal osmotic load and water shift into the intestinal lumen resulting in osmotic diarrhoea. However, gastrointestinal distress is also a potential side effect of acute ingestion of ketone esters, with high prevalence noted after the ingestion of the ketone diester by elite cyclists (Leckey et al., 2017), and increasing incidences occurring with increasing dosages of the ketone monoester (Clarke et al., 2012). Clearly, such issues would be deleterious to exercise performance, and, therefore, require further exploration, either in terms of optimal dosing strategies, or whether repeat exposure to exogenous ketone supplements reduces these symptoms.

In conclusion, acute ingestion of a commercially available β HB salt formulation by trained cyclists resulted in a modest increase in plasma β HB concentrations before and during graded exercise to levels that can be considered acute nutritional ketosis. This resulted in alterations in the metabolic and physiological response to exercise as evidenced by lowering of plasma glucose concentrations and elevated RER and HR values at low-to-moderate exercise intensities compared to ingestion of water. However, no effect was observed on perceived exertion or muscular efficiency or on plasma lactate concentrations. This is in contrast to previous work using β HB infusion or ingestion of a ketone monoester supplement, both of which achieve markedly higher plasma β HB concentrations during exercise. This suggests the likelihood that a dose-response effect exists for exogenous ketone supplements on metabolic responses and exercise performance. Given the gastrointestinal issues observed with the present β HB salts, further work is needed with other methods of increasing circulating ketone concentrations including improved free acid or mineral salt formulations, before the merit, if any, of ketone salts for performance enhancement in athletes is likely to be realised.

Disclosure statement

No potential conflict of interest was reported by the authors.

Funding

Support for this research came from a UCD Institute for Sport and Health student bursary to Mr Evans, who is currently funded by an Irish Research Council Enterprise Partnership Postgraduate Fellowship (EPSPG2016177).

ORCID

Brendan Egan  <http://orcid.org/0000-0001-8327-9016>

References

- Balasse, E. O., Fery, F., & Neef, M. A. (1978). Changes induced by exercise in rates of turnover and oxidation of ketone bodies in fasting man. *Journal of Applied Physiology: Respiratory, Environmental and Exercise Physiology*, 44(1), 5–11.
- Balasse, E. O., & Fery, F. (1989). Ketone body production and disposal: Effects of fasting, diabetes, and exercise. *Diabetes/Metabolism Reviews*, 5(3), 247–270.
- Burke, L. M. (2015). Re-Examining high-Fat diets for sports performance: Did we call the 'nail in the coffin' too soon? *Sports Medicine*, 45(Suppl 1), S33–S49. doi:10.1007/s40279-015-0393-9.
- Clarke, K., Tchabanenko, K., Pawlosky, R., Carter, E., Todd King, M., Musa-Veloso, K., ... Veech, R. L. (2012). Kinetics, safety and tolerability of (R)-3-hydroxybutyl (R)-3-hydroxybutyrate in healthy adult subjects. *Regulatory Toxicology and Pharmacology*, 63(3), 401–408. doi:10.1016/j.yrtph.2012.04.008.
- Cox, P. J., & Clarke, K. (2014). Acute nutritional ketosis: Implications for exercise performance and metabolism. *Extreme Physiology and Medicine*, 3(1), 17. doi:10.1186/2046-7648-3-17.
- Cox, P. J., Kirk, T., Ashmore, T., Willerton, K., Evans, R., Smith, A., ... Clarke, K. (2016). Nutritional ketosis alters fuel preference and thereby endurance performance in athletes. *Cell Metabolism*, 24(2), 256–268. doi:10.1016/j.cmet.2016.07.010.
- Davison, R. C. R., & Woole, A. L. (2007). Cycling. In E. M. Winter, A. M. Jones, R. C. R. Davison, P. D. Bromley, & T. H. Mercer (Eds.), *Sport and exercise physiology testing guidelines: Volume I – sport testing* (pp. 160–164). Oxon, UK: Routledge.
- Edwards, C., Canfield, J., Copes, N., Rehan, M., Lipps, D., & Bradshaw, P. C. (2014). D-beta-hydroxybutyrate extends lifespan in *C. elegans*. *Aging (Albany NY)*, 6(8), 621–644. doi:10.18632/aging.100683.
- Egan, B., & Zierath, J. R. (2013). Exercise metabolism and the molecular regulation of skeletal muscle adaptation. *Cell Metabolism*, 17(2), 162–184. doi:10.1016/j.cmet.2012.12.012.
- Evans, M., Cogan, K. E., & Egan, B. (2017). Metabolism of ketone bodies during exercise and training: Physiological basis for exogenous supplementation. *The Journal of Physiology*, 595(9), 2857–2871. doi:10.1113/jp273185.
- Farquhar, W. B., Paul, E. E., Prettyman, A. V., & Stillabower, M. E. (2005). Blood pressure and hemodynamic responses to an acute sodium load in humans. *Journal of Applied Physiology*, 99(4), 1545–1551. doi:10.1152/jappphysiol.00262.2005.
- Fery, F., Franken, P., Neef, M. A., & Balasse, E. O. (1974). Influence of muscular exercise on the rates of uptake and oxidation of infused ketone bodies in normal man. *Archives Internationales de Physiologie et de Biochimie*, 82(2), 381–385.
- Féry, F., & Balasse, E. O. (1983). Ketone body turnover during and after exercise in overnight-fasted and starved humans. *The American Journal of Physiology*, 245(4), E318–E325.
- Féry, F., & Balasse, E. O. (1988). Effect of exercise on the disposal of infused ketone bodies in humans. *Journal of Clinical Endocrinology and Metabolism*, 67(2), 245–250. doi:10.1210/jcem-67-2-245.
- Frayn, K. N. (1983). Calculation of substrate oxidation rates in vivo from gaseous exchange. *Journal of Applied Physiology*, 55(2), 628–634.
- Gaesser, G. A., & Brooks, G. A. (1975). Muscular efficiency during steady-rate exercise: Effects of speed and work rate. *Journal of Applied Physiology*, 38(6), 1132–1139.
- Gormsen, L. C., Svart, M., Thomsen, H. H., Søndergaard, E., Vendelbo, M. H., Christensen, N., ... Møller, N. (2017). Ketone body infusion with 3-hydroxybutyrate reduces myocardial glucose uptake and increases blood flow in humans: A positron emission tomography study. *Journal of the American Heart Association*, 6(3), e005066. doi:10.1161/jaha.116.005066.
- Guimont, M. C., Desjober, H., Fonfrède, M., Vitoux, D., Benoist, J. F., Launay, J. M., ... Lefèvre, G. (2015). Multicentric evaluation of eight glucose and four ketone blood meters. *Clinical Biochemistry*, 48(18), 1310–1316. doi:10.1016/j.clinbiochem.2015.07.032.
- Hsu, W. Y., Kuo, C. Y., Fukushima, T., Imai, K., Chen, C. M., Lin, P. Y., & Lee, J. A. (2011). Enantioselective determination of 3-hydroxybutyrate in the tissues of normal and streptozotocin-induced diabetic rats of different ages. *Journal of Chromatography. B, Analytical Technologies in the Biomedical and Life Sciences*, 879(29), 3331–3336. doi:10.1016/j.jchromb.2011.07.038.
- Jeukendrup, A. E., & Wallis, G. A. (2005). Measurement of substrate oxidation during exercise by means of gas exchange measurements. *International Journal of Sports Medicine*, 26(Suppl 1), S28–S37.
- Kahle, L. E., Kelly, P. V., Eliot, K. A., & Weiss, E. P. (2013). Acute sodium bicarbonate loading has negligible effects on resting and exercise blood pressure but causes gastrointestinal distress. *Nutrition Research*, 33(6), 479–486. doi:10.1016/j.nutres.2013.04.009.
- Kamysheva, V. A., & Ostrovskaya, R. U. (1980). Effect of sodium hydroxybutyrate on the ammonia level in the rat muscles under physical exercise. *Biulleten Eksperimental'noi Biologii i Meditsiny*, 89(1), 25–27.
- Laffel, L. (1999). Ketone bodies: A review of physiology, pathophysiology and application of monitoring to diabetes. *Diabetes/Metabolism Research and Reviews*, 15(6), 412–426.
- Leckey, J. J., Ross, M. L., Quod, M., Hawley, J. A., & Burke, L. M. (2017). Ketone diester ingestion impairs time-trial performance in professional cyclists. *Frontiers in Physiology*, 8, 41. doi:10.3389/fphys.2017.00806.
- Leston, B., Walden, K., Schmaltz, S., Spychala, J., & Fox, I. H. (1994). beta-Hydroxybutyrate decreases adenosine triphosphate degradation products in human subjects. *The Journal of Laboratory and Clinical Medicine*, 124(2), 199–209.
- Maizels, E. Z., Ruderman, N. B., Goodman, M. N., & Lau, D. (1977). Effect of acetoacetate on glucose metabolism in the soleus and extensor digitorum longus muscles of the rat. *Biochemical Journal*, 162(3), 557–568.
- Mikkelsen, K. H., Seifert, T., Secher, N. H., Grøndal, T., & van Hall, G. (2015). Systemic, cerebral and skeletal muscle ketone body and energy metabolism during acute hyper-D-beta-hydroxybutyrate in post-absorptive healthy males. *The Journal of Clinical Endocrinology and Metabolism*, 100(2), 636–643. doi:10.1210/jc.2014-2608.
- Nair, K. S., Welle, S. L., Halliday, D., & Campbell, R. G. (1988). Effect of beta-hydroxybutyrate on whole-body leucine kinetics and fractional mixed skeletal muscle protein synthesis in humans. *The Journal of Clinical Investigation*, 82(1), 198–205. doi:10.1172/jci113570.

- O'Malley, T., Myette-Cote, E., Durrer, C., & Little, J. P. (2017). Nutritional ketone salts increase fat oxidation but impair high-intensity exercise performance in healthy adult males. *Applied Physiology, Nutrition, and Metabolism*, 42, 1031–1035. doi:10.1139/apnm-2016-0641.
- Oosthuysen, T., & Bosch, A. N. (2010). The effect of the menstrual cycle on exercise metabolism: Implications for exercise performance in eumenorrhoeic women. *Sports Medicine*, 40(3), 207–227. doi:10.2165/11317090-000000000-00000.
- Rhea, M. R. (2004). Determining the magnitude of treatment effects in strength training research through the use of the effect size. *Journal of Strength and Conditioning Research*, 18(4), 918–920. doi:10.1519/14403.1.
- Robinson, A. M., & Williamson, D. H. (1980). Physiological roles of ketone bodies as substrates and signals in mammalian tissues. *Physiological Reviews*, 60(1), 143–187.
- Rodger, S., Plews, D., Laursen, P., & Driller, M. W. (2017). Oral β-hydroxybutyrate salt fails to improve 4-minute cycling performance following submaximal exercise. *Journal of Science and Cycling*, 6(1), 26–31.
- Scofield, R. F., Brady, P. S., Schumann, W. C., Kumaran, K., Ohgaku, S., Margolis, J. M., & Landau, B. R. (1982). On the lack of formation of L-(+)-3-hydroxybutyrate by liver. *Archives of Biochemistry and Biophysics*, 214(1), 268–272.
- Sherwin, R. S., Hendler, R. G., & Felig, P. (1975). Effect of ketone infusions on amino acid and nitrogen metabolism in man. *The Journal of Clinical Investigation*, 55(6), 1382–1390. doi:10.1172/jci108057.
- Spurway, N., & Jones, A. M. (2007). Lactate testing. In E. M. Winter, A. M. Jones, R. C. R. Davison, P. D. Bromley, & T. H. Mercer (Eds.), *Sport and exercise physiology testing guidelines: Volume 1 – sport testing* (pp. 112–119). Oxon: Routledge.
- Stubbs, B. J., Cox, P. J., Evans, R. D., Santer, P., Miller, J. J., Faull, O. K., ... Clarke, K. (2017). On the metabolism of exogenous ketones in humans. *Frontiers in Physiology*, 8, 137. doi:10.3389/fphys.2017.00848.
- Tsai, Y. C., Chou, Y. C., Wu, A. B., Hu, C. M., Chen, C. Y., Chen, F. A., & Lee, J. A. (2006). Stereoselective effects of 3-hydroxybutyrate on glucose utilization of rat cardiomyocytes. *Life Sciences*, 78(12), 1385–1391. doi:10.1016/j.lfs.2005.07.013.
- Wahren, J., Sato, Y., Ostman, J., Hagenfeldt, L., & Felig, P. (1984). Turnover and splanchnic metabolism of free fatty acids and ketones in insulin-dependent diabetics at rest and in response to exercise. *The Journal of Clinical Investigation*, 73(5), 1367–1376. doi:10.1172/jci111340.
- Webber, R. J., & Edmond, J. (1977). Utilization of L-(+)-3-hydroxybutyrate, D-(-)-3-hydroxybutyrate, acetoacetate, and glucose for respiration and lipid synthesis in the 18-day-old rat. *The Journal of Biological Chemistry*, 252(15), 5222–5226.
- Yamada, T., Zhang, S. J., Westerblad, H., & Katz, A. (2010). β-hydroxybutyrate inhibits insulin-mediated glucose transport in mouse oxidative muscle. *The American Journal of Physiology: Endocrinology and Metabolism*, 299(3), E364–E373. doi:10.1152/ajpendo.00142.2010.

Intermittent Running and Cognitive Performance after Ketone Ester Ingestion

MARK EVANS¹ and BRENDAN EGAN^{1,2}

¹School of Health and Human Performance, Dublin City University, Dublin, IRELAND; and ²National Institute for Cellular Biotechnology, Dublin City University, Dublin, IRELAND

ABSTRACT

EVANS, M., and B. EGAN. Intermittent Running and Cognitive Performance after Ketone Ester Ingestion. *Med. Sci. Sports Exerc.*, Vol. 50, No. 11, pp. 2330–2338, 2018. **Purpose:** Ingestion of exogenous ketones alters the metabolic response to exercise and may improve exercise performance, but it has not been explored in variable-intensity team sport activity, or for effects on cognitive function. **Methods:** On two occasions in a double-blind, randomized crossover design, 11 male team sport athletes performed the Loughborough Intermittent Shuttle Test (part A, 5 × 15-min intermittent running; part B, shuttle run to exhaustion), with a cognitive test battery before and after. A 6.4% carbohydrate–electrolyte solution was consumed before and during exercise either alone (PLA) or with 750 mg·kg⁻¹ of a ketone ester (KE) supplement. Heart rate, RPE, and 15-m sprint times were recorded throughout, and serial venous blood samples were assayed for plasma glucose, lactate, and β-hydroxybutyrate. **Results:** KE resulted in plasma β-hydroxybutyrate concentrations of ~1.5 to 2.6 mM during exercise ($P < 0.001$). Plasma glucose and lactate concentrations were lower during KE compared with PLA (moderate-to-large effect sizes). Heart rate, RPE, and 15-m sprint times did not differ between trials. Run time to exhaustion was not different ($P = 0.126$, $d = 0.45$) between PLA (mean = 268 s, 95% confidence interval [CI] = 199–336 s) and KE (mean = 229 s, 95% CI = 178–280 s). Incorrect responses in a multitasking test increased from pre- to postexercise in PLA (mean = 1.8, 95% CI = -0.6 to 4.1) but not in KE (mean = 0.0, 95% CI = -1.8 to 1.8) ($P = 0.017$, $d = 0.70$). **Conclusion:** Compared with carbohydrate alone, coingestion of a KE by team sport athletes attenuated the rise in plasma lactate concentrations but did not improve shuttle run time to exhaustion or 15-m sprint times during intermittent running. An attenuation of the decline in executive function after exhausting exercise suggests a cognitive benefit after KE ingestion. **Key Words:** β-HYDROXYBUTYRATE, LACTATE, LOUGHBOROUGH INTERMITTENT SHUTTLE TEST, RUNNING, TEAM SPORT

Ketone bodies, namely, β-hydroxybutyrate (βHB), acetoacetate, and acetone, are fatty acid metabolites whose production markedly increases in physiological states characterized by reduced glucose availability, such as starvation and ketogenic diets (1,2). Ketone bodies are principally produced as a survival mechanism to provide a substrate for the brain, but they are also oxidized by skeletal muscle and provide up to 10% of energy during exercise in a fasted state (3). Infusion of ketone bodies exerts a range of metabolic actions, such as attenuation of hepatic glucose output, antilipolytic effects in adipose tissue and glucose “sparing,” and stimulation of protein synthesis in skeletal muscle (4–6).

