

# Analysis of Water-soluble Vitamins in Biopharma Raw Materials by Electrophoresis Micro-chips with Contactless Conductivity Detection

Fernando Benito-López,<sup>1</sup> Mercedes Vázquez,<sup>1</sup> José L. García-Cordero<sup>2</sup>  
Dermot Diamond<sup>1,2</sup>

<sup>1</sup>Centre for Bioanalytical Sciences, <sup>2</sup>Biomedical Diagnostics Institute, National Centre for Sensor Research, School of Chemical Sciences, Dublin City University, Dublin 9, Ireland

fernando.lopez@dcu.ie, mercedes.vazquez@dcu.ie, jose.garciacordero3@mail.dcu.ie, dermot.diamond@dcu.ie

## INTRODUCTION

Micro-chip-based electrophoresis represents a promising tool for application in the analysis of raw materials in biologics since analysis times can be reduced to seconds and high separation efficiencies can be achieved using extremely low volume samples, minimal reagent consumption and waste generation, low cost/disposability, portability and ease of mass-production [1].

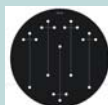
Capacitively Coupled Contactless Conductivity Detection (C<sup>4</sup>D) offers a rather simple and yet sensitive method for detection of ionic species [2]. There is no physical contact of the detection electrodes with the electrolyte solution. Therefore, the integration of this detection mode within the analytical system is rather simple. Furthermore, the background noise is significantly reduced leading to lower detection limits than the conventional contact conductivity detection.

Vitamins are present at very low concentrations in biopharma raw materials and are usually determined using HPLC and CE methods [3]. Electrophoresis micro-chips are a very good alternative to these techniques due to the shorter analysis time and yet very good resolution, among others.

[1] J. West, M. Becker, S. Tombrink, A. Manz, *Anal. Chem.* (2008), 80, 4403-4419.  
[2] P. Kubáň, P. C. Hauser, *Electroanalysis* (2004), 16, 2009.  
[3] M. L. Marszałł, A. Lebedzińska, W. Czarnowski, P. Szefer, *J. Chromatogr. A* (2005), 1094, 91-98.

## MICRO-CHIP FABRICATION

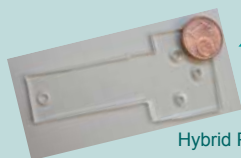
1. MASK FABRICATION: CAD design and high resolution mask printing.



2. CLEAN ROOM: master fabrication.



3. MICRO-CHIP FABRICATION: PDMS (polydimethylsiloxane) layer curing and reversible sealing of PDMS layer to a glass substrate.

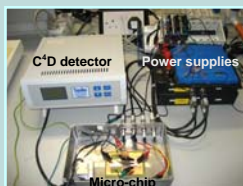


Hybrid PDMS/glass micro-fluidic device

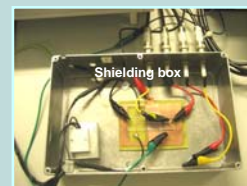
## ACKNOWLEDGMENTS

Irish Research Council for Science, Engineering, and Technology (Fellowship No. 2089), Science Foundation Ireland under Grant No. 05/CE3/B754, IDA (IDA project 116294) and BMS.

## SET-UP



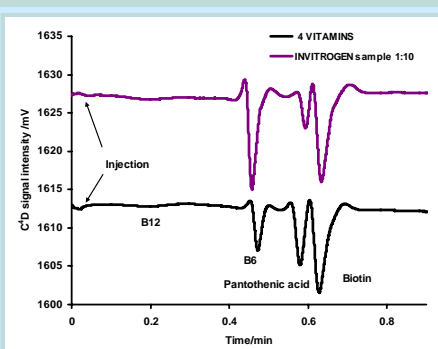
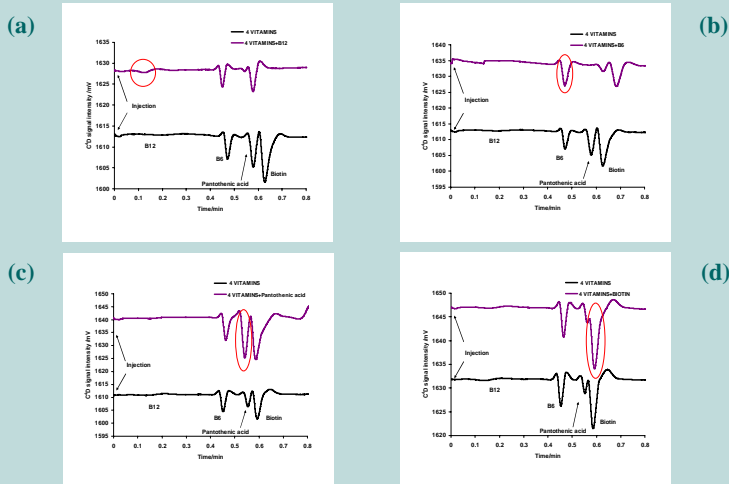
View of the micro-chip shielding box, the C<sup>4</sup>D detector and the power supplies.



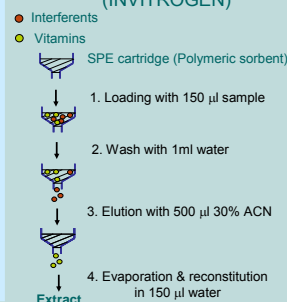
View of the micro-chip setup and C<sup>4</sup>D sensor.

## RESULTS

Electropherograms showing the separation of vitamins in a standard solution of B<sub>12</sub>, B<sub>6</sub>, pantothenic acid and biotin (10 μM each) after spiking B<sub>12</sub> (a), B<sub>6</sub> (b), pantothenic acid (c), or biotin (d). Run buffer: 30 mM Lactic acid (pH 2.6). Micro-chip channels: effective length 3.7 cm, depth 45 μm, width 50 μm. Injection potential: 0.8 kV for 1.5 s. Separation potential: 1.2 kV.



Solid phase extraction (SPE) of a real sample (INVITROGEN)



C<sup>4</sup>D parameters: frequency, 300 kHz; excitation voltage, 20 V<sub>peak-to-peak</sub>