The effects of ketone bodies on substrate utilization during exercise and, consequently, athletic performance are of increasing interest because of the development of exogenous ketone supplements, namely, ketone salts and ketone esters (7–9). These formulations represent a method of acutely inducing nutritional ketosis (plasma βHB >0.5 mM) resulting in a variety of effects on exercise metabolism, performance, and recovery (10–17). Ketone salts in their presently available racemic form produce only modest changes (<1.0 mM) in plasma βHB concentrations (9,12,14,16,17). Although their preexercise ingestion can alter the metabolic response to exercise (12,16), there is no evidence of an ergogenic effect (12,14). Alternatively, exogenous ketone supplements in the form of ketone esters produce markedly greater changes in plasma βHB concentrations than ketone salts in humans (9) and rats (8).

Two ketone esters have been reported in the recent literature: a R,S-1,3-butanediol acetoacetate ketone diester (KDE) (8,15) and a (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester (KME) (7,9,10). Both esters were tested in elite endurance athletes with divergent findings (10,15). Acute ingestion of KME produced plasma βHB concentrations of ~3.0 mM after 20 min and improved 30 min time-trial performance by 2% (10). By contrast, acute ingestion of KDE was less effective at raising serum βHB concentrations (~0.4 mM) and impaired 31.2-km time-trial performance by 2% (15). Consumption of

Address for correspondence: Brendan Egan, Ph.D., School of Health and Human Performance, Dublin City University, Glasnevin, Dublin 9, Ireland; E-mail: brendan.egan@dcu.ie.
Submitted for publication April 2018.
Accepted for publication June 2018.

0195-9131/18/5011-2330/0
MEDICINE & SCIENCE IN SPORTS & EXERCISE®
Copyright © 2018 by the American College of Sports Medicine
DOI: 10.1249/MSS.0000000000001700

KME increased the contribution of ketone bodies to fuel provision during exercise to 16%–18% of total energy provision, in addition to marked metabolic effects, including the attenuation of blood lactate concentrations, “sparing” of muscle glycogen, and increased intramuscular triglyceride utilization (10). The reduction in glycolytic flux may reflect an impairment of carbohydrate utilization rather than glycogen sparing (18–20), a key question that requires further investigation. The former is likely to impair performance in high-intensity sports that demand a high rate of ATP provision from carbohydrate sources (18).

Team sports such as Australian football, soccer, Gaelic games, rugby union, lacrosse, and field hockey are high-intensity and intermittent in nature, consisting of repeated periods of high-intensity activity (sprinting) interspersed with exercise at low-to-moderate intensities (walking and jogging) (21,22). Nutrition guidelines for soccer, for instance, recommend high intakes of carbohydrate before and during competition to maximize muscle glycogen stores with the view to enhancing performance (23). Soccer match play results in a marked reduction in muscle glycogen, and high-intensity running is attenuated in the last 15 min of play (24,25). Therefore, nutrition strategies that could spare glycogen and maintain high-intensity running in the latter parts of matches are of interest to scientists and practitioners, but research on exogenous ketone supplements to date has mostly focused on athletes from endurance sports (10,14–16). Moreover, because ketone bodies are the dominant fuel source for the brain in ketogenic states (26), there is potential for central and/or cognitive effects of exogenous ketone supplements, but to date cognitive effects were only explored in short-term feeding trials in rats (27–29). Therefore, the aim of the present study was to investigate the effects of acute ingestion of a ketone ester on metabolic responses, physical performance, and cognitive performance in team sport athletes in response to an intermittent running protocol that simulated soccer match play.

METHODS

Participants. Eleven male team sport athletes (mean \pm SD: age = 25.4 ± 4.6 yr, height = 1.80 ± 0.05 m, body mass = 78.6 ± 5.3 kg, $\dot{V}O_{2\max} = 53.9 \pm 2.2$ mL·kg⁻¹·min⁻¹) gave written informed consent to participate after written and verbal explanation of the procedures. Ethical approval (permit number: DCUREC2017_130) was obtained from the Dublin City University Research Ethics Committee in accordance with the Declaration of Helsinki. All participants were actively training and competing in high-intensity field-based team sports.

Experimental design. Participants visited the laboratory for exercise tests on three separate occasions over a 14- to 21-d period. During their first visit to the laboratory, each participant's maximal oxygen consumption ($\dot{V}O_{2\max}$) and speed at $\dot{V}O_{2\max}$ were determined using a progressive multi-stage shuttle run test (Yo-Yo intermittent recovery test level 1;

Yo-Yo IR1) (30). These data were used to determine jogging (55% $\dot{V}O_{2\max}$) and cruising (95% $\dot{V}O_{2\max}$) speeds for use during the Loughborough Intermittent Shuttle Test (LIST). The LIST is a validated simulation of the physiological and metabolic responses during soccer match play and consists of two parts: part A comprises a fixed period of variable-intensity shuttle running over 20 m; part B consists of continuous running, alternating every 20 m between 55% and 95% $\dot{V}O_{2\max}$ until volitional fatigue (31). After a 15-min rest after completion of the Yo-Yo IR1, participants were familiarized with the LIST protocol by completing one 15-min block at their personalized running speeds. Cognitive tests were performed before the Yo-Yo IR1 and after familiarization with the LIST to familiarize participants with the cognitive test battery.

Two main experimental trials, each comprising of the LIST (parts A and B) with cognitive tests before and after, were performed during subsequent visits in a double-blinded, randomized crossover design. Both experimental trials included a standardized diet for ~36 h before the exercise test and were identical except for the drinks consumed before and during the LIST, namely, a 6.4% carbohydrate-electrolyte solution, which was either flavored (Symrise, UK) and acted as the control/placebo condition (PLA), or included a ketone ester (KE) (Fig. 1). The primary outcome was endurance capacity measured by run time to exhaustion in the LIST part B, with secondary outcomes including 15-m sprint times during the LIST part A, heart rate (HR), RPE, and plasma glucose, lactate, and β HB concentrations.

Incremental exercise test and familiarization. For determination of $\dot{V}O_{2\max}$, jogging (55% $\dot{V}O_{2\max}$), and cruising (95% $\dot{V}O_{2\max}$) during the LIST, participants completed the Yo-Yo IR1. All participants completed a standardized 5-min warm-up consisting of progressive shuttle runs at 20%, 40%, 60%, and 80% and dynamic stretching (high knees, heel kicks, and groin bridges), followed by a period of self-selected stretching. The Yo-Yo IR1 consists of 40-m shuttle runs (2×20 m) between two sets of cones set 20 m apart. Shuttles progressively increase in speed that is dictated by an audio signal (Teambeep Software, UK). Each 40-m shuttle is separated by a 10-s rest period. The test was terminated when participants failed to complete the second 20-m shuttle on two consecutive occasions or if they reached volitional fatigue. $\dot{V}O_{2\max}$ was calculated as follows:

$$\dot{V}O_{2\max} \text{ (mL}\cdot\text{kg}^{-1}\cdot\text{min}^{-1}\text{)} = \text{Yo-Yo IR1 distance (m)} \times 0.0084 + 36.4.$$

After a 15-min break, participants were familiarized with the LIST by performing one block of intermittent activity, i.e., 15 min of part A, were allowed 3 min of rest, and then completed the part B run to exhaustion. Participants completed a battery of cognitive tests before the Yo-Yo IR1 and after the intermittent run to exhaustion.

Cognitive test battery. The battery of cognitive tests (CANTAB Cognition, UK) was administered via a touch screen tablet lasting ~25 min. An identical test battery was administered before and after each trial. Technical issues,

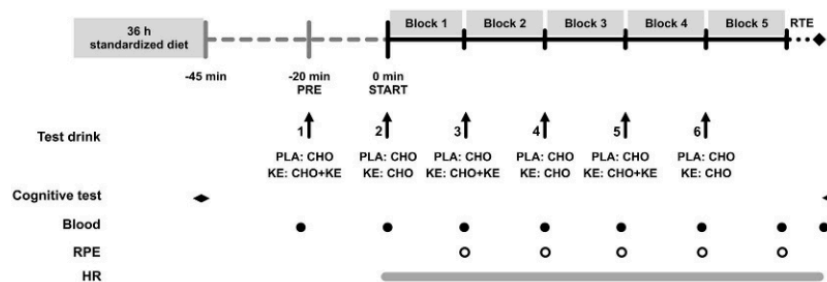


FIGURE 1—Schematic of the study protocol. CHO, carbohydrate–electrolyte solution; PLA, placebo; RTE, shuttle run to exhaustion.

namely, with loss of wireless Internet access during test administration, resulted in the data for the cognitive test battery comprising of $n = 8$ participants.

During the reaction time task, participants select and hold a button at the bottom of the screen, and five circles are presented above. In each case, a yellow dot appears in one of the five circles, and the participant must react as soon as possible, releasing the button at the bottom of the screen and selecting the circle in which the dot appeared. Release time (ms), reaction time (ms), and number of errors were recorded.

The multitasking test is a test of executive function that measures the participant's ability to switch attention between stimuli and ignore task-irrelevant information. White arrows are displayed on a black background, with the arrows located on either the left or right side of the screen, and pointing either to the left or to the right. A cue is displayed at the same time as the arrows, reading either "SIDE" or "DIRECTION." When the "SIDE" cue is presented, the participant is required to press a button on the left or right of the screen corresponding to the side of the screen where the arrow is presented, regardless of the direction the arrow is pointing. Conversely, when the "DIRECTION" cue is presented, the participant is required to touch a button on the left or right of the screen corresponding to the direction the arrow is pointing, regardless of which side of the screen the arrow is presented. Reaction time (ms) and number of correct and incorrect responses were recorded.

The rapid visual information processing task is a test of sustained attention. The participant is presented with a white box in the center of the screen. Single digits ranging from 2 to 9 are presented one at a time in a pseudorandom order inside the box, appearing at a rate of 100 digits per minute. The participant is required to detect specific three-digit sequences, including 2-4-6, 4-6-8, and 3-5-7. As soon as a target sequence is detected, the participant is required to touch a button on the screen. Response latency (ms), correct responses, and false alarms were recorded.

Pretrial preparation. All experimental trials were performed between 3:30 PM and 8:00 PM, but on an individual basis, participants performed their second trial at the same time ± 1 h as their first trial. Pretrial preparation was the same for each experimental trial. Participants were asked to

abstain from alcohol for 48 h and caffeine for 24 h and refrain from strenuous exercise training the day before each trial. The day before experimental trials, participants were provided with a standardized diet (Gourmet Fuel, Ireland), which provided 40 kcal·kg body mass⁻¹ at a macronutrient ratio of 60% carbohydrate, 20% protein, and 20% fat. On the day of experimental trials, participants consumed two meals providing 3 g·kg body mass⁻¹ of carbohydrate before arriving at the laboratory. The second meal was consumed 3 h before the initiation of the LIST. In addition to the energy content and macronutrient ratio, the food itself was identical for both trials. Participants performed the two experimental trials separated by either 7 or 14 d.

Experimental trials. Experimental trials were performed in a double-blinded, randomized crossover design and were identical except for the drinks consumed. During each trial, a bolus of a given drink was ingested 20 min before exercise (drink 1), and during each 3-min seated break during part A (drinks 2 to 6) (Fig. 1). During PLA, a 6.4% carbohydrate–electrolyte solution (Lucozade Sport, Lucozade Ribena Suntory Ltd., UK) was provided at a rate of ~ 1.2 g·min⁻¹ of exercise. During KE, a 6.4% carbohydrate–electrolyte solution was provided at a rate of ~ 1.2 g·min⁻¹ combined with 750 mg·kg body mass⁻¹ of a R- β HB (R)1,3-butanediol ketone ester (KE4, KetoneAid Inc., Falls Church, VA). The ketone ester was mixed directly with the carbohydrate–electrolyte solution for ingestion in three boluses (50:25:25), i.e., at 20 min before exercise (drink 1), and after 30 (drink 3) and 60 min (drink 5) of exercise, respectively (Fig. 1). During PLA, drinks 1, 3, and 5 were flavored with a bitter additive (Symrise, UK) to taste-match with KE, and in both trials, drinks 2, 4, and 6 were provided as the unadulterated carbohydrate–electrolyte solution. All drinks were administered in opaque drinks bottles.

Upon arrival at the laboratory, participants provided a urine sample for assessment of hydration status (PalOSMO, VITECH Scientific, Japan) and then proceeded to complete the described battery of cognitive tests. Thereafter, an indwelling catheter (21G Insyte Autoguard; Becton Dickinson, Franklin Lakes, NJ) was introduced into an antecubital vein for serial venous blood sampling at rest (-20 and 0 min), during each 3-min seated rest period between the 15-min blocks in part A and immediately after the run to exhaustion.

Participants were fitted with a Bluetooth HR monitor (Polar V7; Polar Electro Oy, Kempele, Finland) for continuous recording of HR and then performed the standardized 10-min warm up followed by self-selected stretching. Participants then performed the LIST protocol (part A: 5 × 15 min intermittent activity; part B: run to exhaustion) (31). All exercise intensities were based on percentages of $\dot{V}O_{2\max}$ determined during the Yo-Yo IR1. The repeating order of activity in part A, which occurs in a continuous manner for each 15 min block, comprises of 3 × 20 m at walking speed, 1 × maximal 15 m sprint, 4-s recovery, 3 × 20 m jogging speed (55% $\dot{V}O_{2\max}$), and 3 × 20 m at cruising speed (95% $\dot{V}O_{2\max}$). Sprint times were measured by two sets of wireless infrared photoelectric cells (TC Timing System; Brower Timing, Draper, UT).

Part B consists of single 20-m shuttles alternating between jogging (55% $\dot{V}O_{2\max}$) and cruising (95% $\dot{V}O_{2\max}$) speeds. The shuttle run to exhaustion, measured in seconds, continued until participants were unable to complete two consecutive shuttles at cruising speeds, or until volitional fatigue. All speeds were dictated using audio software (Team Beep Software, UK). All participants received consistent encouragement during the maximal sprinting of part A and the run to exhaustion of part B.

Venous blood samples were collected during the 3-min break between each 15-min block of part A, and RPE (Borg scale) was recorded during the same period. Incidences of gastrointestinal (GI) symptoms were recorded by interview after each trial after completion of the cognitive test battery. After completion of both experimental trials, participants completed an exit interview in which they were asked whether they could identify the KE condition and which trial did they believe that they performed their longest run to exhaustion.

Blood analysis. Blood was collected in plastic tubes (2 mL) containing sodium heparin (Plus Blood Collection Tubes, Becton Dickinson) for subsequent analysis of β HB. A second blood sample was collected in plastic tubes (4 mL) containing sodium fluoride (Plus Blood Collection Tubes, Becton Dickinson). Samples were stored on ice before centrifugation at 3000g for 10 min at 4°C, after which three aliquots of plasma were separated for storage at -80°C until later analysis of plasma β HB, lactate, and glucose (RX Daytona, Randox Laboratories, UK: assay codes RB1007, LC2389, and GL364, respectively).

Statistical analysis. Data were evaluated using Prism 7.0 (GraphPad Software, Inc., CA) and are presented as mean (lower-upper 95% confidence interval [CI] of the mean), except the participant characteristics, which are described as mean ± SD. A paired samples *t*-test was used to determine differences between trials in run time to exhaustion and average HR during part B. Two-way (time-condition) repeated-measures ANOVA was used to determine differences between the two experimental trials for all other with variables with serial measurements. When a main effect of condition or an interaction effect between condition

and time was indicated, *post hoc* testing was performed with Bonferroni's correction with multiplicity-adjusted *P* values applied to compare KE to PLA at the respective time points. The data were tested for normality using the Shapiro-Wilk test before proceeding with the parametric tests described. For null hypothesis statistical testing, the significance level was set at $\alpha = 0.05$ for all tests. Apart from and independent of the outcome of the repeated-measures ANOVA, standardized differences in the mean were used to assess magnitudes of effects between conditions at respective time points. These effect sizes were calculated using Cohen's *d* and interpreted using thresholds of <0.25, >0.25, >0.5, and >1.0 for trivial, small, moderate, and large, respectively (32).

RESULTS

Preexercise hydration status. Hydration status, measured as urine osmolality before each trial, did not differ between trials (mean [95% CI]: KE, 420 [259–581] mOsm·kg⁻¹; PLA, 460 [189–732] mOsm·kg⁻¹; *P* = 0.645, *d* = 0.18).

Plasma β HB, glucose, and lactate concentrations. Fasting plasma concentrations of β HB (mean [95% CI]: KE, 0.11 [0.09–0.13] mM; PLA, 0.11 [0.09–0.13] mM; *P* > 0.99), glucose (KE, 4.81 [4.62–5.00] mM; PLA, 4.84 [4.57–5.01] mM; *P* > 0.99), and lactate (KE, 0.86 [0.71–1.02] mM; PLA, 0.96 [0.82–1.11] mM; *P* > 0.99) concentrations did not differ between trials (Fig. 2). A main effect of time and condition (both *P* < 0.001) and a time-condition interaction effect (*P* < 0.001) were observed for plasma β HB concentrations (Fig. 2A). Ingestion of KE resulted in a rise in plasma β HB concentrations to 1.05 mM (95% CI = 0.83–1.26 mM) (*P* < 0.001) by the start of exercise. Concentrations continued to rise throughout exercise with the highest concentrations during KE observed at cessation of shuttle run to exhaustion at 2.61 mM (95% CI = 2.03–3.10 mM) (*P* < 0.001).

A main effect of time (*P* < 0.001) and condition (*P* = 0.020) was observed for plasma glucose concentrations (Fig. 2B). Plasma glucose concentrations were lower during KE compared with PLA at each time point except block 3, but *post hoc* pairwise comparisons did not reveal significant differences between conditions at any time point. However, standardized differences in the mean indicated moderate effect sizes at each of these time points. Specifically, plasma glucose concentrations were lower during KE compared with PLA by 10.9% (–0.56 [–1.48 to 0.35] mM; *d* = 0.52) after block 1, 7.5% (–0.48 [–1.40 to 0.44] mM; *d* = 0.56) after block 2, 11.6% (–0.76 mM [–1.68 to 0.15] mM; *d* = 0.80) after block 4, and 8.4% (–0.55 [–1.46 to 0.37] mM; *d* = 0.56) after block 5. There was no difference in plasma glucose concentration at the end of the shuttle run to exhaustion (KE, 6.49 [6.01–6.90] mM; PLA, 6.46 [5.64–7.23] mM; *P* > 0.99, *d* = 0.04).

A main effect of time and condition (both *P* < 0.001) and a time-condition interaction effect (*P* = 0.009) were

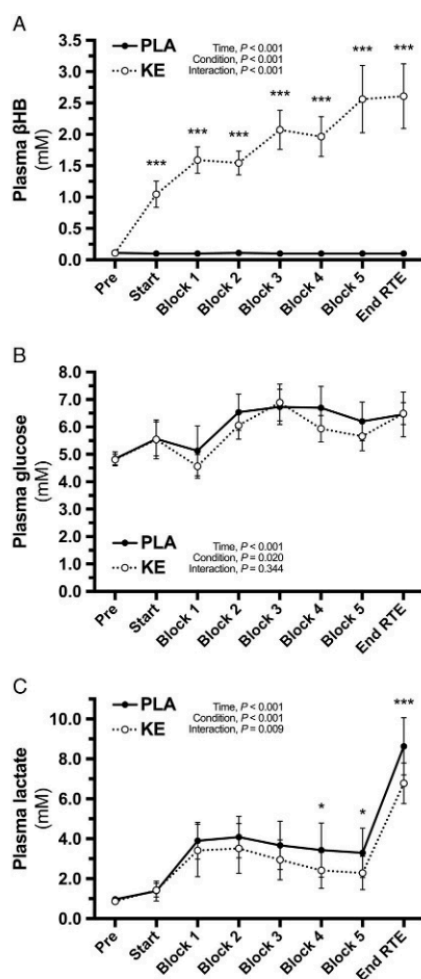


FIGURE 2—Plasma β HB (A), glucose (B), and lactate (C) concentrations during each trial. Data are presented as mean values, with error bars representing 95% CI. * $P < 0.05$ KE vs PLA; *** $P < 0.001$ KE vs PLA.

observed for plasma lactate concentrations (Fig. 2C). Plasma lactate concentrations were elevated from block 1 onward during both conditions but were lower during KE compared with PLA at each time point. *Post hoc* pairwise comparisons revealed significant differences between conditions at block 4 ($P = 0.037$), block 5 ($P = 0.042$), and the end of the shuttle run to exhaustion ($P < 0.001$), and effect sizes indicating small, moderate, and large effects across all time points, i.e., block 1: -11.9% , -0.48 mM (-1.46 to 0.50), $d = 0.27$; block 2: -13.8% , -0.56 mM (-1.54 to 0.42), $d = 0.33$; block 3: -20.2% , -0.73 mM (-1.71 to 0.25), $d = 0.45$; block 4: -29.3% , -1.02 mM (-2.00 to -0.04), $d = 0.58$;

block 5: -30.1% , -1.00 mM (-1.98 to 0.02), $d = 0.63$; end of the shuttle run to exhaustion: -21.5% , -1.85 mM (-2.83 to -0.87), $d = 1.00$).

HR and RPE. For both HR and RPE, main effects of time were observed (both $P < 0.001$), but the absence of main effects of condition or time-condition interaction effects indicates that ingestion of KE did not alter the HR or RPE response during any block of part A of the LIST protocol (Fig. 3). However, the average HR during the shuttle run to exhaustion was lower (-3.9 [-6.4 to -1.4] bpm; $P = 0.007$, $d = 0.42$) during KE (170.7 [163.4 – 177.9] bpm) compared with PLA (174.6 [168.3 – 180.8] bpm).

Fifteen-meter sprint times and shuttle run time to exhaustion. A main effect of time was observed for 15-m sprint times during part A ($P < 0.001$), but no main effect of condition or time-condition interaction effect was observed (Fig. 4A). There was no statistically significant difference in the shuttle run time to exhaustion (KE, 229 [178 – 280] s; PLA, 267 [199 – 336] s; $P = 0.126$), but standardized differences in the mean indicated a small effect size for this difference (-38 [-89 to 13] s; $d = 0.45$).

Cognitive performance. A time-condition interaction effect ($P = 0.021$) was observed for the number of incorrect responses in the executive function multitasking test, which increased from pre- to postexercise in PLA (1.8 [-0.6 to 4.1]), but not in KE (0.0 [-1.8 to 1.8]) ($P = 0.017$, $d = 0.70$) (Table 1). The absence of main effects for time or condition and the time-condition interaction effects indicate that there

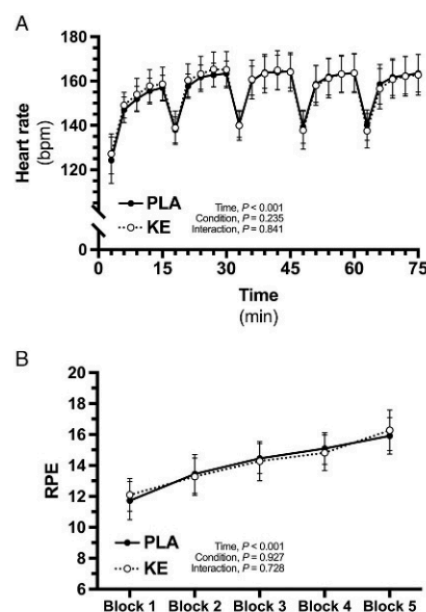


FIGURE 3—HR (A) and RPE (B) during each trial. Data are presented as mean values, with error bars representing 95% CI.

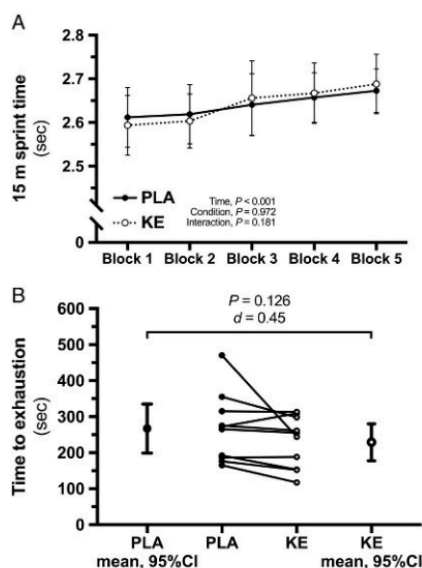


FIGURE 4—Fifteen-meter sprint times (A) and shuttle run time to exhaustion (B) during each trial. Data are presented as mean values, with error bars representing 95% CI.

was no difference between conditions in either reaction time or rapid visual information processing assessed by a sustained attention task (Table 1).

GI symptoms. Out of 11 participants, 4 (36%) reported symptoms of GI distress during PLA and comprised 4 (36%), 3 (27%), 3 (27%), 1 (9%), and 1 (9%) reports of belching, cramps, flatulence, boating, and nausea, respectively. Out of 11 participants, 9 (82%) reported symptoms of GI distress during KE. These comprised 7 (64%), 6 (55%), 4 (36%), 3 (27%), 3 (27%), and 1 (9%) of the participants reporting nausea, cramps, belching, heartburn, flatulence, and vomiting, respectively.

Identification of KE and best performance trials. Out of 11 participants, 8 (73%) correctly identified the trial in which they received KE, identifying KE by the awareness of taste and GI symptoms. However, only five (45%) of the participants correctly identified the trial in which they performed better in the part B run to exhaustion. Only three participants (27%) stated that they believed KE ingestion improved their performance, and two out of those three participants correctly identified their KE trial and their best performance.

DISCUSSION

The aim of the present study was to investigate the effect, if any, of the acute ingestion of a ketone ester on metabolic responses, physical performance, and cognitive performance in team sport athletes in response to an intermittent running

TABLE 1. Cognitive performance measures assessed before and after each trial.

	Reaction Time Test				Errors			
	Pre	Post	Post-Pre	Reaction Time (ms)	Pre	Post	Post-Pre	Errors
KE	393 (360 to 425)	394 (369 to 419)	1 (16 to 18)	218 (177 to 259)	0.4 (-0.1 to 0.8)	0.8 (0.0 to 1.5)	0.4 (-0.6 to 1.4)	0.4 (-0.6 to 1.4)
PLA	404 (371 to 436)	397 (368 to 427)	-8 (-31 to 19)	237 (205 to 269)	0.8 (-0.2 to 1.7)	0.6 (-0.6 to 1.8)	-0.1 (-1.0 to 0.7)	-0.1 (-1.0 to 0.7)
<i>d</i>			-0.28					-0.46
	Multi-tasking Test				Incorrect Responses**			
	Pre	Post	Post-Pre	Reaction Time (ms)	Pre	Post	Post-Pre***	Incorrect Responses**
KE	590 (510 to 669)	550 (483 to 616)	-40 (-78 to -2)	157.9 (156.0 to 159.7)	2.1 (0.7 to 3.6)	2.1 (0.3 to 4.0)	0.0 (-1.8 to 1.8)	2.1 (0.3 to 4.0)
PLA	589 (526 to 652)	543 (499 to 587)	-46 (-74 to -19)	156.0 (152.8 to 159.2)	2.3 (0.2 to 4.3)	4.0 (0.8 to 7.2)****	1.8 (-0.6 to 4.1)	4.0 (0.8 to 7.2)****
<i>d</i>			-0.16				0.70	
	Rapid Visual Information Processing Test				False Alarms			
	Pre	Post	Post-Pre	Reaction Time (ms)	Pre	Post	Post-Pre	False Alarms
KE	449 (389 to 509)	430 (365 to 495)	-19 (-65 to 26)	44.0 (36.7 to 52.3)	2.0 (0.9 to 3.1)	2.3 (0.5 to 4.0)	0.3 (-0.9 to 1.4)	2.3 (0.5 to 4.0)
PLA	460 (395 to 425)	446 (383 to 510)	-14 (-43 to 16)	45.3 (39.0 to 51.5)	2.4 (1.1 to 3.6)	2.5 (0.8 to 4.2)	0.1 (-1.2 to 1.4)	2.5 (0.8 to 4.2)
<i>d</i>			0.12				-0.06	

Data are presented as mean (95% CI), *n* = 8. Effect size calculated as Cohen's *d*.

**P* < 0.01 for main effect of time.

***P* < 0.05 for condition-time interaction effect.

****P* < 0.05 for KE vs PLA.

*****P* < 0.05 for Post vs Pre.

protocol that simulated soccer play. Compared with carbohydrate ingestion alone (PLA), ingestion of the ketone ester with carbohydrate (KE) resulted in an elevation in plasma β HB concentration to >1.5 mM after 15 min of exercise and reached ~ 2.6 mM by the end of exercise. Metabolic consequences included reductions in plasma glucose and lactate concentrations compared with PLA, but no differences in HR or RPE were observed between conditions. KE was without benefit to 15-m sprint times throughout the simulated protocol, or endurance capacity measured by shuttle run to exhaustion. However, cognitive performance in a multitasking executive function test was preserved with KE but declined during PLA.

Recent reports have investigated the effect of acute ingestion of exogenous ketone supplements on physical performance in endurance athletes (10,14,15). Ingestion of a β HB KME increased distance covered in a 30-min cycling time trial by $\sim 2\%$ (411 ± 162 m) (10), whereas, by contrast, ingestion of an acetoacetate KDE impaired performance in a 31.2-km cycling time trial by $2\% \pm 1\%$ (58.2 s) (15). The latter effect was explained by a reduction in average power output by 3.7% and coincided with a high prevalence of GI distress. In addition, ingestion of a β HB salt formulation had no effect on average power output during a 4-min maximal performance cycling test (14). A key distinction between these studies is the form of exogenous ketone supplement ingested. Acute ingestion of KME produces plasma β HB concentrations of ~ 3.0 mM after 20 min, but the KDE and racemic ketone salts result in blood β HB concentrations only in the 0.3- to 0.6-mM range (14–16). Although this concentration range constitutes acute nutritional ketosis and is sufficient to affect the metabolic response to exercise (12,16), performance is unlikely to be affected unless circulating β HB concentrations exceed 1.0 mM (20). In the present study, the KE condition resulted in an elevation of plasma β HB concentrations to >1.5 mM after 15 min of exercise and reached ~ 2.6 mM by the end of exercise, which is broadly similar to previous work (9,10). Our participants ingested $750 \text{ mg} \cdot \text{kg}^{-1}$ body mass split across three boluses with 50% ingested 20 min before commencing exercise, and the remainder split into aliquots of 25% ingested at 30 and 60 min of exercise, respectively. This feeding strategy mimics the previous work with KME ingestion and exercise performance, which resulted in plasma β HB concentrations of ~ 2 mM 20 min after ingestion and ranged from ~ 2.0 to 3.0 mM throughout 90 min of exercise (10). By contrast, our participants only achieved these levels 65 min after ingestion of the initial bolus of KE, but these participants ingested KE in the postprandial state as opposed to the fasted state in the KME work. Ingestion of KME in the postprandial state can attenuate the C_{max} of blood β HB concentrations by 33% and the 4-h β HB AUC by 27% (9).

Accordingly, despite the similar changes in circulating β HB concentrations to the previous investigation of performance effects using the KME (10), the KE condition in the present study was without benefit to 15-m sprint times

throughout the 75-min intermittent running protocol (LIST part A), or on shuttle run time to exhaustion (LIST part B). The metabolic consequences of KME ingestion were recapitulated herein, namely, lower plasma glucose and lactate concentrations during KE compared with PLA. A 10% reduction in plasma glucose was observed 35 min after ingestion of the initial KE bolus and was 8% to 12% lower (moderate effects) during blocks 1, 2, 4, and 5 of part A. The glucose-lowering effect of exogenous ketones is well documented whether ingested alone (9,10,12,16,33) or in combination with carbohydrate and/or protein (9,10,13,15,34). Although the insulinotropic action of ketone bodies is not always observed (5,6), it can occur under certain conditions (2), including when KME is ingested in the fasted state (9,10,33). However, when coingested with carbohydrate and/or protein, the glucose-lowering effect of exogenous ketones occurs despite similar circulating insulin concentrations in response to carbohydrate and/or protein alone at rest (34), during exercise (10), and during recovery from exercise (13). A β HB-mediated glucose-lowering effect is likely a result of an attenuation of hepatic gluconeogenesis and increase in hepatic glucose uptake (6). The rise in plasma lactate concentrations was attenuated during KE compared with carbohydrate ingestion alone, consistent with previous KME work (10). An attenuation in the exercise-induced rise in plasma lactate was previously explained by a reduction in glycolytic flux, sparing of muscle glycogen during exercise, and an increased contribution of ketone bodies and intramuscular triglycerides to energy provision (10). Whereas a 50% reduction in the rise in plasma lactate was observed during a 60-min preloaf at 75% \dot{W}_{max} and 30 min time trial in trained cyclists (10), we observed a reduction ranging from $\sim 10\%$ to 30%. Given the lower aerobic fitness in our team sport athletes, the trained cyclists may have had a greater capacity to extract ketones from circulation and oxidize them as a substrate, resulting in a larger contribution toward total energy production and a greater reduction in glycolytic flux. This is because ketone bodies are transported across the skeletal muscle membrane by monocarboxylate transporters, which are most highly expressed in type I muscle fibers, and are increased in response to endurance exercise training (35).

For that reason, we previously hypothesized that performance benefits of exogenous ketones are most likely to be realized in those individuals with high levels of aerobic fitness and higher proportions of type I muscle fibers and/or monocarboxylate transporter expression (20). A lower level of aerobic fitness and training status, and therefore lesser ability to oxidize circulating ketones, may be one explanation for the lack of performance benefit in the present work. That notwithstanding, our performance test was shorter (~ 2 to 6 min) and intermittent in nature, which may be another factor contributing to the contrasting results. Another explanation may relate to the proposed benefits of exogenous ketone supplements being via their glycogen sparing effect. Given that we used an optimal carbohydrate-based fuelling on the

day before and the day of each trial, our athletes may not have experienced glycogen depletion to an extent that the purported glycogen sparing would have benefited performance in part B. In fact, standardized differences in the mean used to assess magnitudes of effects between KE and PLA indicate a small effect size for a decrement in performance with KE, so it would be remiss not to consider that the effect of exogenous ketones in this instance may have been to impair carbohydrate utilization.

Nutrition strategies such as high fat feeding, ketogenic diets, and exogenous ketone ingestion alter substrate utilization during exercise, which generally results in lower rates of carbohydrate utilization at moderate-to-high exercise intensities (18–20). Whether this shift in substrate utilization reflects a sparing of muscle glycogen, which can then be used later in an exercise challenge, or instead reflects an impairment of muscle glycogen utilization during such exercise intensities is a salient issue for alternative fuelling strategies. The mechanistic basis for reduced carbohydrate utilization in the presence of exogenous ketones is proposed as an attenuation of glycolytic flux via inhibition of pyruvate dehydrogenase (PDH) and phosphofructokinase by increases in NADH:NAD⁺, acetyl-CoA:CoA, or citrate. A similar mechanism is likely to contribute to the impaired performance during moderate- to high-intensity efforts observed under high fat feeding (36,37). The attenuation of PDH activity under such conditions (38) could be problematic for intermittent activity sports that require high-intensity efforts, which rely heavily on ATP provision from glycolytic pathways, performed on a moderate-intensity background. Clearly, this is the nature of the exercise challenge in the present study, but future work will require direct measurement of PDH activity and glycolytic flux in muscle biopsies to make definitive conclusions about the effects of exogenous ketones on utilization of muscle glycogen in this model. Conversely, we observed no benefit or decrement on 15-m sprint times performed at a rate of approximately nine sprints per 15-min block across 75 min of intermittent activity. Maximal short duration sprints rely primarily on the ATP–phosphocreatine system and anaerobic glycolysis for energy provision, but as the number of repeated sprints increases, the contribution of both energy systems decline and the contribution of aerobic glycolysis of circulating glucose and muscle glycogen increase over time (39,40). The lower plasma lactate concentrations in part A during KE suggests a reduction in glycolytic flux, but the reduction ultimately did not affect performance in repeated sprints of <3-s duration.

A higher incidence of GI symptoms occurred during KE compared with PLA, although this did not affect the HR or RPE responses during exercise. GI symptoms are a common side effect of KME and KDE ingestion, and more work is needed on the dose and timing of both these supplements to mitigate this response. KME ingested as part of a meal replacement milkshake drink causes a stepwise increase in symptoms with increasing dosages (7). Furthermore, ingestion of 500 mg·kg⁻¹ body mass of KDE split in two doses

caused symptoms in all participants during a cycling time trial (15). These symptoms are likely to be a large contributor to the performance decrement in that study given the participants' nomination of their symptoms as a distraction or interference to performance. The incidence of GI symptoms was higher in the present study than in previous work with KME (10), but the aforementioned commencement of exercise in a fed as opposed to fasted state, or this protocol involving running as opposed to cycling exercise, may be contributing factors.

A novel finding herein is the preservation of executive function during KE compared with PLA, measured by the number of incorrect responses to a multitasking test. Given that team sport athletes are presented with a multitude of decisions throughout match play, interventions that preserve or improve cognitive performance could positively influence performance outcomes. The primary physiological role of ketogenesis as a survival mechanism during low carbohydrate availability is providing a substrate to the brain in the presence of diminishing blood glucose concentrations (26). Cognitive benefits and a neuroprotective role are established for exogenous ketones in nonexercise contexts (27–29,41). Notably, in a short-term (5-d) feeding study, rats supplemented daily with KME were 38% faster at completing a radial maze task and made more correct decisions before making a mistake during the test (29). This outcome is consistent with our findings and suggests that central effects may be relevant during exercise, although other tests of cognitive function, i.e., reaction time and sustained attention tasks, were unaffected.

In conclusion, in team sport athletes, acute ingestion of a ketone ester elevated plasma β HCB concentrations but did not improve performance in a shuttle run to exhaustion performed after 75 min of intermittent running. Reductions in plasma glucose and attenuated increases plasma lactate during exercise demonstrate the obvious effects of exogenous ketone ingestion on carbohydrate metabolism during exercise. However, participants experienced incidences of GI symptoms. These results underscore the need for future work to explore possible dose–response effects while minimizing any GI distress to athletes. Despite the lack of benefit to physical performance, the novel finding of preserved executive function after exhausting exercise suggests that there remains a possibility that exogenous ketones could enhance sport-specific performance of team sport athletes via other mechanisms.

The authors thank Dr. Sabine Harrison (UCD Institute of Food and Health) for technical assistance during the analysis of blood samples. This work was funded in part by an Irish Research Council Enterprise Partnership Postgraduate Fellowship (EPSPG2016177).

The ketone ester studied in this work was provided in kind by KetoneAid, Inc. KetoneAid provided input into the dosing strategy used but had no role in the implementation, interpretation, or dissemination of this work. No conflict of interest, financial or otherwise, is declared by the authors.

The authors declare that the results of the study are presented clearly, honestly, and without fabrication, falsification, or inappropriate data manipulation and do not constitute endorsement by the American College of Sports Medicine.

REFERENCES

- Robinson AM, Williamson DH. Physiological roles of ketone bodies as substrates and signals in mammalian tissues. *Physiol Rev*. 1980;60(1):143–87.
- Balasse EO, Fery F. Ketone body production and disposal: effects of fasting, diabetes, and exercise. *Diabetes Metab Rev*. 1989;5(3):247–70.
- Féry F, Balasse EO. Response of ketone body metabolism to exercise during transition from postabsorptive to fasted state. *Am J Physiol*. 1986;250(5 Pt 1):E495–501.
- Maizels EZ, Ruderman NB, Goodman MN, Lau D. Effect of acetoacetate on glucose metabolism in the soleus and extensor digitorum longus muscles of the rat. *Biochem J*. 1977;162(3):557–68.
- Nair KS, Welle SL, Halliday D, Campbell RG. Effect of beta-hydroxybutyrate on whole-body leucine kinetics and fractional mixed skeletal muscle protein synthesis in humans. *J Clin Invest*. 1988;82(1):198–205.
- Mikkelsen KH, Seifert T, Secher NH, Grondal T, van Hall G. Systemic, cerebral and skeletal muscle ketone body and energy metabolism during acute hyper-D- β -hydroxybutyratemia in post-absorptive healthy males. *J Clin Endocrinol Metab*. 2015;100(2):636–43.
- Clarke K, Tchabanenko K, Pawlosky R, et al. Kinetics, safety and tolerability of (R)-3-hydroxybutyl (R)-3-hydroxybutyrate in healthy adult subjects. *Regul Toxicol Pharmacol*. 2012;63(3):401–8.
- Kesl SL, Poff AM, Ward NP, et al. Effects of exogenous ketone supplementation on blood ketone, glucose, triglyceride, and lipoprotein levels in Sprague-Dawley rats. *Nutr Metab (Lond)*. 2016;13:9.
- Stubbs BJ, Cox PJ, Evans RD, et al. On the metabolism of exogenous ketones in humans. *Front Physiol*. 2017;8:848.
- Cox PJ, Kirk T, Ashmore T, et al. Nutritional ketosis alters fuel preference and thereby endurance performance in athletes. *Cell Metab*. 2016;24(2):256–68.
- Holdsworth DA, Cox PJ, Kirk T, Stradling H, Impey SG, Clarke K. A ketone ester drink increases postexercise muscle glycogen synthesis in humans. *Med Sci Sports Exerc*. 2017;49(9):1789–95.
- O'Malley T, Myette-Côté E, Durrer C, Little JP. Nutritional ketone salts increase fat oxidation but impair high-intensity exercise performance in healthy adult males. *Appl Physiol Nutr Metab*. 2017;42:1031–1035.
- Vandoorne T, De Smet S, Ramaekers M, et al. Intake of a ketone ester drink during recovery from exercise promotes mTORC1 signaling but not glycogen resynthesis in human muscle. *Front Physiol*. 2017;8:310.
- Rodger S, Plews D, Laursen P, Driller MW. Oral β -hydroxybutyrate salt fails to improve 4-minute cycling performance following submaximal exercise. *J Sci Cycling*. 2017;6(1):26–31.
- Leckey JJ, Ross ML, Quod M, Hawley JA, Burke LM. Ketone diester ingestion impairs time-trial performance in professional cyclists. *Front Physiol*. 2017;8:806.
- Evans M, Patchett E, Nally R, Kearns R, Lamey M, Egan B. Effect of acute ingestion of β -hydroxybutyrate salts on the response to graded exercise in trained cyclists. *Eur J Sport Sci*. 2018;18(3):376–86.
- Waldman HS, Basham SA, Price FG, et al. Exogenous ketone salts do not improve cognitive responses after a high-intensity exercise protocol in healthy college-aged males. *Appl Physiol Nutr Metab*. 2018;43(7):711–7.
- Burke LM. Re-examining high-fat diets for sports performance: did we call the 'nail in the coffin' too soon? *Sports Med*. 2015;45(1 Suppl):S33–49.
- Pinckaers PJ, Churchward-Venne TA, Bailey D, van Loon LJ. Ketone bodies and exercise performance: the next magic bullet or merely hype? *Sports Med*. 2017;47(3):383–91.
- Evans M, Cogan KE, Egan B. Metabolism of ketone bodies during exercise and training: physiological basis for exogenous supplementation. *J Physiol*. 2017;595(9):2857–71.
- Spencer M, Bishop D, Dawson B, Goodman C. Physiological and metabolic responses of repeated-sprint activities: specific to field-based team sports. *Sports Med*. 2005;35(12):1025–44.
- Cummins C, Orr R, O'Connor H, West C. Global positioning systems (GPS) and microtechnology sensors in team sports: a systematic review. *Sports Med*. 2013;43(10):1025–42.
- Burke LM, Loucks AB, Broad N. Energy and carbohydrate for training and recovery. *J Sports Sci*. 2006;24(7):701–13.
- Jacobs I, Westlin N, Karlsson J, Rasmussen L. Muscle glycogen and diet in elite soccer players. *Eur J Appl Physiol Occup Physiol*. 1982;48(3):297–302.
- Mohr M, Krstrup P, Bangsbo J. Match performance of high-standard soccer players with special reference to development of fatigue. *J Sports Sci*. 2003;21(7):519–28.
- Owen OE, Morgan AP, Kemp HG, Sullivan JM, Herrera MG, Cahill GF Jr. Brain metabolism during fasting. *J Clin Invest*. 1967;46(10):1589–95.
- Ari C, Kovács Z, Juhasz G, et al. Exogenous ketone supplements reduce anxiety-related behavior in Sprague-Dawley and Wistar albino Glaxo/Rijswijk rats. *Front Mol Neurosci*. 2016;9:137.
- Kovács Z, D'Agostino DP, Dobolyi A, Ari C. Adenosine A1 receptor antagonism abolished the anti-seizure effects of exogenous ketone supplementation in Wistar albino Glaxo/Rijswijk rats. *Front Mol Neurosci*. 2017;10:235.
- Murray AJ, Knight NS, Cole MA, et al. Novel ketone diet enhances physical and cognitive performance. *FASEB J*. 2016;30(12):4021–32.
- Bangsbo J, Iaia FM, Krstrup P. The Yo-Yo intermittent recovery test: a useful tool for evaluation of physical performance in intermittent sports. *Sports Med*. 2008;38(1):37–51.
- Nicholas CW, Nuttall FE, Williams C. The Loughborough Intermittent Shuttle Test: a field test that simulates the activity pattern of soccer. *J Sports Sci*. 2000;18(2):97–104.
- Rhea MR. Determining the magnitude of treatment effects in strength training research through the use of the effect size. *J Strength Cond Res*. 2004;18(4):918–20.
- Stubbs BJ, Cox PJ, Evans RD, Cyranka M, Clarke K, de Wet H. A ketone ester drink lowers human ghrelin and appetite. *Obesity*. 2018;26(2):269–73.
- Myette-Côté É, Neudorf H, Rafiei H, Clarke K, Little JP. Prior ingestion of exogenous ketone monoester attenuates the glycaemic response to an oral glucose tolerance test in healthy young individuals. *J Physiol*. 2018;596(8):1385–95.
- Thomas C, Bishop DJ, Lambert K, Mercier J, Brooks GA. Effects of acute and chronic exercise on sarcolemmal MCT1 and MCT4 contents in human skeletal muscles: current status. *Am J Physiol Regul Integr Comp Physiol*. 2012;302(1):R1–14.
- Havemann L, West SJ, Goedecke JH, et al. Fat adaptation followed by carbohydrate loading compromises high-intensity sprint performance. *J Appl Physiol*. 2006;100(1):194–202.
- Burke LM, Ross ML, Garvican-Lewis LA, et al. Low carbohydrate, high fat diet impairs exercise economy and negates the performance benefit from intensified training in elite race walkers. *J Physiol*. 2017;595(9):2785–807.
- Stellingwerff T, Spriet LL, Watt MJ, et al. Decreased PDH activation and glycogenolysis during exercise following fat adaptation with carbohydrate restoration. *Am J Physiol Endocrinol Metab*. 2006;290(2):E380–8.
- Gaitanos GC, Williams C, Boobis LH, Brooks S. Human muscle metabolism during intermittent maximal exercise. *J Appl Physiol*. 1993;75(2):712–9.
- Parolin ML, Chesley A, Matsos MP, Spriet LL, Jones NL, Heigenhauser GJ. Regulation of skeletal muscle glycogen phosphorylase and PDH during maximal intermittent exercise. *Am J Physiol*. 1999;277(5 Pt 1):E890–900.
- Svart M, Gormsen LC, Hansen J, et al. Regional cerebral effects of ketone body infusion with 3-hydroxybutyrate in humans: reduced glucose uptake, unchanged oxygen consumption and increased blood flow by positron emission tomography. A randomized, controlled trial. *PLoS One*. 2018;13(2):e0190556.

Search documents



. . . Published ahead of Print

**No Benefit of Ingestion of a Ketone Monoester Supplement
on 10-km Running Performance**

Mark Evans¹, Fionn T. McSwiney², Aidan J. Brady¹, Brendan Egan^{1,3}

¹School of Health and Human Performance, Dublin City University, Glasnevin, Dublin, Ireland;

²School of Health Sciences, Waterford Institute of Technology, Waterford, Ireland; ³National Institute for Cellular Biotechnology, Dublin City University, Glasnevin, Dublin, Ireland

Accepted for Publication: 21 May 2019

Medicine & Science in Sports & Exercise® Published ahead of Print contains articles in unedited manuscript form that have been peer reviewed and accepted for publication. This manuscript will undergo copyediting, page composition, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered that could affect the content.

Copyright © 2019 American College of Sports Medicine

No Benefit of Ingestion of a Ketone Monoester Supplement on 10-km Running Performance

Mark Evans¹, Fionn T. McSwiney², Aidan J. Brady¹, Brendan Egan^{1,3}

¹School of Health and Human Performance, Dublin City University, Glasnevin, Dublin, Ireland;

²School of Health Sciences, Waterford Institute of Technology, Waterford, Ireland; ³National Institute for Cellular Biotechnology, Dublin City University, Glasnevin, Dublin, Ireland

CORRESPONDING AUTHOR

Brendan Egan, PhD

School of Health and Human Performance

Dublin City University

Glasnevin, Dublin 9

Ireland

e: brendan.egan@dcu.ie

t: +353 1 7008803

f: +353 1 7008888

This work was funded in part by an Irish Research Council Enterprise Partnership Postgraduate Fellowship (EPSPG2016177). The ketone monoester studied in this work was purchased from HVMN, Inc. (San Francisco, USA), and we thank Dr. Brianna Stubbs (HVMN, Inc.) for advice regarding the formulation and flavoring of the placebo condition. HVMN had no role in the implementation, interpretation, or dissemination of this work. No conflict of interest, financial or otherwise, is declared by the authors. The authors declare the results of the study are presented clearly, honestly, and without fabrication, falsification, or inappropriate data manipulation and do not constitute endorsement by the American College of Sports Medicine.

ABSTRACT

Purpose: Pre-exercise ingestion of exogenous ketones alters the metabolic response to exercise, but effects on exercise performance have been equivocal. **Methods:** On two occasions in a double-blind, randomized crossover design, eight endurance-trained runners performed 1 h of submaximal exercise at ~65% $\text{VO}_{2\text{max}}$ immediately followed by a 10-km self-paced TT on a motorized treadmill. An 8% carbohydrate-electrolyte solution was consumed before and during exercise, either alone (CHO+PLA), or with 573 $\text{mg}\cdot\text{kg}^{-1}$ of a ketone monoester supplement (CHO+KME). Expired air, heart rate (HR), and rating of perceived exertion (RPE) were monitored during submaximal exercise. Serial venous blood samples were assayed for plasma glucose, lactate and β -hydroxybutyrate concentrations. **Results:** CHO+KME produced plasma β -hydroxybutyrate concentrations of ~1.0 to 1.3 mM during exercise ($P < 0.001$), but plasma glucose and lactate concentrations were similar during exercise in both trials. VO_2 , running economy, respiratory exchange ratio, HR and RPE were also similar between trials. Performance in the 10-km TT was not different ($P = 0.483$) between CHO+KME (mean = 2402 s; 95% confidence interval [CI] = 2204, 2600 s) and CHO+PLA (mean = 2422 s; 95% CI = 2217, 2628 s). Cognitive performance, measured by reaction time and a multi-tasking test, did not differ between trials. **Conclusion:** Compared with carbohydrate alone, co-ingestion of KME by endurance-trained athletes elevated plasma β -hydroxybutyrate concentrations, but did not improve 10-km running TT or cognitive performance.

KEYWORDS: athletes; β -hydroxybutyrate; cognition; endurance; lactate; time trial;

INTRODUCTION

The therapeutic and performance potential of exogenous ketone supplements has been the subject of increasing interest in recent years (1, 2). Metabolic effects the ketone bodies (KB), namely β -hydroxybutyrate (β HB) and acetoacetate (AcAc), are well-established in many organs, including attenuation of glycolysis, hepatic glucose output and adipose tissue lipolysis (3), but their potential role in modulating substrate utilization has garnered attention for athletic performance (4, 5). In the fasted state, KB provide up to 10% of energy to skeletal muscle during exercise (6), and after acute ingestion of exogenous ketone supplements, this contribution can apparently increase to 16 to 18% when circulating β HB is elevated to the 3 to 4 mM range (5). Moreover, this increase in β HB oxidation coincides with a reduction in glycolytic flux, as evidenced by an attenuation in the exercise-induced rise in plasma lactate and glycolytic intermediates, and an increase in intramuscular triglyceride utilization during exercise (5).

Circulating KB concentrations are <0.1 mM in the postprandial state, whereas hyperketonaemia is accepted as KB concentrations exceeding 0.2 mM (3). Ingestion of a variety of exogenous ketone supplements can acutely produce nutritional ketosis (4, 5, 7-17), which has been defined as circulating KB concentrations >0.5 mM (18). The most potent of these exogenous ketone supplements is the (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester (KME). When ingested at rest in the fasted state, KME produces a dose-dependent increase in circulating β HB concentrations of up to 6 mM 20 min after the ingestion of up to 573 mg kg^{-1} body mass (5, 9). This elevation in β HB concentration coincides with decreases in plasma glucose, free fatty acids (FFAs), triglycerides and ghrelin concentrations (5, 8, 9, 13). Exercise attenuates the rise in β HB concentrations, as ingestion of 573 mg. kg^{-1} body mass KME prior to 45 min cycling at 45% and 75% peak power output (W_{max}) resulted in circulating β HB of ~ 4.0 mM and ~ 3.0 mM, respectively. As a consequence of the aforementioned effects on

substrate utilization, acute ingestion of KME attenuates the rise in plasma glucose and lactate concentrations during exercise, whether in an endurance cycling or intermittent running context (4, 5).

These metabolic consequences have been proposed to explain the observation that the co-ingestion of KME in addition to a carbohydrate-based fuelling strategy improved performance in a 30 min maximum distance cycling time-trial by 2% when preceded by 1 h of submaximal 'pre-load' exercise (5). In contrast, high-intensity shuttle running capacity (~4 to 6 min) performed after 75 min of intermittent running was not improved in team sport athletes with KME co-ingestion compared to carbohydrate alone (4). While the former study considered a 'sparing' of muscle glycogen to be major factor in the performance benefit (5), the latter study speculated that the attenuation of glycolytic flux in the presence of elevated circulating β HB may have been a factor in the lack of performance benefit in that exercise model (4). Performance in exercise of long duration that incorporates high intensity efforts (i.e. sprint finishes, climbs) is largely dependent on carbohydrate utilization (19). Therefore, nutrition strategies that could spare muscle glycogen and maintain high intensities in the latter parts of races are of interest to scientists and practitioners (20). However, if glycogen sparing occurs via an attenuation of glycolytic flux that cannot be overcome when higher intensity efforts are required, this would instead be likely to impair performance (19). Moreover, the recent observation that acute ingestion of KME prior to intermittent exercise in team sport athletes resulted in preserved executive function as measured by a decision-making task after volitional exhaustion (4), remains to be confirmed in other exercise settings.

Therefore, the aim of the present study was to investigate the effects of acute ingestion of an exogenous ketone supplement in the form of a commercially-available KME on physiological

responses, and physical and cognitive performance in endurance-trained runners in response to 1 h submaximal exercise immediately followed by a 10-km time trial.

METHODS

Participants. Eight trained, middle and long distance runners (M/F, 7/1; age, 33.5 ± 7.3 y; height, 1.79 ± 0.07 m; body mass, 68.8 ± 9.7 kg; body fat, $8.0 \pm 4.1\%$; $\text{VO}_{2\text{max}}$, 62.0 ± 5.6 mL.kg⁻¹.min⁻¹) gave written informed consent to participate after written and verbal explanations of the procedures. Ethical approval (permit number: DCUREC2018_039) was obtained from the Dublin City University Research Ethics Committee in accordance with the Declaration of Helsinki.

Experimental design. Participants visited the laboratory for exercise tests on four separate occasions over a 21 to 28 day period, comprising one baseline, one familiarization and two main experimental trials. During their first visit to the lab, each participant's maximal rate of oxygen consumption ($\text{VO}_{2\text{max}}$) was determined using an incremental treadmill test to volitional exhaustion. The exercise protocol performed in the familiarization visit (visit 2) and two main experimental trials (visits 3 and 4) comprised of a pre-load of 1 h of treadmill running at $65\% \text{VO}_{2\text{max}}$ followed by a self-paced 10-km time-trial (TT) performance test performed on a motorized treadmill (Fig. 1). A battery of cognitive tests was performed before and after the exercise protocol. The main experimental trials were performed in a double-blind, placebo-controlled, randomized crossover design. Visits 2, 3 and 4 were identical in terms of the pre-test preparation (standardized physical activity and diet for 24 h prior to each visit) and the exercise protocol. The visits differed only in the drinks consumed before and during exercise, namely an 8% carbohydrate-electrolyte solution, which was co-ingested with either a flavored placebo condition (CHO+PLA), or included the (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester (CHO+KME). The primary outcome was endurance performance measured by time to

complete the self-paced 10-km TT, with secondary outcomes including cognitive performance, oxygen consumption (VO_2), running economy, respiratory exchange ratio (RER), heart rate (HR), rating of perceived exertion (RPE), and plasma βHB , glucose, and lactate concentrations.

Assessment of $\text{VO}_{2\text{max}}$ and submaximal running speeds. Body mass was measured to the nearest 0.2 kg using a calibrated digital scales (SECA, Hamburg, Germany), and height was measured to the nearest 0.01 m using a wall-mounted stadiometer (Holtain, Crymych, UK). Body fat was determined by bioelectrical impedance analysis (DC-430U Dual Frequency Analyzer; Tanita, Arlington Heights, IL USA). All exercise testing and experimental trials were conducted on a motorized treadmill (T200; COSMED, Rome Italy). Initially, for the determination of the responses in VO_2 and blood lactate concentration at submaximal running speeds, participants ran for 4 min stages at a progressively increasing speeds, interspersed with a 1 min rest interval for determination of blood lactate concentrations (Lactate Pro 2; Arkay, Kyoto, Japan), RPE (Borg scale) and HR (Polar H7; Polar, Kempele Finland). The first stage was $4 \text{ km}\cdot\text{h}^{-1}$ slower than the average speed corresponding to each participant's personal best time for a 10-km race. For each subsequent stage, the running speed was increased by $1 \text{ km}\cdot\text{h}^{-1}$ until the running speed exceeded the speed corresponding to their personal best 10-km race speed. After a 10 min rest, participants began running at a speed corresponding to the last completed speed of the preceding test. Treadmill speed was increased by $2.0 \text{ km}\cdot\text{h}^{-1}$ every 2 min for two stages, after which treadmill gradient was increased by 1.0% every 1 min until volitional fatigue. Expired air was collected and analyzed throughout these tests using the Quark RMR metabolic cart (COSMED, Rome, Italy). VO_2 , carbon dioxide production (VCO_2), and RER were calculated from an average of breath-by-breath measurements during the last 30 s of each stage during the submaximal running stages and the assessment of $\text{VO}_{2\text{max}}$. $\text{VO}_{2\text{max}}$ was considered to have been achieved if two of the following criteria were achieved: (i) plateauing of

VO₂ despite increasing treadmill speed (increase in VO₂ of less than 2.0 mL.kg⁻¹.min⁻¹), (ii) HR within 5% of the age-predicted HR_{max} (208 – 0.7 x age in years), and (iii) an RER ≥1.10.

Cognitive test battery. The battery of cognitive tests (CANTAB Cognition, Cambridge, UK) was administered via a touch screen tablet lasting ~10 min. An identical test battery was administered before and after each trial in visits 2, 3 and 4.

During the reaction time (RTI) test, participants select and hold a button at the bottom of the screen and five circles are presented above. In each case, a yellow dot appears in one of the five circles, and the participants must react as soon as possible, releasing the button at the bottom of the screen, and selecting the circle in which the dot appeared. Release time (msec), reaction time (msec), and number of errors were recorded.

The multi-tasking test (MTT) is a test of executive function that measures the participant's ability to switch attention between stimuli, and ignore task-irrelevant information. White arrows are displayed on a black background, with the arrows located on either the left or right side of the screen, and pointing either to the left or to the right. A cue is displayed at the same time as the arrows, reading either "SIDE" or "DIRECTION". When the "SIDE" cue is presented, the participant is required to press a button on the left or right of the screen corresponding to the side of the screen where the arrow is presented, regardless of the direction the arrow is pointing. Conversely, when the "DIRECTION" cue is presented, the participants are required to touch a button on the left or right of the screen corresponding to the direction the arrow is pointing, regardless of which side of the screen the arrow is presented. Reaction time (msec), and number of correct and incorrect responses were recorded.

Pre-trial preparation. All experimental trials commenced between 0730 and 1130, and were completed within a period of 4.0-4.5 h (Fig. 1). On an individual basis, participants performed their second main experimental trial at the same time ±1 h as their first main trial. Pre-

trial preparation was the same for the familiarization visit and each main experimental trial. Participants were asked to abstain from alcohol for 48 h and caffeine for 24 h, and refrain from strenuous exercise training on the day prior to each trial. For the day prior to experimental trials, participants were provided with a prescribed meal plan that provided ~2800 kcal (~41 kcal.kg⁻¹) at a macronutrient ratio of 60% carbohydrate (~6.2 g.kg⁻¹), 20% protein and 20% fat. Participants performed the two main experimental trials separated by either 7 or 14 days.

Main experimental trials. The protocol for the familiarization and main experimental trials were identical except for the drinks consumed before and during exercise (Fig. 1). Participants arrived to the laboratory in a fasted state 2 h prior to the commencement of exercise, and immediately consumed a standardized breakfast of quick-cook porridge oats and cereal bars providing ~300-400 kcal (~4.4-5.8 kcal.kg⁻¹) and ~1.0 g.kg⁻¹ of carbohydrate, and 500 mL of water. Participants proceeded to complete the cognitive test battery 45 min after breakfast. Thereafter, an indwelling catheter (21G Insyte Autoguard; Becton Dickinson, Franklin Lakes, NJ USA) was introduced into an antecubital vein for serial blood sampling at rest (-30 and 0 min), during submaximal exercise (20, 40 and 60 min) and immediately after the 10-km TT.

For each trial, a bolus of a given drink was ingested 30 min prior to exercise (drink 1), at 20 min intervals during the 1 h of submaximal running (drinks 2 to 4), and at the 5-km mark of the 10-km TT (drink 5) (Fig. 1). The carbohydrate-based fuelling strategy (CHO) consisted of a 6.4% carbohydrate-electrolyte solution (Lucozade Sport; Lucozade Ribena Suntory Ltd., Uxbridge, UK) with maltodextrin (Cargill Inc, Minneapolis, MN USA) added to make an 8.0% carbohydrate-electrolyte solution that was provided at a rate of ~1.0 g.min⁻¹ of exercise. During CHO+PLA, CHO was supplemented with denatonium benzoate, malic acid and arrow root extract to mimic the bitter taste and mouth-feel of the KME. During CHO+KME, CHO was supplemented with 573 mg.kg body mass⁻¹ of a (R)-3-hydroxybutyl (R)-3-hydroxybutyrate

ketone monoester (HVMN™ Ketone; HVMN, Inc., San Francisco, CA USA). The commercially-available ketone ester was mixed directly with the carbohydrate-electrolyte solution for ingestion, and the 573 mg.kg⁻¹ dose was divided into three boluses at a ratio 50:25:25 ingested at -30 min (drink 1), 20 min (drink 2) and 60 min (drink 4), respectively (Fig. 1). During CHO+PLA, drinks 1, 2 and 4 were flavored with the bitter additives to taste match with CHO+KME, and in both trials, drinks 3 and 5 were provided as the unadulterated 8% carbohydrate-electrolyte solution. All drinks were administered in opaque drinks bottles.

For the exercise protocol, participants first performed a standardized 5 min warm up on the motorized treadmill (8 km.h⁻¹) followed by self-selected stretching. Participants then performed 1 h of treadmill running at a speed corresponding to ~65%VO_{2max} (Table 1). Immediately after completion of the 1 h pre-load, participants completed a 10-km TT. The pre-load followed by TT protocol was modeled on the previous work demonstrating a benefit of KME on cycling TT performance (5), and has been similarly applied to treadmill running in previous studies (21, 22). Prior to each TT, participants were told to complete the distance as fast as possible i.e. to race the 10-km. They were allowed to adjust the treadmill speed as often and by as much as desired by manually-adjusting a side-mounted control panel on the treadmill. Increments or decrements in speed were 0.1 km.h⁻¹ in response to each press of an up or down arrow button, respectively. The 10-km TT began with the participant accelerating from a standing start. Participants were blinded to the speed of the treadmill and the time elapsed at all times, but were aware of the distance covered throughout the TT, including the 5-km mark when drink 5 was provided. After completing the 10-km TT, participants completed the same cognitive test battery as completed prior to exercise.

Venous blood samples were collected at 30 min prior to exercise, at 20 min intervals during submaximal exercise, and immediately after the 10-km TT. HR and RPE were recorded at

20 min intervals during submaximal exercise. Expired air was collected during the first 10 min, 25 to 30 min, and 55 to 60 min of the submaximal exercise for the monitoring of exercise intensity, and calculation of RER and running economy. Running economy is expressed as the volume of oxygen required to run 1 km relative to body mass ($\text{mL.kg}^{-1}.\text{km}^{-1}$) (23). Incidences of gastrointestinal (GI) symptoms were recorded by interview after each trial. At the end of visit 4, participants completed an exit interview in which they were asked whether they could identify the CHO+KME condition, and to identify which experimental trial they believed that they performed their best TT.

Blood analysis. Blood was collected in plastic tubes (2 mL) containing sodium heparin (Plus Blood Collection Tubes; Becton Dickinson, Franklin Lakes, NJ USA) for subsequent analysis of β HB. A second blood sample was collected in plastic tubes (4 mL) containing sodium fluoride (Plus Blood Collection Tubes; Becton Dickinson, Franklin Lakes, NJ USA) for subsequent analysis of glucose and lactate. All collection tubes were pre-chilled, and blood samples were stored on ice before centrifugation at 3000 g for 10 min at 4°C, after which aliquots of plasma were separated for storage at -80°C until later analysis. Plasma β HB was determined by colorimetric assay as per the manufacturer's instructions (MAK041; Sigma-Aldrich, Arklow, Ireland). Plasma glucose and lactate were measured using the RX Daytona™ chemical autoanalyser and appropriate reagents as per the manufacturer's instructions (Randox Laboratories, Crumlin, UK: assay codes GL3815 and LC3980, respectively).

Statistical analysis. The required sample size was calculated *a priori* using performance in the 10-km TT as the primary outcome measure. Based on the reliability data for the pre-loaded 10-km TT protocol employed (22), the assessment of other running TT protocols (24), and the variability of real-world performance in races of similar distance (25), we estimated a coefficient of variation (CV) of 1.5% for performance in the 10-km TT. We aimed to detect a 2.5% change

in 10-km TT performance based on the smallest worthwhile difference (SWD) described by Russell et al. for this pre-loaded 10-km TT protocol being 2.1% (22). Consequently, the sample size calculation at an α level of 0.05 and power (1- β) of 0.8 revealed that six participants would be sufficient to detect a 2.5% change in 10-km TT performance. However, considering the adequacy of sample sizes in similar studies, as a conservative measure we recruited a final sample size of $n=8$. Data were evaluated using Prism v8.0 (GraphPad Software, Inc., San Diego, CA USA) and are presented as mean [lower, upper 95% confidence interval (CI) of the mean], except for the participant characteristics, which are described as mean \pm SD. A one-way repeated measures analysis of variance (ANOVA) was used to determine whether a trial order effect existed across visits 2, 3 and 4 in the time to complete the 10-km TT. A paired samples t-test was used to determine differences between trials in time to complete the 10-km TT. The SWD was set at 0.2 between-subject SD, which is suggested to represent a practically-relevant change in performance in athletes. Thus, the SWD corresponded to 48 sec, or 2.0%, for 10-km TT performance in this study. Two-way (time x condition) repeated measures ANOVA was used to determine differences between the two experimental trials for all variables with serial measurements. When a main effect of condition, or an interaction effect between condition and time was indicated, *post-hoc* testing was performed with Bonferroni's correction with multiplicity-adjusted *P* values applied to compare CHO+KME to CHO+PLA at the respective time points. The data were tested for normality using Shapiro-Wilk test prior to proceeding with the parametric tests described. For null hypothesis statistical testing, the significance level was set at $\alpha = 0.05$ for all tests.

RESULTS

Plasma β HB, glucose and lactate concentrations. Postprandial plasma concentrations of β HB (mean [95% CI]: CHO+KME, 0.27 [0.22-0.33] mM; CHO+PLA, 0.28 [0.14-0.43] mM),

glucose (CHO+KME, 3.96 [3.22-4.70] mM; CHO+PLA, 3.70 [3.06-4.35] mM), and lactate (CHO+KME, 1.04 [0.79-1.29] mM; CHO+PLA, 1.02 [0.84-1.20] mM) did not differ between trials (all $P > 0.99$). A main effect of time and condition (both $P < 0.001$) and a time-condition interaction effect ($P < 0.001$) were observed for plasma β HB concentrations (Fig. 2A). Ingestion of CHO+KME resulted in a rise in plasma β HB concentrations to 0.99 (0.85-1.14) mM at 0 min. β HB concentrations peaked at 1.33 (1.13-1.52) mM during submaximal exercise at 40 min, with similar concentrations observed at the cessation of the 10-km TT at 1.33 (0.95-1.70) mM.

A main effect of time ($P < 0.001$) and condition ($P = 0.027$) was observed for plasma glucose concentrations (Fig. 2B). Plasma glucose concentrations were lower in CHO+KME at 0 min, i.e. 30 min after ingestion of the first bolus of either CHO+KME or CHO+PLA (CHO+KME, 3.87 [3.22-4.70] mM; CHO+PLA, 4.52 [3.91-5.13] mM; $P = 0.016$) (Fig. 2B). Plasma glucose concentrations rose throughout submaximal exercise (Fig. 2B) with the highest concentrations observed at cessation of the 10-km TT (CHO+KME, 6.94 [5.60-8.28] mM; CHO+PLA 7.24 [5.93-8.54] mM), with no difference between trials ($P > 0.99$).

A main effect of time ($P < 0.001$) was observed for plasma lactate concentrations, but were similar between trials at all time points (Fig. 2C). Peak plasma lactate concentrations were observed at cessation of the 10-km TT (CHO+KME, 6.94 [4.15, 9.73] mM; CHO+PLA, 7.48 [5.46-9.51] mM; $P = 0.738$).

Submaximal exercise. Running speeds were identical between trials as per the study design. There was no difference in % $\text{VO}_{2\text{max}}$, VO_2 , running economy, VCO_2 , RER, HR, and RPE between CHO+KME and CHO+PLA during the submaximal exercise period (Table 1). Main effects of time were observed for the decline in RER ($P < 0.001$), and the increase in RPE ($P < 0.001$) during the submaximal exercise bout (Table 1)

10-km TT performance. No trial order effect was observed for 10-km TT performance between visit 2 (2388 [2187-2588] s), visit 3 (2415 [2223-2607] s), and visit 4 (2409 [2197-2621] s) ($P = 0.742$). There was no statistically significant difference (-20 [-86-45] s; $P = 0.483$) in 10-km TT performance between trials (CHO+KME, 2402 [2204-2600] s; CHO+PLA, 2422 [2217-2628] s) (Fig. 3A). Compared to CHO+PLA, three participants demonstrated improvements in performance with CHO+KME that were greater than the SWD, and one participant demonstrated a decrement in performance with CHO+KME that was greater than the SWD (Fig. 3B). The remaining participants' differences in performance between trials were less than the SWD. Running speeds for each 2 km split during the 10-km TT did not differ between trials, but did increase progressively throughout the TT (main effect of time, $P < 0.001$) (Fig. 3C).

Cognitive performance. In the reaction time test (RTI), main effects of time ($P = 0.026$) and condition ($P = 0.026$) were observed for release time, but no interaction effect was present ($P = 0.535$), whereas an interaction effect was observed for reaction time ($P = 0.014$) (Table 2). In the multi-tasking test (MTT), a main effect of time was observed for response latency ($P = 0.010$), correct responses ($P = 0.049$) and incorrect responses ($P = 0.036$), but no main effects of time, or interaction effects were observed across these parameters (all $P > 0.05$) (Table 2). Overall, there was no difference in cognitive performance between conditions in either the RTI, or MTT assessments (Table 2).

Gastrointestinal symptoms. Out of 8 participants, 4 (50%) reported symptoms of GI distress during CHO+PLA and comprised 4 (50%), 3 (38%), 1 (13%), 1 (13%), and 1 (13%) incidences of belching, flatulence, reflux, urge to defecate, and diarrhea, respectively. Out of 8 participants, 5 (63%) reported symptoms of GI distress during CHO+KME and comprised 3 (38%), 2 (25%), 1 (13%), 1 (13%), 1 (13%), and 1 (13%) incidences of belching, urge to defecate, cramps, reflux, nausea, and stitch, respectively.

Identification of CHO+KME and best performance trials. Out of 8 participants, 2 (25%) correctly identified the trial in which they received CHO+KME, identifying CHO+KME by taste and a perceived alteration of performance. Six (75%) participants declared that they could not differentiate between CHO+PLA and CHO+KME. Seven (88%) participants correctly identified the trial in which they performed their best 10-km TT.

DISCUSSION

The present study investigated whether the acute ingestion of a commercially-available ketone monoester supplement altered metabolic responses, physical and cognitive performance in endurance-trained runners in response to 1 h of submaximal exercise immediately followed by a treadmill-based self-paced 10-km TT. Compared with placebo (CHO+PLA), ingestion of the ketone monoester (CHO+KME) elevated plasma β HB to ~ 1.0 mM at the onset of submaximal exercise, and reached ~ 1.3 mM at the end of the 10-km TT. However, CHO+KME did not alter the metabolic or cardiorespiratory responses to exercise, or demonstrate benefit to physical or cognitive performance compared to CHO+PLA ingestion.

The present study adds to the growing body of literature investigating the effects on exercise performance of elevating ketone body concentrations by exogenous means. The term “exogenous ketone supplement” encompasses a range of different forms of supplements, with each having differential effects on the metabolic response to exercise, and exercise performance. These studies have included the acute ingestion of a (R)-3-hydroxybutyl (R)-3-hydroxybutyrate ketone monoester (KME) (4, 5), and a R,S-1,3-butanediol acetoacetate ketone diester (KDE) (12), racemic ketone salts (KS) (7, 10, 11, 14), and the ketogenic compound 1,3-butanediol (BD) (21, 26) prior to and/or during an exercise challenge. One of the key metabolic consequences of ingesting exogenous ketone supplements is the elevation in circulating β HB, but we speculate that exercise performance is unlikely to be affected unless β HB concentrations exceed 1.0 mM

(27). To date, the only supplement to consistently exceed this threshold prior to an exercise challenge is the KME supplement (4, 5). KS and KDE elevate β HB concentrations into the 0.3 to 0.6 mM range (7, 10, 12), and ingestion of BD elevates β HB concentrations into the 0.6 to 0.8 mM range (21, 26).

Specifically focusing on KME ingestion and exercise studies, ingestion of 573 mg.kg⁻¹ of KME in the fasted state elevated β HB concentrations to ~2.0 mM 20 min after ingestion where it remained throughout 1 h cycling exercise at 75%W_{max} and a subsequent 30 min TT (5). In the fed state, ingestion of 750 mg.kg⁻¹ of KME elevated β HB concentrations to >1.5 mM after 15 min of exercise, and ~2.6 mM by the end of 75 min of intermittent running followed by a short duration shuttle run to exhaustion (4). In contrast to this previous work, plasma β HB concentrations in the present study were elevated to ~1.3 mM during the exercise protocol, which is lower than previously observed at the same 573 mg.kg⁻¹ dose (5). These previous studies have used a split dosing strategy to achieve to the total KME dose described (4, 5), and therefore we employed the same approach. The presently-observed attenuated rise in plasma β HB concentrations compared to these studies is unsurprising given that ingestion of KME in the fasted states consistently elevates circulating β HB to >3.0 mM (8, 9), whereas ingestion of KME in the postprandial state results in circulating β HB in the range from ~1.0 to 2.5 mM (4, 5, 9). For instance, ingestion of 395 mg.kg⁻¹ in the fasted state produces peak β HB concentrations of ~3.0 mM but only ~2.0 mM in the fed state, a 33% reduction in C_{max} and coincides with a 27% reduction in 4 h β HB AUC in resting participants (9). Given that our participants were fed a lower initial dose of KME of 287 mg.kg⁻¹, that this ingestion occurred in a postprandial state, and that exercise commenced 30 min later, it is not surprising that we observed lower β HB concentrations prior to and during exercise compared to previous work (4, 5).

Therefore, although the present protocol achieved acute nutritional ketosis, a benefit to endurance performance was not observed. This finding is consistent with a number of studies that have failed to find a performance benefit of exogenous ketone supplements in various exercise models (4, 10-12, 14). The variety of exogenous ketones supplements used, the large range of changes in circulating β HB produced, and a lack of consistency in the nutrients co-ingested and type of exercise challenge performed, make it difficult to make broad conclusions on the efficacy of these supplements. However, only one study to date has demonstrated a performance benefit with the ingestion of KME, which when co-ingested with CHO increased the distance covered in a 30 min cycling TT by $\sim 2\%$ (mean \pm SEM, 411 \pm 162 m; $n=8$), when preceded by 1 h pre-load exercise at 75% W_{\max} (5). The proposed mechanism for this improvement in performance was a shift in the contribution to energy provision from substrate utilization of carbohydrate to fat, as demonstrated by reduction in glycolytic flux resulting in a 'sparing' of muscle glycogen, and a concomitant increase in intramuscular triglyceride utilization during exercise (5).

The mechanistic basis whereby elevated ketones reduce carbohydrate utilization during exercise is likely an attenuation of glycolytic flux via an inhibition of pyruvate dehydrogenase and phosphofructokinase by increases in NADH:NAD⁺, acetyl-CoA:CoA, or citrate. A reduction in glycolytic flux has been proposed to explain the attenuated exercise-induced rise in plasma lactate observed in previous studies providing KME (4, 5). This attenuation was $\sim 50\%$ during 60 min at 75% W_{\max} and 30 min TT in trained cyclists (5), and $\sim 10\%$ to 30% during 75 min of intermittent running in team sport athletes (4). However, no differences in plasma lactate were observed between trials in the present study either during the pre-load or TT periods. The submaximal exercise intensity of $\sim 65\%VO_{2\max}$ employed was below lactate threshold for all participants, and therefore an intensity too low to observe an attenuation, if any, of the exercise-

induced rise in plasma lactate. However, plasma β HB concentrations were elevated >1.0 mM before and at the cessation of the 10-km TT, yet no difference in plasma lactate was observed between trials.

Similarly, while a glucose-lowering effect of KME ingestion is well-documented whether ingested alone (5, 8, 9), or co-ingested with carbohydrate or protein (4, 5, 9, 13, 15), we observed an attenuation in the rise in plasma glucose concentrations only at 30 min after ingestion of the first bolus of CHO+KME compared to CHO+PLA. This difference in plasma glucose between trials was absent during the submaximal exercise period, and upon completion of the 10-km TT. When effects of KME ingestion on plasma glucose have been observed, the mechanism proposed has been an attenuation of hepatic gluconeogenesis and an increase in hepatic glucose uptake (13). Under certain conditions, elevated KB concentrations may have an insulintropic action (6), but is not always observed (28, 29). When co-ingested with carbohydrate and/or protein, the effect of exogenous ketones to attenuate postprandial glycemia occurs despite similar circulating insulin concentrations between conditions (5, 13, 15).

We propose that the lack of differences between trials for plasma glucose and lactate, in contrast to previous work (4, 5), suggests that the nature of the exercise challenge, or the degree of nutritional ketosis are key determinants of the metabolic effects of exogenous ketone supplements during exercise. While plasma β HB concentrations were elevated to ~ 1.3 mM at the cessation of the 10-km TT, concentrations were ~ 1.2 mM lower than observed in studies demonstrating effects on plasma glucose and lactate during exercise (4, 5). The lower plasma β HB concentrations are a consequence of the aforementioned particulars of the dosing and feeding strategy, and future research should be cognizant of these issues when designing study protocols.

The brain is the primary site of ketone body utilization under conditions of low carbohydrate availability (30). Elevated β HB concentrations are associated with a neuroprotective role in non-exercise contexts (31-33), and short-term (5 days) feeding of a diet supplemented with KME improved performance of rats in a radial maze task by 38%, and improved decision-making during the test (34). Moreover, in our previous work, acute ingestion of KME preserved cognitive performance, measured by the number of incorrect responses to a multi-tasking test (4). This test was performed at the cessation of a short duration intermittent run to exhaustion proceeding the Loughborough Intermittent Shuttle Test (LIST), a variable intensity running protocol that mimics soccer match-play (35). In contrast to previous results, we observed no difference in cognitive performance with the addition of KME in the present study. The specifics of the exercise challenge may play a role in these divergent findings. The LIST is a cognitively-demanding task that requires participants to be aware of current and subsequent running speeds for 75 min. Mental fatigue has a negative impact on aspects of cognitive performance, including altered attentional focus (36), and slower and less accurate reaction times (37), suggesting that the more cognitively-demanding the task, the larger a deficit in cognitive performance should be evident. In the present study, we observed no decline in cognitive performance in either condition. The absence of decline is important to note because in our previous work, it was a preservation of cognitive performance observed with KME, not an absolute improvement (4). These results suggest the exercise challenge presently employed was not sufficiently cognitively-demanding to negatively impact reaction time or executive function, and therefore, potential benefits were unlikely to be observed.

Concerns have been raised about the practical use of exogenous ketone supplements by athletes due to the high rates of occurrence of GI distress in previous work using BD (26), KS (7, 17), KDE (12), and KME (4). However, in the present study, incidences of GI distress were

similar between conditions, and this is consistent with previous work using KME (5). Typically, rates of occurrence of GI distress are higher with exogenous ketones than with ingestion of water or carbohydrate alone, and GI distress occurs at a higher rate with increasing doses of exogenous ketones (4, 7, 38). Importantly, no participants nominated GI distress as a distraction or detriment to performance during CHO+KME trials.

The present study has attempted to incorporate several elements of experimental design that are consistent with reviews of best practice when undertaking studies of nutrition supplements and sports performance (24, 39-41). These include recruitment of trained participants who compete in the chosen mode of exercise, the inclusion of a familiarization trial to improve the reliability of the performance TT, standardization of nutrient intakes before and during each trial, the inclusion of an appropriate placebo coupled to the interrogation of the success of the blinding, and a fuelling strategy that mimics real-world practice i.e. exercise undertaken in fed conditions rather than fasted, and supported by optimal carbohydrate provision during performance. However, the study is not without limitations. While the sample size calculations suggested a small sample would be sufficient to detect meaningful change using this experimental protocol, an *n*-size of eight participants is underpowered to explore relationships between performance differences and inter-individual differences in $\text{VO}_{2\text{max}}$ and peak plasma βHB concentrations achieved. These are two parameters that we speculate are important determinants of the performance benefits, if any, of exogenous ketone supplements (27). For example, that three out of eight participants had an improvement in 10-km TT performance that was greater than the SWD is suggestive of potential benefits to performance in certain athletes. Another limitation, despite the strength of the experimental protocol as described above, is that the performance measure employed lacks ecological validity, and is not an entirely accurate representation of a real-world performance scenario because of the pre-load protocol, and use of

a motorized treadmill with self-paced adjustments of speed. The requirement for participants to manually change the treadmill speed using console buttons is dependent upon their perception of an ability to run faster or slower, but may not be sufficiently sensitive to detect small differences in performance (42, 43).

In conclusion, the addition of a commercially-available ketone monoester supplement to a carbohydrate-based fuelling strategy prior to and during exercise did not improve performance in a self-paced, treadmill-based 10-km TT. Ingestion of the ketone monoester attenuated the rise in plasma glucose prior to exercise, but concentrations were similar between trials thereafter, and no effect on the increase in plasma lactate concentrations during the 10-km TT was observed. Moreover, no differences between trials were observed for a range of physiological responses, and assessments of cognitive performance. Future research should evaluate different dosing strategies and exercise models to elucidate whether a threshold of plasma β HB concentration must be exceeded in order to exert performance benefits, and in which exercise contexts these benefits, if any, might be realized.

ACKNOWLEDGEMENTS

The authors thank Dr. Sabine Harrison (Institute of Food and Health, University College Dublin) and Mr. John Noone (National Institute for Cellular Biotechnology, Dublin City University) for technical assistance during the analysis of blood samples, and Ms. Una Raymond-Hoey and Mr. Eamonn Blake for assistance with data collection. This work was funded in part by an Irish Research Council Enterprise Partnership Postgraduate Fellowship (EPSPG2016177).

The ketone monoester studied in this work was purchased from HVMN, Inc. (San Francisco, USA), and we thank Dr. Brianna Stubbs (HVMN, Inc.) for advice regarding the formulation and flavoring of the placebo condition. HVMN had no role in the implementation, interpretation, or dissemination of this work. No conflict of interest, financial or otherwise, is declared by the authors.

The authors declare the results of the study are presented clearly, honestly, and without fabrication, falsification, or inappropriate data manipulation and do not constitute endorsement by the American College of Sports Medicine.

REFERENCES

1. Egan B, D'Agostino DP. Fueling Performance: Ketones Enter the Mix. *Cell Metab.* 2016;24(3):373-5.
2. Koutnik AP, D'Agostino DP, Egan B. Anticatabolic Effects of Ketone Bodies in Skeletal Muscle. *Trends Endocrinol Metab.* 2019;30(4):227-9.
3. Robinson AM, Williamson DH. Physiological roles of ketone bodies as substrates and signals in mammalian tissues. *Physiol Rev.* 1980;60(1):143-87.
4. Evans M, Egan B. Intermittent Running and Cognitive Performance after Ketone Ester Ingestion. *Med Sci Sports Exerc.* 2018;50(11):2330-8.
5. Cox PJ, Kirk T, Ashmore T et al. Nutritional Ketosis Alters Fuel Preference and Thereby Endurance Performance in Athletes. *Cell Metab.* 2016;24(2):256-68.
6. Balasse EO, Fery F. Ketone body production and disposal: effects of fasting, diabetes, and exercise. *Diabetes Metab Rev.* 1989;5(3):247-70.
7. Evans M, Patchett E, Nally R, Kearns R, Larney M, Egan B. Effect of acute ingestion of beta-hydroxybutyrate salts on the response to graded exercise in trained cyclists. *Eur J Sport Sci.* 2018;18(3):376-86.
8. Stubbs BJ, Cox PJ, Evans RD, Cyranka M, Clarke K, de Wet H. A Ketone Ester Drink Lowers Human Ghrelin and Appetite. *Obesity (Silver Spring).* 2018;26(2):269-73.
9. Stubbs BJ, Cox PJ, Evans RD et al. On the metabolism of exogenous ketones in humans. *Front Physiol.* 2017;8:848.
10. Rodger S, Plews D, Laursen P, Driller MW. Oral β -hydroxybutyrate salt fails to improve 4-minute cycling performance following submaximal exercise. *J Sci Cycling.* 2017;6(1):26-31.

11. O'Malley T, Myette-Cote E, Durrer C, Little JP. Nutritional ketone salts increase fat oxidation but impair high-intensity exercise performance in healthy adult males. *Appl Physiol Nutr Metab*. 2017;42(10):1031-5.
12. Leckey JJ, Ross ML, Quod M, Hawley JA, Burke LM. Ketone Diester Ingestion Impairs Time-Trial Performance in Professional Cyclists. *Front Physiol*. 2017;8:806.
13. Myette-Côté É, Neudorf H, Rafiei H, Clarke K, Little JP. Prior ingestion of exogenous ketone monoester attenuates the glycaemic response to an oral glucose tolerance test in healthy young individuals. *J Physiol*. 2018;596(8):1385-95.
14. Waldman HS, Basham SA, Price FG et al. Exogenous ketone salts do not improve cognitive responses after a high-intensity exercise protocol in healthy college-aged males. *Appl Physiol Nutr Metab*. 2018;43(7):711-7.
15. Vandoorne T, De Smet S, Ramaekers M et al. Intake of a Ketone Ester Drink during Recovery from Exercise Promotes mTORC1 Signaling but Not Glycogen Resynthesis in Human Muscle. *Front Physiol*. 2017;8:310.
16. Holdsworth DA, Cox PJ, Kirk T, Stradling H, Impey SG, Clarke K. A Ketone Ester Drink Increases Postexercise Muscle Glycogen Synthesis in Humans. *Med Sci Sports Exerc*. 2017;49(9):1789-95.
17. Fischer T, Och U, Klawon I et al. Effect of a Sodium and Calcium DL-beta-Hydroxybutyrate Salt in Healthy Adults. *J Nutr Metab*. 2018;2018:9812806.
18. Volek JS, Noakes T, Phinney SD. Rethinking fat as a fuel for endurance exercise. *Eur J Sport Sci*. 2015;15(1):13-20.
19. Hawley JA, Leckey JJ. Carbohydrate Dependence During Prolonged, Intense Endurance Exercise. *Sports Med*. 2015;45 Suppl 1:S5-12.

20. Pinckaers PJ, Churchward-Venne TA, Bailey D, van Loon LJ. Ketone Bodies and Exercise Performance: The Next Magic Bullet or Merely Hype? *Sports Med.* 2017;47(3):383-91.
21. Scott BE, Laursen PB, James LJ et al. The effect of 1,3-butanediol and carbohydrate supplementation on running performance. *J Sci Med Sport.* 2019;22(6):702-6.
22. Russell RD, Redmann SM, Ravussin E, Hunter GR, Larson-Meyer DE. Reproducibility of endurance performance on a treadmill using a preloaded time trial. *Med Sci Sports Exerc.* 2004;36(4):717-24.
23. Barnes KR, Kilding AE. Running economy: measurement, norms, and determining factors. *Sports Med Open.* 2015;1(1):8.
24. Currell K, Jeukendrup AE. Validity, reliability and sensitivity of measures of sporting performance. *Sports Med.* 2008;38(4):297-316.
25. Hopkins WG, Hewson DJ. Variability of competitive performance of distance runners. *Med Sci Sports Exerc.* 2001;33(9):1588-92.
26. Shaw DM, Merien F, Braakhuis A, Plews D, Laursen P, Dulson DK. The Effect of 1,3-Butanediol on Cycling Time-Trial Performance. *Int J Sport Nutr Exerc Metab.* 2019:1-27.
27. Evans M, Cogan KE, Egan B. Metabolism of ketone bodies during exercise and training: physiological basis for exogenous supplementation. *J Physiol.* 2017;595(9):2857-71.
28. Nair KS, Welle SL, Halliday D, Campbell RG. Effect of beta-hydroxybutyrate on whole-body leucine kinetics and fractional mixed skeletal muscle protein synthesis in humans. *J Clin Invest.* 1988;82(1):198-205.
29. Mikkelsen KH, Seifert T, Secher NH, Grondal T, van Hall G. Systemic, cerebral and skeletal muscle ketone body and energy metabolism during acute hyper-D-beta-

- hydroxybutyratemia in post-absorptive healthy males. *J Clin Endocrinol Metab.* 2015;100(2):636-43.
30. Owen OE, Morgan AP, Kemp HG, Sullivan JM, Herrera MG, Cahill GF, Jr. Brain metabolism during fasting. *J Clin Invest.* 1967;46(10):1589-95.
 31. Ari C, Kovacs Z, Juhasz G et al. Exogenous Ketone Supplements Reduce Anxiety-Related Behavior in Sprague-Dawley and Wistar Albino Glaxo/Rijswijk Rats. *Front Mol Neurosci.* 2016;9:137.
 32. Kovacs Z, D'Agostino DP, Dobolyi A, Ari C. Adenosine A1 Receptor Antagonism Abolished the Anti-seizure Effects of Exogenous Ketone Supplementation in Wistar Albino Glaxo Rijswijk Rats. *Front Mol Neurosci.* 2017;10:235.
 33. Svart M, Gormsen LC, Hansen J et al. Regional cerebral effects of ketone body infusion with 3-hydroxybutyrate in humans: Reduced glucose uptake, unchanged oxygen consumption and increased blood flow by positron emission tomography. A randomized, controlled trial. *PLoS One.* 2018;13(2):e0190556.
 34. Murray AJ, Knight NS, Cole MA et al. Novel ketone diet enhances physical and cognitive performance. *FASEB J.* 2016;30(12):4021-32.
 35. Nicholas CW, Nuttall FE, Williams C. The Loughborough Intermittent Shuttle Test: a field test that simulates the activity pattern of soccer. *J Sports Sci.* 2000;18(2):97-104.
 36. Boksem MA, Meijman TF, Lorist MM. Effects of mental fatigue on attention: an ERP study. *Brain Res Cog Brain Res.* 2005;25(1):107-16.
 37. Boksem MA, Meijman TF, Lorist MM. Mental fatigue, motivation and action monitoring. *Biol Psych.* 2006;72(2):123-32.

38. Clarke K, Tchabanenko K, Pawlosky R et al. Kinetics, safety and tolerability of (R)-3-hydroxybutyl (R)-3-hydroxybutyrate in healthy adult subjects. *Regul Toxicol Pharmacol*. 2012;63(3):401-8.
39. Burke LM, Peeling P. Methodologies for Investigating Performance Changes With Supplement Use. *Int J Sport Nutr Exerc Metab*. 2018;28(2):159-69.
40. Jeacocke NA, Burke LM. Methods to standardize dietary intake before performance testing. *Int J Sport Nutr Exerc Metab*. 2010;20(2):87-103.
41. Halperin I, Pyne DB, Martin DT. Threats to internal validity in exercise science: a review of overlooked confounding variables. *Int J Sports Physiol Perform*. 2015;10(7):823-9.
42. Laursen PB, Francis GT, Abbiss CR, Newton MJ, Nosaka K. Reliability of time-to-exhaustion versus time-trial running tests in runners. *Med Sci Sports Exerc*. 2007;39(8):1374-9.
43. Whitham M, McKinney J. Effect of a carbohydrate mouthwash on running time-trial performance. *J Sports Sci*. 2007;25(12):1385-92.

FIGURE LEGENDS

FIGURE 1 – Schematic of the study protocol. CHO, carbohydrate-electrolyte solution; HR, heart rate; KME, ketone monoester; PLA, placebo; RPE, rating of perceived exertion.

FIGURE 2 – Plasma β HB (A), glucose (B), and lactate (C) concentrations during each trial. Data are presented as mean values, with error bars representing 95% confidence intervals. * $P < 0.05$ for CHO+KME vs. CHO+PLA; * $P < 0.001$ for CHO+KME vs. CHO+PLA.**

FIGURE 3 – 10-km time-trial performance (A), individual differences between CHO+KME compared to CHO+PLA (B), and running speeds for each 2 km split during the 10-km time-trials (C). Data in (A) and (C) are presented as mean values, with error bars representing 95% confidence intervals. The shaded area in (B) represents the range for the smallest worthwhile difference in 10-km time-trial performance in this cohort.

Figure 1

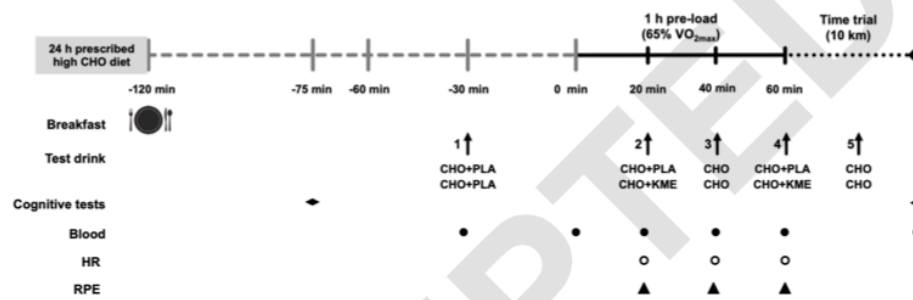


Figure 1

Figure 2

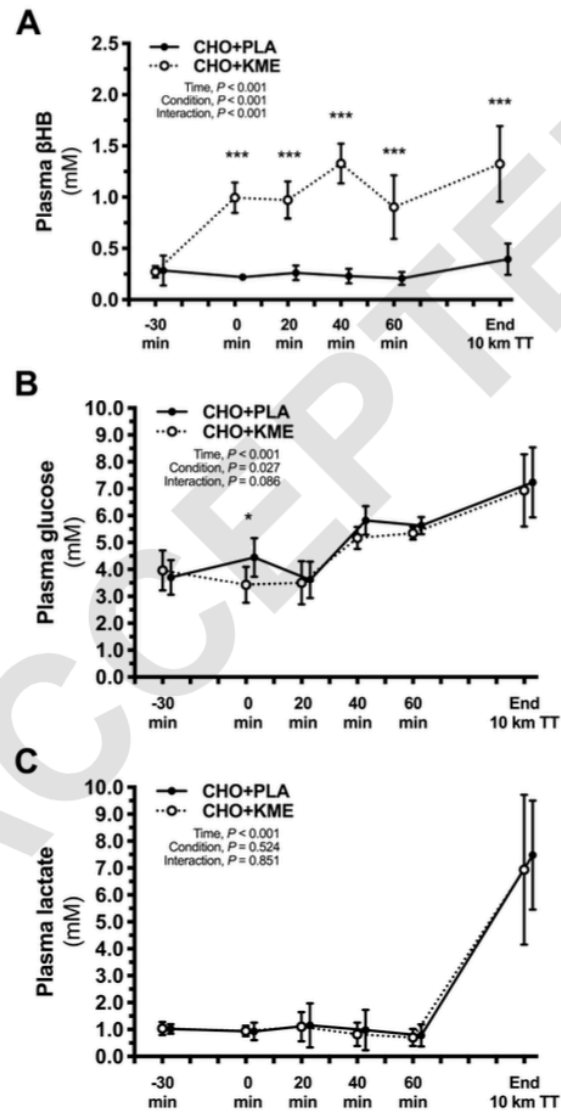


Figure 3

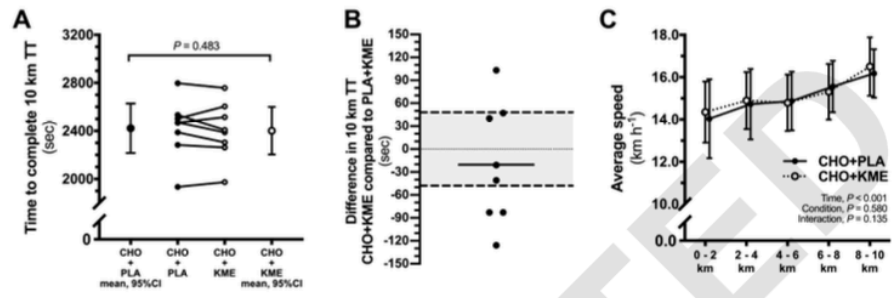


TABLE 1. Physiological responses to 1 h of treadmill running at ~65%VO_{2max} when carbohydrate was co placebo (CHO+PLA) or a ketone monoester (CHO+KME).

	Time			
	0-10 min	10-30 min	30-60 min	
Running speed (km.h ⁻¹)	12.4 (11.3, 13.5)	12.4 (11.3, 13.5)	12.3 (11.1, 13.5)	
VO₂ (L.min ⁻¹)				Time,
CHO+PLA	2.84 (2.52, 3.16)	2.84 (2.56, 3.12)	2.81 (2.53, 3.09)	Condition,
CHO+KME	2.78 (2.42, 3.13)	2.79 (2.42, 3.13)	2.72 (2.49, 2.95)	Interaction,
%VO_{2max}				Time,
CHO+PLA	67.0 (62.8, 71.2)	66.9 (64.5, 69.4)	66.2 (63.8, 69.4)	Condition,
CHO+KME	65.3 (60.9, 69.8)	65.8 (62.6, 69.8)	64.1 (63.2, 65.0)	Interaction,
Running economy (mL.kg ⁻¹ .km ⁻¹)				Time,
CHO+PLA	202 (184, 219)	203 (185, 220)	202 (185, 219)	Condition,
CHO+KME	196 (181, 212)	199 (181, 217)	196 (179, 213)	Interaction,
VCO₂ (L.min ⁻¹)				Time,
CHO+PLA	2.67 (2.36, 2.99)	2.60 (2.30, 2.90)	2.55 (2.26, 2.84)	Condition,
CHO+KME	2.63 (2.28, 2.98)	2.58 (2.28, 2.89)	2.50 (2.26, 2.74)	Interaction,
RER				Time,
CHO+PLA	0.94 (0.92, 0.96)	0.91 (0.89, 0.94)	0.91 (0.88, 0.93)	Condition,

CHO+KME	0.95 (0.92, 0.97)	0.92 (0.89, 0.96)	0.92 (0.89, 0.95)	Interaction,
HR				Time,
(bpm)				Condition,
CHO+PLA	141 (133, 149)	146 (137, 155)	145 (137, 154)	Interaction,
CHO+KME	140 (131, 150)	144 (134, 154)	143 (134, 152)	Time,
RPE				Condition,
CHO+PLA	10 (9, 12)	11 (10, 13)	12 (10, 13)	Interaction,
CHO+KME	10 (8, 12)	11 (9, 12)	11 (9, 13)	

Data are presented as mean (95% CI), n = 8. *** $P < 0.001$.

TABLE 2. Measures of cognitive performance assessed before and after each trial consisting of 1 h of treadmill running at ~65% VO_{2max} followed by a 10-km time-trial during which carbohydrate was co-ingested with either placebo ketone monoester (CHO+KME).

	Reaction time test (RTI)						
	Release time ^{#,§} (msec)			Reaction time [†] (msec)			
	Pre	Post	Post-Pre	Pre	Post	Post-Pre	Pre
CHO+PLA	417 (373, 461)	401 (356, 446)	-16 (-35, 2)	223 (171, 276)	221 (165, 278)	-2 (-18, 14)	0.3 (-0.1, 0.6)
CHO+KME	430 (383, 477)	409 (368, 450)*	-21 (-40, -2)	214 (176, 252)	232 (183, 282)*	18 (2, 34)	0.6 (-0.3, 1.5)

	Multi-tasking test (MTT)						
	Response latency [#] (msec)			Correct responses [#]			
	Pre	Post	Post-Pre	Pre	Post	Post-Pre	Pre
CHO+PLA	599 (500, 698)	561 (447, 674)***	-38 (-58, -18)	159 (157, 160)	157 (155, 159)	-2 (-4, 1)	1 (0, 3)
CHO+KME	583 (513, 653)	541 (461, 622)***	-41 (-62, -21)	158 (157, 160)	157 (156, 159)	-1 (-4, 2)	2 (0, 3)

Data are presented as mean (95%CI), *n*=8. Symbols are [#]*P* < 0.05 for main effect of Time; [§]*P* < 0.05 for main effect of Condition; **P* < 0.05 for Time x Condition interaction effect; [†]*P* < 0.05 for Post vs. Pre; ****P* < 0.001 for Post vs. Pre.